

The infectivity assays of papaya ringspot virus contained the mutation at the dual coat protein cleavage sites

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ABSTRACT

The genome of Papaya ringspot virus (PRSV) contains 10,326 nucleotides that encode a 381 kDa protein processed by three viral proteinase P1, HC-Pro, and NIa. NIa is responsible for the processing of at least six cleavage sites in the C-terminal part of the polyprotein. According to the cleavage rule of NIa, there were two consensus cleavage sequences located between N1b and coat protein (CP), each represented as CP1 and CP2 cut site, and as a results, two in-frame heterologous Nterminal CP would be produced. The double cleavage sites between N1b and CP is only found in PRSV among Potyvirus. The purpose of our study is to characterize the role of the dual cutting sites in PRSV during virus infection. By using the approach of PCR mutagenesis, nine PRSV CP mutants were constructed, eight of them contained the replaced amino acids at CP1 and/or CP2 sites, including HA-GFPCP1QS, HA-GFP-CP1GS, HA-GFP-CP1MS, HA-GFP-CP2ES, HA-GFP-CP2GS, HA-GFP-CP2MS, HA-GFP-CP12MS/GS, and HA-GFP-CP12QS/ES, and the other mutant denoted as HA-GFP-CPdel had 20-amine acid deletion between CP1 and CP2. All constructs were check by restriction enzyme Hind and EcoR digestion and then confirmed by nucleotide sequencing. Because all the constructs of the CP mutants contained a GFP gene, it facilitates the observation of viral replication and movement in host plants. The plasmids of wild type virus and nine PRSV CP mutants were inoculated into systemic host Carica papaya and local lesion host Chenopodium quinoa for observing symptom expression. Papaya plants inoculated by HA-GFP-CP1QS and HA-GFP-CP1GS showed symptoms at 18 d.p.i. and 20 d.p.i., respectively, while the other mutants were unable to caused symptoms. In order to detect the activity of the virus on the inoculated papaya leaves, Western blot analyses using the antiserum against PRSV were further conducted. It was found that only those leaves inoculated by HA-GFP-CP1QS, HA-GFP-CP1GS and HAGFP-CPdel were able to detect the existence of viruses. As compared with papaya -viplants inoculated with wild type viruses, the symptoms showed up about one week slower. When all the 9 PRSV CP mutants were inoculated into a local-lesion host quinoa, all mutants were able to caused local lesions. Our preliminary results suggested that the mutations in CP1 and CP2 cut site did hamper the ability of virus infection in host plants. The role of the larger CP derived from CP1 cutting site may involve in the movement of virus and have less impact on virus replication.

Keywords : Papaya ringspot virus, NIa proteinase, Coat protein, Western blotting

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REFERENCES

- Adams, M.J., Antoniow, J.F. and Beaudoin, F. (2005) Overview and analysis of the polyprotein cleavage sites in the family Potyviridae. *Mol. Plant Pathol.*, 471-487.
- Boevink P. and Oparka, K.J. (2005) Virus-host interactions during movement processes. *Plant Physiol.* 138, 1815-1821
- Carrington, J.C. and Dougherty, W.G. (1988) A viral cleavage site cassette: identification of amino acid sequences required for tobacco etch virus polyprotein processing. *Proc Natl Acad Sci U S A* 85, 3391-5.
- Carrington JC, H.R., Dolja VV, Restrepo-Hartwig MA. (1993) Internal cleavage and trans-proteolytic activities of the VPg-proteinase (NIa) of tobacco etch potyvirus in vivo. *J Virol.* 67, 1-6.
- Chiang, C.H. and Yeh, S.D. (1997) Infectivity assays of in vitro and in vivo transcripts of papaya ringspot potyvirus. *Bot. Bull. Acad. Sin.* 38, 153-163.
- Christie, R.G., and Edwardson, J. R. (1977) Light and electron microscopy of plant virus inclusionS. Dolja, V.V., Haldeman, R., Robertson, N.L., Dougherty, W.G. and Carrington, J.C.(1994) Distinct functions of capsid protein in assembly and movement of tobacco etch potyvirus in plants. *EMBO J* 13(6), 1482-91.
- Dougherty, W.G., Cary, S.M. and Parks, T.D. (1989) Molecular genetic analysis of a plant virus polyprotein cleavage site: a model. *Virology* 171(2), 356-64.
- Edwardson, J.R. (1974) some properties of the potato virus y group. *F1. Agric. Exp. St.Monogr* 4, 225.
- Ghabrial, S.A., Smith, H.A., Parks, T.D. and Dougherty, W.G. (1990) Molecular genetic analyses of the soybean mosaic virus NIa proteinase. *The Journal of general virology* 71(9), 1921-1927.
- Hamilton, R.I., Edwardson, J. R. I. B., (1981) Guidelines for the identification and characterization of plant viruses. *J Gen Virol* 54, 223-241.
- Johansen, E., Edwards, M. C., and Hampton, R. O. (1994) Seed transmission of viruses : current perspectives. *Annu Rev Phytopathol* 32, 363-386.
- Lin S.S., H., R. F., Yeh S.D., (2001) Complete genome sequence and genetic organization of a Taiwan isolate of Zucchini yellow mosaic virus. *Botanical Bulletin of Academia Sinica* 42, 243-250.
- Ozeki, J., Hashimoto, M., Komatsu, K., Maejima, K., Himeno, M., Senshu,

