

腦垂體腺? 酸環化? 激活? 扣顫d郭魚體外培養性腺組織之表現及調控研究

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摘要

腦垂體腺?酸環化?激活?(pituitary adenylate cyclase activating polypeptide, PACAP)為一神經胜? (neuropeptide)，屬於血管活性腸?(vasoactive intestinal peptide, VIP)/胰泌素(secretin)/昇醣素(glucagon)/生長激素釋放素(growth hormone releasing hormone, GHRH)家族成員之一。PACAP於生物體內具有兩種不同構形，其胺基酸序列從N端組成38個及27個胺基酸兩種構形，分別命名為PACAP38與PACAP27，二者於哺乳類具有調控細胞凋亡、代謝、內分泌及免疫系統等重要之功能，PACAP及其受體於斑馬魚(zebrafish)、薩克愛鮭(sockeye salmon)、泰國鯰(Thai catfish)、彩虹鱒魚(rainbow trout)及紅鰭東方豚(fugu rubripes)等其他魚種器官之分佈已有探討，但於吳郭魚除腦外，於其他各器官並無相關之研究。本研究除將針對tPACAP38及PAC1-R於吳郭魚各臟器及組織之分佈與表現進行分析外，另以dibutyryl-cAMP、ovine PACAP38、forskolin及H89等藥物對體外培養之吳郭魚性腺組織(精巢及卵巢)進行劑量及時間相關之試驗，以探討這些藥物對性腺tPACAP38分泌之影響。以半定量反轉錄?-聚合?鏈鎖反應(semi-quantitative RT-PCR)及PCR雜合反應(PCR hybridization)分析發現tPACAP38及PAC1-R，除肝臟外，於吳郭魚之腦、膽囊、鰓、心臟、腸、腎臟、肌肉、胃、精巢及卵巢等組織中皆有表現，且於雄、雌魚腦中tPACAP38及其受體之表現量均顯著高於其他組織者($P < 0.05$)，於其他組織之表現量則無顯著之差異($P > 0.05$)。另以免疫組織化學染色法(immunohistochemistry)分析PACAP及其受體蛋白質表現與分佈所得之結果，與前述各組織之表現相吻合。以不同濃度之cAMP(dibutyryl-cAMP; 0.25, 1.5 and 5 mM)誘導於體外培養之吳郭魚性腺組織2小時後，或以1.5 mM之cAMP濃度或5 nM ovine PACAP38誘導經不同培養時間(0, 2, 4 and 8 h)後，tPACAP38之mRNA表現量於2小時後之表現量較高，並隨著誘導時間增加其表現量則有明顯下降之趨勢。續以forskolin(1, 5 and 10 μ M)及H89(10 μ M)進行誘導tPACAP38表現之結果顯示，發現forskolin雖為腺?環化?(adenylate cyclase)之活化劑，但其之誘導作用可受蛋白激?A(protein kinase A, PKA)抑制劑H89所抑制，因此tPACAP38之表現可能與cAMP-PKA之訊號傳遞路徑有關，推測tPACAP38可能於硬骨魚類扮演旁分泌或自分泌(paracrine/autocrine)調控者之角色。

