

# Expression of a Human Extracellular Superoxide Dismutase Gene by Turnip Mosaic Viral Vector

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## ABSTRACT

Active oxygen species (AOS), such as superoxide(O<sub>2</sub><sup>-</sup>) are generated as by-products of normal metabolism in aerobic organisms. Superoxide dismutase (SOD) is the first line of defense to eliminate superoxide in the cellular environment. Extracellular SOD (EC-SOD), an isoform of SOD, is predominantly located in blood and extracellular matrix of tissue, and play a role to scavenge O<sub>2</sub><sup>-</sup> generated in extracellular space. Because SOD is a highly economic value protein, the objective of our study is to use Turnip mosaic virus (TuMV) viral vector to express human ec-sod gene in plant system. Four TuMV-ECSOD recombinant viruses were obtained by replacing the NSs gene in TuMV-NSs with ec-sod constructs. There were four different ec-sod constructs with or without a signal peptide at the N-terminus, and with or without an endoplasmic reticulum (ER) retention signal KDEL at the carboxyl-terminus. The resulting recombinant virus TuMV-ECSOD-S expressed the EC-SOD protein with a secreted N-terminal extracellular (EC) signal peptide. Recombinant TuMV-ECSOD-SK contained the EC-SOD with a secreted N-terminal signal peptide and a KDEL signal at C-terminus. Recombinant TuMV-ECSOD-N contained none of the signal peptide at both ends and recombinant TuMV-ECSOD-K had the EC-SOD protein with a KDEL at C-terminus. The four TuMV-ECSOD recombinant viruses were introduced into plants of *Chenopodium quinoa* Willd. All plants developed local lesion symptoms at 5 to 7 days post inoculation, except for the plants inoculated by TuMV-ECSOD-SK shown one day delay of symptom expression. The size of the local lesions on plants of quinoa infected by TuMV-ECSOD were similar to the plants inoculated by TuMV-NSs. Local lesions were collected and then inoculated into systemic hosts *Brassica chinensis* L. and *Brassica chinensis* L. CV. At approximately 10 to 21 days, the recombinants TuMV-ECSOD-N and TuMV-ECSOD-K inoculated plants showed mosaic, with some necrosis and hard, crisp systematic symptom and finally the entire plant dwarf, while the plants inoculated with the local lesions collected from TuMV-ECSOD-S and TuMV-ECSOD-Sk infected *Chenopodium quinoa* plants showed no symptom. Western blotting analyses with the antisera against EC-SOD and TuMV, respectively, were able to detect either the EC-SOD protein with the molecular weight of 24 kDa or TuMV coat protein with the molecular weight of 35 kDa. The results showed that TuMV-ECSOD-N and TuMV-ECSOD-K were infectious and were able to express EC-SOD protein correctly. In SOD activity assays with NBT, Riboflavin, KCN and H<sub>2</sub>O<sub>2</sub>, the protein of CuZn-SOD extracted from plants inoculated with TuMV-ECSOD showed higher expression than those plants inoculated with TuMV-NSs or healthy plants. The results suggest that the expression of CuZn-SOD in plants can be stimulated by the inoculation of recombinant TuMV-ECSOD. A very faint extra SOD protein under the bands of CuZn-SOD was observed from the extracts of TuMV-ECSOD infected plant was considered as the EC-SOD protein. In addition, the TuMV-ECSOD infected plants showed better resistance to herbicide paraquat.

Keywords : Extracellular SOD (EC-SOD) ; Turnip mosaic virus (TuMV) ; *Chenopodium quinoa* Willd. ; *Brassica chinensis* L. ; *Brassica chinensis* L. CV. ; Paraquat