H.,Kawanishi, T., Kagiwada, S., Yamaji, Y. and Namba, S. (2009) The N-terminal region of the *Plantago asiatica* mosaic virus coat protein is required for cell-to-cell movement but is dispensable for virion assembly. *Mol Plant Microbe Interact* 22(6), 677-85. Rao, A.L. and Grantham, G.L. (1996) Molecular studies on bromovirus capsid protein. II. Functional analysis of the amino-terminal arginine-rich motif and its role in encapsidation, movement, and pathology. *Virology* 226(2), 294-305. Riechmann, J.L., Lain, S. and Garcia, J.A. (1990) Infectious in vitro transcripts from a plum pox potyvirus cDNA clone. *Virology* 177(2), 710-6. Riechmann, J.L., Lain, S. and Garcia, J.A. (1992) Highlights and prospects of potyvirus molecular biology. *J Gen Virol* 73 (Pt 1), 1-16. Shukla, D.D. and Ward, C.W. (1989) Identification and classification of potyviruses on the basis of coat protein sequence data and serology. Brief review. *Arch Virol* 106(3-4), 171-200. Siaw, M.F., Shahabuddin, M., Ballard, S., Shaw, J.G. and Rhoads, R.E. (1985) Identification of a protein covalently linked to the 5' terminus of tobacco vein mottling virus RNA. *Virology* 142(1), 134-43. Torrance L, Lukhovitskaya NI, Schepetilnikov MV, Cowan GH, Ziegler A and El., S.(2009) Unusual long-distance movement strategies of Potato mop-top virus RNAs in *Nicotiana benthamiana*. *MPMI*. 22, 1-9. Urcuqui-Inchima, S., Haenni, A.L. and Bernardi, F. (2001) Potyvirus proteins: a wealth of functions. *Virus Res* 74(1-2), 157-75. Varrelmann, M. and Maiss, E. (2000) Mutations in the coat protein gene of plum pox virus suppress particle assembly, heterologous encapsidation and complementation in transgenic plants of *Nicotiana benthamiana*. *J Gen Virol* 81(Pt 3), 567-76. Verchot-Lubicz, J. (2005) A new cell-to-cell transport model for Potexviruses. *Mol Plant Microbe Interact* 18(4), 283-90. Yeh, S.D., Jan, F.J., Chiang, C.H., Doong, T.J., Chen, M.C., Chung, P.H. and Bau, H.J.(1992) Complete nucleotide sequence and genetic organization of papaya ringspot virus RNA. *J Gen Virol* 73 (Pt 10), 2531-41. Zakir Ullah, a, b., Benli Chai , b., Sue Hammar a, Benny Raccab b, b, A.G.-O. and a,R.G. (2003) Effect of substitution of the amino termini of coat proteins of distinct potyvirus species on viral infectivity and host specificity *Physiological and Molecular Plant Pathology* 63, 129-139.