關鍵詞：吳郭魚、腦垂體腺?酸環化?激活?、腦垂體腺?酸環化?激活?第一型受體、性腺、訊號傳遞

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- 參考文獻 邵廣昭。1996。台灣常見魚貝類圖說（下）。台灣省漁業局。pp. 125-146。 Adams, B.A., Lescheid, D.W. and Vickers, E.D. (2002) Pituitary adenylate cyclase activating polypeptide and growth hormone-releasing hormone-like peptide in sturgeon, white, grayling, flounder and halibut: cDNA sequence, exon skipping and evolution. *Regul Pept*, 109: 27-37. Adamou, J.E., Aiyar, N., Van Horn, S. and Elshourbagy, N.A. (1995). Cloning and functional characterization of the human vasoactive intestinal peptide (VIP)-2 receptor. *Biochem Biophys Res Commun*, 209: 385-392. Apa, R., Lanzone, A., Mastrandrea, M., Miceli, F., Macchione, E., Fulghesu, A.M., Caruso, A. and Canipari, R. (1997). Effect of pituitary adenylate cyclase-activating peptide on meiotic maturation in follicle-enclosed, cumulus-enclosed, and denuded rat oocytes. *Biol Reprod*, 57: 1074-1079. Arimura, A. and Shioda, S. (1995). Pituitary adenylate cyclase activating polypeptide (PACAP) and its receptors: neuroendocrine and endocrine interaction. *Front Neuroendocrinol*, 16: 53-88. Arimura, A. (1998). Perspectives on pituitary adenylate cyclase activating polypeptide (PACAP) in the neuroendocrine, endocrine, and nervous systems. *Jpn J Physiol*, 48(5): 301-331. Arimura, A., Somogyvari, V.A., Weill, C., Fiore, R.C., Tatsuno, I., Bay, V. and Brenneman, D.E. (1994). PACAP function as a neurotrophic factor. *Ann N Y Acad Sci*, 739: 228-243. Basille, M., Gonzalez, B.J., Desrues, L., Demas, M., Fournier, A. and Vaudry, H. (1995). Pituitary adenylate cyclase-activating polypeptide (PACAP) stimulates adenylyl cyclase and phospholipase C activity in rat cerebellar neuroblasts. *J Neurochem*, 65(3): 1318-1324. Barberi, M., Muciaccia, B., Morelli, M.B., Stefanini, M., Ceconni, S. and Canipari, R. (2007). Expression localisation and functional activity of pituitary adenylate cyclase-activating polypeptide, vasoactive intestinal polypeptide and their receptors in mouse ovary. *Reproduction*, 134(2): 281-292. Bhogal, R., Sheldrick, R.L. and Coleman, R.A. (1994). The effects of PACAP and VIP on guinea pig tracheal smooth muscle in vitro. *Peptides*, 15(7): 1237-1241. Canny, B.J., Rawlings, S.R. and Leong, D.A. (1992). Pituitary adenylate cyclase-activating polypeptide specifically increases cytosolic calcium ion concentration in rat gonadotropes and somatotropes. *Endocrinology*, 130(1): 211-215. Cardoso, J.C., Vet, E.C.D., Louro, B., Elgar, G., Clark, M.S. and Power, D.M. (2007). Persistence of duplicated PAC1 receptors in the teleost, *Sparus auratus*. *BMC Evol Biol*, 7(1): 221. Cauvin, A., Robberecht, P., and De Neef, P. (1991). Properties and distribution of receptors for pituitary adenylate cyclase activating peptide (PACAP) in rat brain and spinal cord. *Regul Pept*, 35(2): 161-173. Chartrel, N., Tonon, M.C. and Vaudry, H. (1991). Primary structure of frog pituitary adenylate cyclase-activating polypeptide (PACAP) and effects of ovine PACAP on frog pituitary. *Endocrinology*, 129(6): 3367-371. Delgado, M., De La Ruente, M. and Martine Z.C. (1995). Pituitary adenylate cyclase activating polypeptide (PACAP27 and PACAP38) inhibit the mobility of murine thymocytes and splenic lymphocytes: comparison with VIP and implication of cAMP. *J Neuroimmunol*, 62(2): 137-146. Filipsson, K., Martina, K.R. and Bo, A. (2001). The neuropeptide pituitary adenylate cyclase-activating polypeptide and islet function. *Diabetes*, 50(9): 1959-1969. Filipsson, K., Sundler, F., Hannibal, J. and Ahren, B. (1998). PACAP and PACAP receptors in insulin producing tissues: localization and effects. *Regul Pept*, 74: 167-175. Fradinger, E.A. and Sherwood, N.M. (2000). Characterization of the gene encoding both growth hormone-releasing hormone (GRF) and pituitary adenylate cyclaseactivating polypeptide (PACAP) in the zebrafish. *Mol Cell Endocrinol*, 165: 211-219. Fradinger, E.A., Tello, J.A., Rivier, J.E. and Sherwood, N.M. (2005). Characterization of four receptor cDNAs: PAC1, VPAC1, a novel PAC1 and a partial GHRH in zebrafish. *Mol Cell Endocrinol*, 231(1-2): 49-63. Ghatei, M.A., Takahashi, K. and Suzuki, Y. (1993). Distribution, molecular characterization of pituitary adenylate cyclase-activating polypeptide and its precursor encoding messenger RNA in human and rat tissues. *J Endocrinology*, 136(1): 1559-1566. Girard, B.A., Lelievre, V., Brass, K.M., Bazinia, T., Vizzard, M.A. and Ioffe, Y. (2006). Noncompensation in peptide/receptor gene expression and distinct behavioral phenotypes in VIP- and PACAP-deficient mice. *J Neurochem*, 99: 499-513. Gottschall, P.E., Tatsuno, I. and Miyata, A. (1990). Haracterization and distribution of binding sites for the hypothalamic peptide, pituitary adenylate cyclase activating polypeptide. *Endocrinology*, 127(1): 272-277. Gras, S., Hannibal, J., Georg, B. and Fahrenkrug, J. (1996). Transient periovulatory expression of pituitary adenylate cyclase activating peptide in rat ovarian cells. *Endocrinology*, 137: 4779-4785. Gras, S., Hedetoft, C., Pedersen, S.H. and Fahrenkrug, J. (2000). Pituitary adenylate cyclase-activating peptide stimulates acute progesterone production in