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## REFERENCES

- 王妙玲、陳小玲、陳全木 2005 利用基因工程乳酸菌產製外源性胞外超氧歧化?。大葉大學分子生物科技學系碩士學位論文。彰化，台灣。
- Adachi, T., Morihara, N., Yamazaki, N., Yamada, H., Futenma, A., Kato, K., and Hirano, K. (1996). An arginine-213 to glycine mutation in human extracellular superoxide dismutase reduces susceptibility to trypsin-like proteinases. *J. Biochem.* 120, 184-188.
- Alscher, R. G., Erturk, N., and Heath, L. S. (2002). Role of superoxide dismutases (SODs) in controlling oxidative stress in plants. *J. Exp. Bot.* 53, 1331-1341.
- Arakawa, T., Yu, J., and Langridge, W. (2001). Synthesis of a cholera toxin B subunit-rotavirus NSP4 fusion protein in potato. *Plant Cell Reports* 20, 343-348.
- Asada, K. (1994). Production and action of active oxygen species in photosynthetic tissue. In: Foyer CH, Mullineaux PM, eds. *Causes of photooxidative stress and amelioration of defense systems in plants*. Boca Raton CRC Press, 77-104.
- Baldensperger, J. B. (1978). An iron containing superoxide dismutase from the chemo lithotrophic *Thiobacillus denitrificans* rt strain. *Arch. Microbiol.* 119, 237-444.
- Barra, D., Schinina, M. E., Bossa, F., Puget, K., Durosay, P., Guissani, A., and Michelson, A. M. (1990). A tetrameric iron superoxide dismutase from the eucaryote *Tetrahymena pyriformis*. *J. Biol. Chem.* 265, 17680-17687.
- Beevers, G., Lip, G. Y., and O'Brien, E. (2001). ABC of hypertension: The pathophysiology of hypertension. *B. m. j.* 322, 912-916.
- Benedetto, M. T., Anzai, Y., and Gordon, J. W. (1991). Isolation and analysis of the mouse genomic sequence encoding Cu(2+)-Zn(2+) superoxide dismutase. *Gene* 99, 191-195.
- Bowler, C., Slooten, L., Vandenbranden, S., De Rycke, R., Botterman, J., Sybesma, C., Van Montagu, M., and Inze, D. (1991). Manganese superoxide dismutase can reduce cellular damage mediated by oxygen radicals in transgenic plants. *EMBO* 10, 1723-1732.
- Bowler, C., Van Camp, W., Van Montagu, M., and Inze, D. (1994). Superoxide dismutases in plants. *Crit. Rev. Plant Sci.* 13, 199-218.
- Butterfield, D. A., Hensley, K., Harris, M., Mattson, M., and Carney, J. (1994). beta-Amyloid peptide free radical fragments initiate synaptosomal lipoperoxidation in a sequence-specific fashion: implications to Alzheimer's disease. *Biochem. Biophys. Res. Commun.* 200, 710-715.
- Cardinale, A., Filesi, I., Vetrugno, V., Pocchiari, M., Sy, M. S., and Biocca, S. (2005). Trapping prion protein in the endoplasmic reticulum impairs PrPC maturation and prevents PrPSc accumulation. *J. Biol. Chem.* 280, 685-694.
- Carlsson, L. M., Jonsson, J., Edlund, T., and Marklund, S. L. (1995). Mice lacking extracellular superoxide dismutase are more sensitive to hyperoxia. *Proc. Natl. Acad. Sci. USA* 92, 6264-6268.
- Chan, P. H., Yang, G. Y., Chen, S. F., Carlson, E., and Epstein, C. J. (1991). Cold-induced brain edema and infarction are reduced in transgenic mice overexpressing CuZn-superoxide dismutase. *Ann. Neuro.* 29, 482-486.
- Chen, C. C. (2006). Development of turnip mosaic virus as a plant viral vector for expressing foreign proteins and generation of attenuated strains for cross protection. . Ph.D. Thesis, Department of Plant Pathology, National Chung Hsing University, Taiwan, ROC.
- Chen, Y., Hou, M., Li, Y., Traverse, J. H., Zhang, P., Salvemini, D., Fukai, T., and Bache, R. J. (2005). Increased superoxide production causes coronary endothelial dysfunction and depressed oxygen consumption in the failing heart. *Am. J. Physiol. Heart. Circ. Physiol.* 288, H133-H141.
-

Davuluri, G. R., van Tuinen, A., Mustilli, A. C., Manfredonia, A., Newman, R., Burgess, D., Brummell, D. A., King, S. R., Palys, J., Uhlig, J., Pennings, H. M., and Bowler, C. (2004). Manipulation of DET1 expression in tomato results in photomorphogenic phenotypes caused by post-transcriptional gene silencing. *Plant J.* 40, 344-354.

19. Edlund, A., Edlund, T., Hjalmarsson, K., Marklund, S. L., Sandstrom, J., Stromqvist, M., and Tibell, L. (1992). A non-glycosylated extracellular superoxide dismutase variant. *Biochem. J.* 288, 451-456.

