

# Analysis of the cAMP Production in Human 293T Cells Transfected with Zebrafish (*Danio rerio*) Mutated Gonadotropin Receptor

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## ABSTRACT

Biosafety has been a new conflict as the biotechnology making continuous progress and many transgenic and cloned animals have been produced. Sterility is a necessary adjunct to the exploitation of transgenic animals unless completely locked facility is available. The hypothalamus-pituitary-gonads axis and the expression of related genes in the axis play an important role in controlling the reproduction. Follicle-stimulating hormone receptor (FSHR) and luteinizing hormone receptor (LHR) are the members of G protein-coupled receptors, and the intracellular loop 2 and 3 (IL2 and IL3) of these receptors can regulate the G protein activity. In this study, wild type (WT) and point mutated amino acid sequences in conserved regions and intracellular loop 2 regions of zebrafish (*Danio rerio*) FSHR and LHR genes were constructed into reporter vectors and then transfected into human embryonic kidney 293 cells (293T cells). Pregnant mare serum gonadotropin (PMSG) and human chorionic gonadotropin (hCG) were applied to evaluate the cAMP accumulation in the transfected 293T cells, respectively. Dominant negative mutant (DNM) is a competitive effect between wild type and mutant type genes. It may competition with another one. Results of enzyme immunoassay showed that PMSG induced cAMP production only in zFSHR-WT transfected cells, however, cAMP production of zLHR-WT cells could be induced by either PMSG or hCG. The cAMP levels of the 293T cells transfected with zFSHR- or zLHR-DNM were not significantly higher than those of zFSHR- or zLHR-WT cells. After hCG induction, the cAMP levels of zFSHR-DNM cells were significantly higher than those zFSHR-WT, however, contrast results were observed in zLHR-DNM and zLHR-WT cells. Similarly, significantly higher cAMP levels of zFSHR-R448H and zLHR-R474A cells after PMSG induction were observed, indicating negative and positive feedback-like mechanisms, respectively. These DMMs may result in sterile of animals, however, further in vivo experiments should be conducted to verify this hypothesis. The current results may be applied in the fertility control of transgenic or cloned animals and in further biomedical researches.

Keywords : zebrafish, dominant negative mutant, point mutation, G protein-coupled receptors, follicle stimulating hormone receptor, luteinizing hormone receptor, cAMP

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## REFERENCES

- 徐永年。(2006)。斑馬魚發育初期促性腺素受體基因失活之研究。大葉大學分子生物科技學系碩士論文。 Abrahams, M. V. and Sutterlin, A. (1999). The foraging and antipredator behaviour of growth-enhanced transgenic Atlantic salmon. *Anim Behav* 58, 933-942. Angelova, K. and Puett, D. (2002). Differential responses of an invariant region in the ectodomain of three glycoprotein hormone receptors to mutagenesis and assay conditions. *Endocrine* 19, 147-54. Apaja, P. M., Tuusa, J. T., Pietila, E. M., Rajaniemi, H. J. and Petaja-Repo, U. E.(2006). Luteinizing hormone receptor ectodomain splice variant misroutes the full-length receptor into a subcompartment of the endoplasmic reticulum. *Mol Biol Cell* 17, 2243-55. Bluhm, A. P., Toledo, R. A., Mesquita, F. M., Pimenta, M. T., Fernandes, F. M., Ribela, M. T. and Lazari, M. F. (2004). Molecular cloning, sequence analysis and expression of the snake follicle-stimulating hormone receptor. *Gen Comp Endocrinol* 137, 300-11. Burns, F. J., Chen, S., Xu, G., Wu, F. and Tang, M. S. (2002). The action of a dietary retinoid on gene expression and cancer induction in electron-irradiated rat skin. *J Radiat Res (Tokyo)* 43 Suppl, S229-32. Cepni, I., Erkan, S., Ocal, P. and Ozturk, E. (2006). Spontaneous ovarian hyperstimulation syndrome presenting with acute abdomen. *J Postgrad Med* 52,154-5. De Leener, A., Caltabiano, G., Erkan, S., Idil, M., Vassart, G., Pardo, L. and

Costagliola, S. (2008). Identification of the first germline mutation in the extracellular domain of the follitropin receptor responsible for spontaneous ovarian hyperstimulation syndrome. *Hum Mutat* 29, 91-8.

De Leener, A., Montanelli, L., Van Durme, J., Chae, H., Smits, G., Vassart, G. and Costagliola, S. (2006). Presence and absence of follicle-stimulating hormone receptor mutations provide some insights into spontaneous ovarian hyperstimulation syndrome pathophysiology. *J Clin Endocrinol Metab* 91,555-62.

Dierich, A., Sairam, M. R., Monaco, L., Fimia, G. M., Gansmuller, A., LeMeur, M. and Sassone-Corsi, P. (1998). Impairing follicle-stimulating hormone (FSH) signaling in vivo: targeted disruption of the FSH receptor leads to aberrant gametogenesis and hormonal imbalance. *Proc Natl Acad Sci U S A* 95, 13612-7.