rat granulosa/lutein cells via two receptor subtypes. *Biol Reprod*, 63: 206-212. Hashimoto, H., Ishihara, T. and Shigemoto, R. (1993). Molecular cloning and tissue distribution of a receptor for pituitary adenylate cyclase-activating polypeptide. *Neuron*, 11(2): 333-342. Heindel, J.J., Sneeden, J., Powell, C.J., Davis, B. and Culler, M.D. (1996). A novel hypothalamic peptide, pituitary adenylate cyclase-activating peptide, regulates the function of rat granulosa cells in vitro. *Biol Reprod*, 54: 523-530. Hou, X., Vandermeers, A. and Gourlet, P. (1994). Structural requirements for the occupancy of rat brain PACAP receptors and adenylate cyclase activation. *Neuropharmacology*, 33(10): 1189-1195. Hu, Z., Lelievre, V., Tam, J., Cheng, J.W., Fuenzalida, G., Zhou, X. and Waschek, J.A. (2000). Molecular cloning of growth hormone-releasing hormone/pituitary adenylyl cyclase-activating polypeptide in the frog *Xenopus laevis*: brain distribution and regulation after castration. *Endocrinology*, 141: 3366-3376. Inagaki, N., Yoshida, H., Mizuta, M., Mizuno, N., Fujii, Y., Gonoi, T., Miyazaki, J. and Seino, S. (1994). Cloning and functional characterization of a third pituitary adenylate cyclase-activating polypeptide receptor subtype expressed in insulin-secreting cells. *Proc Natl Acad Sci USA*, 91: 2679-2683. Ishihara, T., Shigemoto, R., Mori, K., Takahashi, K. and Nagata, S. (1992). Functional expression and tissue distribution of a novel receptor for vasoactive intestinal polypeptide. *Neuron*, 8(4): 811-819. Issac, R. and Sherwood, N.M. (2008). Pituitary adenylate cyclase-activating polypeptide (PACAP) is important for embryo implantation in mice. *Mol Cell Endocrinol*, 280(1-2): 13-19. Ito, Y., Kozawa, O. and Tokuda, H. (1994). Glucocorticoid inhibits cAMP production induced by vasoactive agents in aortic smooth muscle cells. *Atherosclerosis*, 110(1): 69-76. Jaworski, D.M. and Proctor, M.D. (2000). Developmental regulation of pituitary adenylate cyclase-activating polypeptide and PAC(1) receptor mRNA expression in the rat central nervous system. *Brain Res Dev Brain Res*, 120(1): 27-39. Kanemura, T., Tamaoki, J. and Chiyotani, A. (1993). Role of Na⁺-K⁺-ATPase in airway smooth muscle relaxation by vasoactive intestinal peptide and pituitary adenylate cyclase activating peptide. *Res Commun Chem Pathol Pharmacol*, 79(1): 11-22. Kantor, O., Heinzlmann, A., Suzuki, N., Vincze, E., Kocsis, K. and Kovacs, K. (2002). Distribution of PACAP and its mRNA in several nonneuronal tissues of rats demonstrated by sandwich enzyme immunoassay and RT-PCR technique. *Regul Pept*, 109: 103-105. Kastner, A., Bruch, L. and Will, S.L. (1995). Pituitary adenylate cyclase activating peptide are endothelium-independent dilators of human and porcine coronary arteries. *Agents Actions*, 45: 283-289. Katsoulis, S., Clemens, A. and Schworer, H. (1993). Pituitary adenylate cyclase activating polypeptide (PACAP) is a potent relaxant of the rat ileum. *Peptides*, 14(3): 587-592. Kimura, C., Ohkubo, S., Ogi, K., Hosoya, M., Itoh, Y., Onda, H., Miyata, A., Jian, L., Dahi, R.R., Stibbs, H.H., Arimura, A. and Fujino, M. (1990). A novel peptide which stimulates adenylate cyclase: molecular cloning and characterization of the ovine and human cDNAs. *Biochem Biophys Res Commun*, 166(1): 81-89. Kivipelto, L., Abood, A. and Arimura, A. (1992). The distribution of pituitary adenylate cyclase activating polypeptide-like immunoreactivity is distinct from helodermin and helospectin-like immunoreactivities in the rat brain. *J Chem Neuroanat*, 5(1): 85-94. Ko, C., In, Y.H. and Park-Sarge, O.K. (1999). Role of progesterone receptor activation in pituitary adenylate cyclase activating polypeptide gene expression in rat ovary. *Endocrinology*, 140: 5185-5194. Ko, C. and Park-Sarge, O.K. (2000). Progesterone receptor activation mediates LH induced type-I pituitary adenylate cyclase activating