20. Enghild, J. J., Thogersen, I. B., Oury, T. D., Valnickova, Z., Hojrup, P., and Crapo, J. D. (1999). The heparin-binding domain of extracellular superoxide dismutase is proteolytically processed intracellularly during biosynthesis. *J. Biol. Chem.* 274, 14818-14822.

21. Esterbauer, H., Gebicki, J., Puhl, H., and Jurgens, G. (1992). The role of lipid peroxidation and antioxidants in oxidative modification of LDL. *Free Radic. Biol. Med.* 13, 341-90.

22. Fahn, S., and Cohen, G. (1992). The oxidant stress hypothesis in Parkinson's disease: evidence supporting it. *Ann Neurol* 32, 804-812.

23. Fattman, C. L., Enghild, J. J., Crapo, J. D., Schaefer, L. M., Valnickova, Z., and Oury, T. D. (2000). Purification and characterization of extracellular superoxide dismutase in mouse lung. *Biochem. Biophys. Res. Commun.* 275, 542-548.

24. Fattman, C. L., Schaefer, L. M., and Oury, T. D. (2003). Extracellular superoxide dismutase in biology and medicine. *Free Radic. Biol. Med.* 35, 236-256.

25. Folz, R. J., Abushamaa, A. M., and Suliman, H. B. (1999). Extracellular superoxide dismutase in the airways of transgenic mice reduces inflammation and attenuates lung toxicity following hyperoxia. *J. Clin. Invest.* 103, 1055-1066.

26. Folz, R. J., and Crapo, J. D. (1994). Extracellular superoxide dismutase (SOD3): tissue-specific expression, genomic characterization, and computer-assisted sequence analysis of the human EC SOD gene. *Genomics* 22, 162-171.

27. Folz, R. J., Guan, J., Seldin, M. F., Oury, T. D., Enghild, J. J., and Crapo, J. D. (1997). Mouse extracellular superoxide dismutase: primary structure, tissue-specific gene expression, chromosomal localization, and lung in situ hybridization. *Am. J. Respir. Cell Mol Biol* 17, 393-403.

28. Forman, H. J., and Fridovich, I. (1973). On the stability of bovine superoxide dismutase. The effects of metals. *J. Biol. Chem.* 248, 2645-2649.

29. Foyer, C. H., Descourvieres, P., and Kunert, K. J. (1994). Protection against oxygen radicals: an important defense mechanism studied in transgenic plants. *Plant Cell and Environment* 17, 507-523.

30. Foyer, C. H., and Halliwell, B. (1976). The presence of glutathione and glutathione reductase in chloroplasts: a proposed role in ascorbic acid metabolism. *Planta* 133, 21-25.

31. Fridovich, I. (1986). Superoxide dismutases. *Adv. Enzymol. Relat. Areas Mol. Biol.* 58, 61-97.

32. Fridovich, I. (1995). Superoxide radical and superoxide dismutases. *Annu. Rev. Biochem.* 64, 97-112.

33. Frigerio, L., Pastres, A., Prada, A., and Vitale, A. (2001). Influence of KDEL on the fate of trimeric or assembly-defective phaseolin: selective use of an alternative route to vacuoles. *Plant Cell* 13, 1109-1126.

34. Fukai, T., Folz, R. J., Landmesser, U., and Harrison, D. G. (2002). Extracellular superoxide dismutase and cardiovascular disease. *Cardiovasc. Res.* 55, 239-49.

35. Fukai, T., Galis, Z. S., Meng, X. P., Parthasarathy, S., and Harrison, D. G. (1998). Vascular expression of extracellular superoxide dismutase in atherosclerosis. *J. Clin. Invest.* 101, 2101-11.

36. Fukai, T., Siegfried, M. R., Ushio-Fukai, M., Cheng, Y., Kojda, G., and Harrison, D. G. (2000). Regulation of the vascular extracellular superoxide dismutase by nitric oxide and exercise training. *J. Clin. Invest.* 105, 1631-1639.

37. Fukai, T., Siegfried, M. R., Ushio-Fukai, M., Griendling, K. K., and Harrison, D. G. (1999). Modulation of extracellular superoxide dismutase expression by angiotensin II and hypertension. *Circ. Res.* 85, 23-28.

38. Gao, F., Kinnula, V. L., Myllarniemi, M., and Oury, T. D. (2008). Extracellular superoxide dismutase in pulmonary fibrosis. *Antioxid Redox Signal* 10, 343-354.