Dorsam, R. T. and Gutkind, J. S. (2007). G-protein-coupled receptors and cancer. *Nat Rev Cancer* 7, 79-94.

Dufau, M. L. (1998). The luteinizing hormone receptor. *Annu Rev Physiol* 60,461-96.

Dufau, M. L., Baukal, A. J. and Catt, K. J. (1980). Hormone-induced guanyl nucleotide binding and activation of adenylate cyclase in the Leydig cell. *Proc Natl Acad Sci U S A* 77, 5837-41.

Fredriksson, R., Hoglund, P. J., Gloriam, D. E., Lagerstrom, M. C. and Schiöth, H. B. (2003). Seven evolutionarily conserved human rhodopsin G protein-coupled receptors lacking close relatives. *FEBS Lett* 554, 381-8.

Heckert, L. L. and Griswold, M. D. (2002). The expression of the follicle-stimulating hormone receptor in spermatogenesis. *Recent Prog Horm Res* 57, 129-48.

Hillier, S. G. (2001). Gonadotropic control of ovarian follicular growth and development. *Mol Cell Endocrinol* 179, 39-46.

Huhtaniemi, I. (2006). Mutations along the pituitary-gonadal axis affecting sexual maturation: novel information from transgenic and knockout mice. *Mol Cell Endocrinol* 254-255, 84-90.

Kawate, N. and Menon, K. M. (1994). Palmitoylation of luteinizing hormone/human chorionic gonadotropin receptors in transfected cells. Abolition of palmitoylation by mutation of Cys-621 and Cys-622 residues in the cytoplasmic tail increases ligand-induced internalization of the receptor. *J Biol Chem* 269,30651-8.

Klabunde, T. and Hessler, G. (2002). Drug design strategies for targeting G-protein-coupled receptors. *Chembiochem* 3, 928-44.

Kumar, R. S., Ijiri, S. and Trant, J. M. (2001). Molecular biology of channel catfish gonadotropin receptors: 1. Cloning of a functional luteinizing hormone receptor and preovulatory induction of gene expression. *Biol Reprod* 64,1010-8.

Kumar, T. R., Palapattu, G., Wang, P., Woodruff, T. K., Boime, I., Byrne, M. C. and Matzuk, M. M. (1999). Transgenic models to study gonadotropin function: the role of follicle-stimulating hormone in gonadal growth and tumorigenesis. *Mol Endocrinol* 13, 851-65.

Kwok, H. F., So, W. K., Wang, Y. and Ge, W. (2005). Zebrafish gonadotropins and their receptors: I. Cloning and characterization of zebrafish follicle-stimulating hormone and luteinizing hormone receptors--evidence for their distinct functions in follicle development. *Biol Reprod* 72, 1370-81.

Laan, M., Richmond, H., He, C. and Campbell, R. K. (2002). Zebrafish as a model for vertebrate reproduction: characterization of the first functional zebrafish (*Danio rerio*) gonadotropin receptor. *Gen Comp Endocrinol* 125,349-64.

Latronico, A. C., Chai, Y., Arnhold, I. J., Liu, X., Mendonca, B. B. and Segaloff, D. L. (1998). A homozygous microdeletion in helix 7 of the luteinizing hormone receptor associated with familial testicular and ovarian resistance is due to both decreased cell surface expression and impaired effector activation by the cell surface receptor. *Mol Endocrinol* 12, 442-50.

Latronico, A. C. and Segaloff, D. L. (2007). Insights learned from L457(3.43)R, an activating mutant of the human lutropin receptor. *Mol Cell Endocrinol* 260-262,287-93.

Lei, H. Q. and Xu, Y. X. (1993). Effects of gossypol acetic acid on the motility of rat oviduct smooth muscle. *Yao Xue Xue Bao* 28, 6-10.

Lei, Z. M., Mishra, S., Zou, W., Xu, B., Foltz, M., Li, X. and Rao, C. V. (2001). Targeted disruption of luteinizing hormone/human chorionic gonadotropin receptor gene. *Mol Endocrinol* 15, 184-200.

Logan, K. A., Juengel, J. L. and McNatty, K. P. (2002). Onset of steroidogenic enzyme gene expression during ovarian follicular development in sheep. *Biol Reprod* 66, 906-16.

Lunenfeld, B. and Insler, V. (1993). Follicular development and its control. *Gynecol Endocrinol* 7, 285-91.

Mertani, H. C., Pechoux, C., Garcia-Caballero, T., Waters, M. J. and Morel, G. (1995). Cellular localization of the growth hormone receptor/binding protein in the human anterior pituitary gland. *J Clin Endocrinol Metab* 80, 3361-7.

Miwa, S., Yan, L. and Swanson, P. (1994). Localization of two gonadotropin receptors in the salmon gonad by in vitro ligand autoradiography. *Biol Reprod* 50, 629-42.