39. Gao, Y., Ma, Y., Li, M., Cheng, T., Li, S. W., Zhang, J., and Xia, N. S. (2003). Oral immunization of animals with transgenic cherry tomatillo expressing HBsAg. *World J Gastroenterol* 9, 996-1002.

40. Gauldie, J., Jordana, M., and Cox, G. (1993). Cytokines and pulmonary fibrosis. *Thorax* 48, 931-935.

41. Gerlach, D., Reichardt, W., and Vettermann, S. (1998). Extracellular superoxide dismutase from *Streptococcus pyogenes* type 12 strain is manganese-dependent. *FEMS Microbiol. Lett.* 160, 217-224.

42. Giddings, G. (2001). Transgenic plants as protein factories. *Curr. Opin. Biotechnol.* 12, 450-454.

43. Gil, F., Brun, A., Wigdorovitz, A., Catala, R., Martinez-Torrecedrera, J. L., Casal, I., Salinas, J., Borca, M. V., and Escribano, J. M. (2001). High-yield expression of a viral peptide vaccine in transgenic plants. *FEBS Lett.* 488, 13-17.

44. Giordano, F. J. (2005). Oxygen, oxidative stress, hypoxia, and heart failure. *J. Clin. Invest.* 115, 500-508.

45. Gomez, J. M., Jimenez, A., Olmos, E., and Sevilla, F. (2004). Location and effects of long-term NaCl stress on superoxide dismutase and ascorbate peroxidase isoenzymes of pea (*Pisum sativum* cv. Puget) chloroplasts. *J. Exp. Bot.* 55, 119-130.

46. Halliwell, B. (1989). Free radicals, reactive oxygen species and human disease: a critical evaluation with special reference to atherosclerosis. *Br. J. Exp. Pathol.* 70, 737-757.

47. Halliwell, B. (1993). Free radicals and vascular disease: how much do we know? *B. m. j.* 307, 885-886.

48. Halliwell, B., Gutteridge, J. M., and Cross, C. E. (1992). Free radicals, antioxidants, and human disease: where are we now? *J. Lab. Clin. Med.* 119, 598-620.

49. Hetherington, P. R., McKersie, B. D., and Borochoy, A. (1987). Ice Encasement Injury to Microsomal Membranes from Winter Wheat Crowns : I. Comparison of Membrane Properties after Lethal Ice Encasement and during a Post-Thaw Period. *Plant Physiol* 85, 1068-1072.

50. Hjalmarsson, K., Marklund, S. L., Engstrom, A., and Edlund, T. (1987). Isolation and sequence of complementary DNA encoding human extracellular superoxide dismutase. *Proc. Natl. Acad. Sci. USA* 84, 6340-6344.

51. Hood, E. E., Woodard, S. L., and Horn, M. E. (2002). Monoclonal antibody manufacturing in transgenic plants--myths and realities. *Curr. Opin. Biotechnol.* 13, 630-635.

52. Hsu, C. H., Lin, S. S., Liu, F. L., Su, W. C., and Yeh, S. D. (2004). Oral administration of a mite allergen expressed by zucchini yellow mosaic virus in cucurbit species downregulates allergen-induced airway inflammation and IgE synthesis. *J. Allergy. Clin. Immunol.* 113, 1079-85.

53. Juul, K., Tybjaerg-Hansen, A., Marklund, S., Heegaard, N. H., Steffensen, R., Sillesen, H., Jensen, G., and Nordestgaard, B. G. (2004). Genetically reduced antioxidative protection and increased ischemic heart disease risk: The Copenhagen City Heart Study. *Circulation* 109, 59-65.

54. Kanematsu, S., and Asada, K. (1978). Superoxide dismutase from an anaerobic photosynthetic bacterium, *Chromatium vinosum*. *Arch. Biochem. Biophys.* 185, 473-482.

55. Kanematsu, S., and Asada, K. (1991). Chloroplast and cytosol isozymes of CuZn-superoxide dismutase: their characteristic amino acid sequences. *Free Radic. Res. Commun.* 12-13 Pt 1, 383-390.

56. Karlsson, K., and Marklund, S. L. (1987). Heparin-induced release of extracellular superoxide dismutase to human blood plasma. *Biochem. J.* 242, 55-59.

57.

Karlsson, K., and Marklund, S. L. (1988). Extracellular superoxide dismutase in the vascular system of mammals. *Biochem J* 255, 223-228. 58.

Karlsson, K., and Marklund, S. L. (1989). Binding of human extracellular-superoxide dismutase C to cultured cell lines and to blood cells. *Lab. Invest.* 60, 659-666. 59.

Kim, Y. H., Yoo, H. Y., Jung, G., Kim, J. Y., and Rho, H. M. (1993). Isolation and analysis of the rat genomic sequence encoding Cu/Zn superoxide dismutase. *Gene* 133, 267-271. 60.

Kinouchi, H., Epstein, C. J., Mizui, T., Carlson, E., Chen, S. F., and Chan, P. H. (1991). Attenuation of focal cerebral ischemic injury in transgenic mice overexpressing CuZn superoxide dismutase. *Proc. Natl. Acad. Sci. USA* 88, 11158-11162. 61.

Kirby, T. W., Lancaster, J. R., Jr., and Fridovich, I. (1981). Isolation and characterization of the iron-containing superoxide dismutase of *Methanobacterium bryantii*. *Arch. Biochem. Biophys.* 210, 140-148. 62.

Kojda, G., and Harrison, D. (1999). Interactions between NO and reactive oxygen species: pathophysiological importance in atherosclerosis, hypertension, diabetes and heart failure. *Cardiovasc. Res.* 43, 562-571. 63.

Kong, Q., Richter, L., Yang, Y. F., Arntzen, C. J., Mason, H. S., and Thanavala, Y. (2001). Oral immunization with hepatitis B surface antigen expressed in transgenic plants. *Proc. Natl Acad. Sci. USA* 98, 11539-11544. 64.

Ku, H. H., Brunk, U. T., and Sohal, R. S. (1993). Relationship between mitochondrial superoxide and hydrogen peroxide production and longevity of mammalian species. *Free Radic. Biol. Med.* 15, 621-627. 65.

Kusunose, E., Ichihara, K., Noda, Y., and Kusunose, M. (1976). Superoxide dismutase from *Mycobacterium tuberculosis*. *J. Biochem.* 80, 1343-1352. 66.

Landmesser, U., Merten, R., Spiekermann, S., Buttner, K., Drexler, H., and Hornig, B. (2000). Vascular extracellular superoxide dismutase activity in patients with coronary artery disease: relation to endothelium-dependent vasodilation. *Circulation* 101, 2264-2270. 67.

Landmesser, U., Spiekermann, S., Dikalov, S., Tatge, H., Wilke, R., Kohler, C., Harrison, D. G., Hornig, B., and Drexler, H. (2002). Vascular oxidative stress and endothelial dysfunction in patients with chronic heart failure: role of xanthine-oxidase and extracellular superoxide dismutase. *Circulation* 106, 3073-3078. 68.

Lee, K. W., Meyer, N., and Ortwerth, B. J. (1999). Chromatographic comparison of the UVA sensitizers present in brunescant cataracts and in calf lens proteins ascorbylated in vitro. *Exp. Eye. Res.* 69, 375-384. 69.

Levanon, D., Lieman-Hurwitz, J., Dafni, N., Wigderson, M., Sherman, L., Bernstein, Y., Laver-Rudich, Z., Danciger, E., Stein, O., and Groner, Y. (1985). Architecture and anatomy of the chromosomal locus in human chromosome 21 encoding the Cu/Zn superoxide dismutase. *EMBO J* 4, 77-84. 70.

Lookene, A., Stenlund, P., and Tibell, L. A. (2000). Characterization of heparin binding of human extracellular superoxide dismutase. *Biochemistry* 39, 230-236. 71.

Lou, X. M., Yao, Q. H., Zhang, Z., Peng, R. H., Xiong, A. S., and Wang, H. K. (2007). Expression of the human hepatitis B virus large surface antigen gene in transgenic tomato plants. *Clin. Vaccine Immunol.* 14, 464-469. 72.

Lu, Z., Xu, X., Hu, X., Zhu, G., Zhang, P., van Deel, E. D., French, J. P., Fassett, J. T., Oury, T. D., Bache, R. J., and Chen, Y. (2008). Extracellular superoxide dismutase deficiency exacerbates pressure overload-induced left ventricular hypertrophy and dysfunction. *Hypertension* 51, 19-25. 73.