Munshi, U. M., Clouser, C. L., Peegel, H. and Menon, K. M. (2005). Evidence that palmitoylation of carboxyl terminus cysteine residues of the human luteinizing hormone receptor regulates postendocytic processing. *Mol Endocrinol* 19, 749-58.

Nakayama, Y., Yamamoto, T., Oba, Y., Nagahama, Y. and Abe, S. (2000). Molecular cloning, functional characterization, and gene expression of a follicle-stimulating hormone receptor in the testis of newt *Cynops pyrrhogaster*. *Biochem Biophys Res Commun* 275, 121-8.

Oba, Y., Hirai, T., Yoshiura, Y., Yoshikuni, M., Kawachi, H. and Nagahama, Y. (1999). Cloning, functional characterization, and expression of a gonadotropin receptor cDNA in the ovary and testis of amago salmon (*Oncorhynchus rhodurus*). *Biochem Biophys Res Commun* 263, 584-90.

Schubert, R. L. and Puett, D. (2003). Single-chain human chorionic gonadotropin analogs containing the determinant loop of the beta-subunit linked to the alpha-subunit. *J Mol Endocrinol* 31, 157-68.

So, W. K., Kwok, H. F. and Ge, W. (2005). Zebrafish gonadotropins and their receptors: II. Cloning and characterization of zebrafish follicle-stimulating hormone and luteinizing hormone subunits--their spatial-temporal expression patterns and receptor specificity. *Biol Reprod* 72, 1382-96.

Takeda, J. and Inada, E. (2002). Next generation of drugs used in anesthesia. *Masui* 51 Suppl, S183-8.

Themmen, A. P. N. and Huhtaniemi, I. T. (2000). Mutations of gonadotropins and gonadotropin receptors: elucidating the physiology and pathophysiology of pituitary-gonadal function. *Endocr Rev* 21, 551-83.

Timossi, C., Maldonado, D., Vizcaino, A., Lindau-Shepard, B., Conn, P. M. and Ulloa-Aguirre, A. (2002). Structural determinants in the second intracellular loop of the human follicle-stimulating hormone receptor are involved in G(s)protein activation. In *Mol Cell Endocrinol* 189, 157-68.

Ulloa-Aguirre, A., Maldonado, A., Damian-Matsumura, P. and Timossi, C. (2001). Endocrine regulation of gonadotropin glycosylation. *Arch Med Res* 32,520-32.

Ulloa-Aguirre, A., Uribe, A., Zarinan, T., Bustos-Jaimes, I., Perez-Solis, M. A. and Dias, J. A. (2007). Role of the intracellular domains of the human FSH receptor in G(alphaS) protein coupling and receptor expression. *Mol Cell Endocrinol* 260-262, 153-62.

Uribe, A., Zarinan, T., Perez-Solis, M. A., Gutierrez-Sagal, R., Jardon-Valadez, E., Pineiro, A., Dias, J. A. and Ulloa-Aguirre, A. (2008). Functional and structural roles of conserved cysteine residues in the carboxyl-terminal domain of the follicle-stimulating hormone receptor in human embryonic kidney 293 cells. *Biol Reprod* 78,

869-82. Vassart, G., Pardo, L. and Costagliola, S. (2004). A molecular dissection of the glycoprotein hormone receptors. *Trends Biochem Sci* 29, 119-26. Wang, Y. and Ge, W. (2003). Gonadotropin regulation of follistatin expression in the cultured ovarian follicle cells of zebrafish, *Danio rerio*. *Gen Comp Endocrinol* 134, 308-15. Weiss, J., Axelrod, L., Whitcomb, R. W., Harris, P. E., Crowley, W. F. and Jameson, J. L. (1992). Hypogonadism caused by a single amino acid substitution in the beta subunit of luteinizing hormone. *N Engl J Med* 326,179-83. Wood, A. J. (2002). Variability in beta-adrenergic receptor response in the vasculature: Role of receptor polymorphism. *J Allergy Clin Immunol* 110,S318-21. Yamashita, S., Nakamura, K., Omori, Y., Tsunekawa, K., Murakami, M. and Minegishi, T. (2005). Association of human follitropin (FSH) receptor with splicing variant of human lutropin/choriogonadotropin receptor negatively controls the expression of human FSH receptor. *Mol Endocrinol* 19, 2099-111. Yan, L., Swanson, P. and Dickhoff, W. W. (1992). A two-receptor model for salmon gonadotropins (GTH I and GTH II). *Biol Reprod* 47, 418-27. Zhang, F. P., Kero, J. and Huhtaniemi, I. (1998). The unique exon 10 of the human luteinizing hormone receptor is necessary for expression of the receptor protein at the plasma membrane in the human luteinizing hormone receptor, but deleterious when inserted into the human follicle-stimulating hormone receptor. *Mol Cell Endocrinol* 142, 165-74. Zhang, F. P., Poutanen, M., Wilbertz, J. and Huhtaniemi, I. (2001). Normal prenatal but arrested postnatal sexual development of luteinizing hormone receptor knockout (LuRKO) mice. *Mol Endocrinol* 15, 172-83.