MacNee, W., and Rahman, I. (1995). Oxidants/antioxidants in idiopathic pulmonary fibrosis. *Thorax* 1, S53-S58. 74.

Mahajan, A. S., Babbar, R., Kansal, N., Agarwal, S. K., and Ray, P. C. (2007). Antihypertensive and antioxidant action of amlodipine and vitamin C in patients of essential hypertension. *J. Clin. Biochem. Nutr.* 40, 141-147. 75.

Maria, C. S., Revilla, E., Ayala, A., de la Cruz, C. P., and Machado, A. (1995). Changes in the histidine residues of Cu/Zn superoxide dismutase during aging. *FEBS Lett.* 374, 85-88. 76.

Marklund, S. L. (1982). Human copper-containing superoxide dismutase of high molecular weight. *Proc. Natl. Acad. Sci. USA* 79, 7634-7638. 77.

Marklund, S. L. (1984). Extracellular superoxide dismutase in human tissues and human cell lines. *J. Clin. Invest.* 74, 1398-1403. 78.

Marklund, S. L., Nilsson, P., Israelsson, K., Schampi, I., Peltonen, M., and Asplund, K. (1997). Two variants of extracellular-superoxide dismutase: relationship to cardiovascular risk factors in an unselected middle-aged population. *J. Intern. Med.* 242, 5-14. 79.

McCord, J. M., and Fridovich, I. (1969). The utility of superoxide dismutase in studying free radical reactions. I. Radicals generated by the interaction of sulfite, dimethyl sulfoxide, and oxygen. *J. Biol. Chem.* 244, 6056-6063. 80.

McCord, J. M., and Fridovich, I. (1970). The utility of superoxide dismutase in studying free radical reactions. II. The mechanism of the mediation of cytochrome c reduction by a variety of electron carriers. *J. Biol. Chem.* 245, 1374-1377. 81.

McKersie, B. D., Bowley, S. R., Harjanto, E., and Leprince, O. (1996). Water-Deficit Tolerance and Field Performance of Transgenic Alfalfa Overexpressing Superoxide Dismutase. *Plant Physiol.* 111, 1177-1181. 82.

McKersie, B. D., Bowley, S. R., and Jones, K. S. (1999). Winter survival of transgenic alfalfa overexpressing superoxide dismutase. *Plant Physiol.* 119, 839-848. 83.

McKersie, B. D., Murnaghan, J., and Bowley, S. R. (1997). Manipulating freezing tolerance in transgenic plants. *Acta. Physiol. Plant.* 19, 485-495. 84.

McKersie, B. D., Murnaghan, J., Jones, K. S., and Bowley, S. R. (2000). Iron-superoxide dismutase expression in transgenic alfalfa increases winter survival without a detectable increase in photosynthetic oxidative stress tolerance. *Plant Physiol.* 122, 1427-1437. 85.

Mugge, A., Brandes, R. P., Boger, R. H., Dwenger, A., Bode-Boger, S., Kienke, S., Frolich, J. C., and Lichtlen, P. R. (1994). Vascular release of superoxide radicals is enhanced in hypercholesterolemic rabbits. *J. Cardiovasc. Pharmacol.* 24, 994-998. 86.

Ohara, Y., Peterson, T. E., and Harrison, D. G. (1993). Hypercholesterolemia increases endothelial superoxide anion production. *J. Clin. Invest.* 91, 2546-2551. 87.

Ohta, H., Adachi, T., and Hirano, K. (1993). The nature of heterogeneous components of extracellular-superoxide dismutase purified from human umbilical cords. *Free Radic. Biol. Med.* 15, 151-158. 88.

Olsen, D. A., Petersen, S. V., Oury, T. D., Valnickova, Z., Thogersen, I. B., Kristensen, T., Bowler, R. P., Crapo, J. D., and Enghild, J. J. (2004). The intracellular proteolytic processing of extracellular superoxide dismutase (EC-SOD) is a two-step event. *J. Biol. Chem.* 279, 22152-22157. 89.

Orr, W. C., and Sohal, R. S. (1994). Extension of life-span by overexpression of superoxide dismutase and catalase in *Drosophila melanogaster*. *Sci.* 263, 1128-1130. 90.

Oury, T. D., Chang, L. Y., Marklund, S. L., Day, B. J., and Crapo, J. D. (1994). Immunocytochemical localization of extracellular superoxide dismutase in human lung. *Lab. Invest.* 70, 889-898. 91.

Oury, T. D., Day, B. J., and Crapo, J. D. (1996). Extracellular superoxide dismutase in vessels and airways of humans and baboons. *Free Radic. Biol. Med.* 20, 957-965. 92.

Puget, K., and Michelson, A. M. (1974). Iron containing superoxide dismutases from luminous bacteria. *Biochimie* 56, 1255-1267. 93.

Pukacki, P. M., Kendall, E. J., and McKersie, B. D. (1991). Membrane injury during freezing stress to winter wheat *Triticum aestivum* L. crowns. *J. Plant*

Physiol. 138, 516-521. 94. Romanos, M. A., Scorer, C. A., and Clare, J. J. (1992). Foreign gene expression in yeast: a review. *Yeast* 8, 423-488. 95. Rosen, G. M., Pou, S., Ramos, C. L., Cohen, M. S., and Britigan, B. E. (1995). Free radicals and phagocytic cells. *Faseb. J.* 9, 200-209. 96. Salin, M. L., and Bridges, S. M. (1980). Isolation and characterization of an iron-containing superoxide dismutase from a eucaryote, *Brassica campestris*. *Arch. Biochem. Biophys.* 201, 369-374. 97. Samis, K., Bowley, S., and McKersie, B. (2002). Pyramiding Mn-superoxide dismutase transgenes to improve persistence and biomass production in alfalfa. *J. Exp. Bot.* 53, 1343-1350. 98. Sandstrom, J., Carlsson, L., Marklund, S. L., and Edlund, T. (1992). The heparin-binding domain of extracellular superoxide dismutase C and formation of variants with reduced heparin affinity. *J. Biol. Chem.* 267, 18205-18209. 99. Sandstrom, J., Karlsson, K., Edlund, T., and Marklund, S. L. (1993). Heparin-affinity patterns and composition of extracellular superoxide dismutase in human plasma and tissues. *Biochem. J.* 294 ( Pt 3), 853-857. 100. Sandstrom, J., Nilsson, P., Karlsson, K., and Marklund, S. L. (1994). 10-fold increase in human plasma extracellular superoxide dismutase content caused by a mutation in heparin-binding domain. *J. Biol. Chem.* 269, 19163-19166. 101. Santi, L., Batchelor, L., Huang, Z., Hjelm, B., Kilbourne, J., Arntzen, C. J., Chen, Q., and Mason, H. S. (2008). An efficient plant viral expression system generating orally immunogenic Norwalk virus-like particles. *Vaccine* 26, 1846-1854. 102. Scholthof, H. B., Scholthof, K. B., and Jackson, A. O. (1996). Plant virus gene vectors for transient expression of foreign proteins in plants. *Annu. Rev. Phytopathol.* 34, 299-323. 103. Schubert, B. K. (1994). Glutathione, glutathione reductase and freezing stress in alfalfa *Medicago sativa* L. MSc thesis, University of Guelph. 104. Searcy, K. B., and Searcy, D. G. (1981). Superoxide dismutase from the Archaeobacterium *Thermoplasma acidophilum*. *Biochim. Biophys. Acta.* 670, 39-46. 105. Senaratna, T., McKersie, B. D., and Stinson, R. H. (1984). Association between Membrane Phase Properties and Dehydration Injury in Soybean Axes. *Plant Physiol.* 76, 759-762. 106. Senaratna, T., McKersie, B. D., and Stinson, R. H. (1985a). Antioxidant Levels in Germinating Soybean Seed Axes in Relation to Free Radical and Dehydration Tolerance. *Plant Physiol.* 78, 168-171. 107. Senaratna, T., McKersie, B. D., and Stinson, R. H. (1985b). Simulation of Dehydration Injury to Membranes from Soybean Axes by Free Radicals. *Plant Physiol.* 77, 472-474. 108. Sessa, W. C., Pritchard, K., Seyedi, N., Wang, J., and Hintze, T. H. (1994). Chronic exercise in dogs increases coronary vascular nitric oxide production and endothelial cell nitric oxide synthase gene expression. *Circ. Res.* 74, 349-353. 109. Smolenska, L., Roberts, I. M., Learmonth, D., Porter, A. J., Harris, W. J., Wilson, T. M., and Santa Cruz, S. (1998). Production of a functional single chain antibody attached to the surface of a plant virus. *FEBS Lett* 441, 379-382. 110. Steinberg, D. (1995). Role of oxidized LDL and antioxidants in atherosclerosis. *Adv. Exp. Med. Biol.* 369, 39-48. 111. Strausz, J., Muller-Quernheim, J., Stepling, H., and Ferlinz, R. (1990). Oxygen radical production by alveolar inflammatory cells in idiopathic pulmonary fibrosis. *Am. Rev. Respir. Dis.* 141, 124-128. 112. Stromqvist, M., Holgersson, J., and Samuelsson, B. (1991). Glycosylation of extracellular superoxide dismutase studied by high-performance liquid chromatography and mass spectrometry. *J. Chromatogr.* 548, 293-301. 113. W. Y., Folz, R., Chen, J. S., Crapo, J. D., and Chang, L. Y. (1997). Extra-cellular superoxide dismutase mRNA expressions in the human lung by in situ hybridization. *Am. J. Respir. Cell. Mol. Biol.* 16, 162-170. 114. Tackaberry, E. S., Dudani, A. K., Prior, F., Tocchi, M., Sardana, R., Altsaar, I., and Ganz, P. R. (1999). Development of biopharmaceuticals in plant expression systems: cloning, expression and immunological reactivity of human cytomegalovirus glycoprotein B (UL55) in seeds of transgenic tobacco. *Vaccine* 17, 3020-3029. 115. Tainer, J. A., Getzoff, E. D., Richardson, J. S., and Richardson, D. C. (1983). Structure and mechanism of copper, zinc superoxide dismutase. *Nature* 306, 284-287. 116. Tanaka, Y., Hibino, T., Hayashi, Y., Tanaka, A., Kishitani, S., Takabe, T., Yokota, S., and Takabe, T. (1999). Salt tolerance of transgenic rice overexpressing yeast mitochondrial Mn-SOD in chloroplasts. *Plant Sci.* 148, 131-138. 117. Tibell, L., Aasa, R., and Marklund, S. L. (1993). Spectral and physical properties of human extracellular superoxide dismutase: a comparison with CuZn superoxide dismutase. *Arch. Biochem. Biophys.* 304, 429-433. 118. van Poppel, G., and van den Berg, H. (1997). Vitamins and cancer. *Cancer Lett.* 114, 195-202. 119. Vance, C. K., and Miller, A. F. (2001). Novel insights into the basis for *Escherichia coli* superoxide dismutase's metal ion specificity from Mn-substituted FeSOD and its very high E(m). *Biochemistry* 40, 13079-13087. 120. Walmsley, A. M., and Arntzen, C. J. (2000). Plants for delivery of edible vaccines. *Curr. Opin. Biotechnol.* 11, 126-129. 121. White, C. R., Brock, T. A., Chang, L. Y., Crapo, J., Briscoe, P., Ku, D., Bradley, W. A., Gianturco, S. H., Gore, J., Freeman, B. A., and et al. (1994). Superoxide and peroxynitrite in atherosclerosis. *Proc. Natl. Acad. Sci. USA* 91, 1044-1048. 122. Yamada, H., Yamada, Y., Adachi, T., Fukatsu, A., Sakuma, M., Futenma, A., and Kakumu, S. (2000). Protective role of extracellular superoxide dismutase in hemodialysis patients. *Nephron* 84, 218-223. 123. Yamada, H., Yamada, Y., Adachi, T., Goto, H., Ogasawara, N., Futenma, A., Kitano, M., Miyai, H., Fukatsu, A., Hirano, K., and Kakumu, S. (1997). Polymorphism of extracellular superoxide dismutase (EC-SOD) gene: relation to the mutation responsible for high EC-SOD level in serum. *Jpn. J. Hum. Genet.* 42, 353-356. 124. Yost, F. J., Jr., and Fridovich, I. (1973). An iron-containing superoxide dismutase from *Escherichia coli*. *J. Biol. Chem.* 248, 4905-4908. 125. Yu, J., and Langridge, W. H. (2001). A plant-based multicomponent vaccine protects mice from enteric diseases. *Nat. Biotechnol.* 19, 548-552. 126. Zelko, I. N., Mariani, T. J., and Folz, R. J. (2002). Superoxide dismutase multigene family: a comparison of the CuZn-SOD (SOD1), Mn-SOD (SOD2), and EC-SOD (SOD3) gene structures, evolution, and expression. *Free Radic. Biol. Med.* 33, 337-349.