polypeptide receptor (PAC1) gene expression in rat granulosa cells. *Biochem Biophys Res Commun*, 277: 270-279. Koh, P.O., Kwak, S.D., Kang, S.S., Cho, G.J., Chun, S.Y., Kwon, H.B. and Choi, W.S. (2000). Expression of pituitary adenylate cyclase activating polypeptide (PACAP) and PACAP type I A receptor mRNAs in granulosa cells of preovulatory follicles of the rat ovary. *Mol Reprod Dev*, 55: 379-386. Koh, P.O., Noh, H.S., Kim, Y.S., Cheon, E.W., Kim, H.J., Kang, S.S., Cho, G.J. and Choi, W.S. (2003). Cellular localization of pituitary adenylate cyclase-activating polypeptide in the rat testis. *Mol Cells*, 15(2): 271-276. Kwok, Y.Y., Chu, Y.S., Vaudry, H., Yon, L., Anouar, Y. and Chow, K.C. (2006). Cloning and characterization of a PAC1 receptor hop-1 splice variant in goldfish (*Carassius auratus*). *Gen Comp Endocrinol*, 145(2): 188-196. Kozicz, T., Vigh, S. and Arimura, A. (1998). The source of origin of PACAP- and VIP-immunoreactive fibers in the laterodorsal division of the bed nucleus of the stria terminalis in the rat. *Brain Res*, 810(1-2): 211-219. Lacombe, A., Lelievre, V., Roselli, C.E., Salameh, W., Lue, Y.H., Lawson, G., Muller, J.M., Waschek, J.A. and Vilain, E. (2006). Delayed testicular aging in pituitary adenylate cyclase-activating peptide (PACAP) null mice. *Proc Natl Acad Sci USA*, 103(10): 3793-3798. Lee, J., Park, H.J., Choi, H.S., Kwon, H.B., Arimura, A., Lee, B.J., Choi, W.S. and Chun, S.Y. (1999). Gonadotropin stimulation of pituitary adenylate cyclase activating polypeptide (PACAP) messenger ribonucleic acid in the rat ovary and the role of PACAP as a follicle survival factor. *Endocrinology*, 140: 818-826. Lee, L.T.O., Siu, F.K.Y., Tam, J.K.V., Lau, I.T.Y., Wong, A.O.L., Lin, M.C.M., Vaudry, H. and Chow, B.K.C. (2007). Discovery of growth hormone-releasing hormones and receptors in nonmammalian vertebrates. *Proc Natl Acad Sci USA*, 104(7): 2133-2138. Lo, C.W., Chang, S.L. and Weng, C.F. (2007). Pituitary adenylate cyclase activating polypeptide (PACAP) regulates the expression of PACAP in cultured tilapia astrocytes. *Exp Biol Med*, 232(2): 262-276. Lutz, E.M., Sheward, W.J., West, K.M., Morrow, J.A., Fink, G. and Harmar, A.J. (1993). The VIP2 receptor: molecular characterisation of a cDNA encoding a novel receptor for vasoactive intestinal peptide. *FEBS Lett*, 334(1): 3-8. Masuo, Y., Suzuki, N., Matsumoto, H. (1993). Regional distribution of pituitary adenylate cyclase activating polypeptide (PACAP) in the rat central nervous system as determined by sandwich-enzyme immunoassay. *Brain Res*, 602(1): 57-63. Masuda, Y., Ohtaki, T. and Kitada, C. (1990). Solubilization of receptor for pituitary adenylate cyclase activating polypeptide from bovine brain. *Biochem Biophys Res Commun*, 172(2): 709-714. Matsudaa, K., Kashimoto, K., Higuchi, T., Yoshida, T., Uchiyama, M., Shioda, S., Arimura, A. and Okamura, T. (2000). Presence of pituitary adenylate cyclase-activating polypeptide (PACAP) and its relaxant activity in the rectum of a teleost, the stargazer, *uranoscopus japonicus*. *Peptides*, 21(6): 821-827. McRory, J.E., Parker, D.B., Ngamvongchon, S. and Sherwood, N.M. (1995). Sequence and expression of cDNA for pituitary adenylate cyclase activating polypeptide (PACAP) and growth hormone-releasing hormone (GHRH)-like peptide in catfish. *Mol Cell Endocrinol*, 108(1-2): 169-177. McRory, J.E., Parker, R.L. and Sherwood, N.M. (1997). Expression and alternative processing of a chicken gene encoding both growth hormone releasing hormone and pituitary adenylate cyclase activating polypeptide. *DNA Cell Biol*, 16(1): 95-102. Miyata, A., Arimura, A., Dahl, R.R., Minamino,

N., Uehara, A., Jiang, L., Culler, M.D. and Coy, D.H. (1989). Isolation of a novel 38 residue hypothalamic polypeptide which stimulates adenylyl cyclase in pituitary cells. *Biochem Biophys Res Commun*, 164(1): 567-574. Miyata, A., Jiang, L., Dahl, RD., Kitada, C., Kubo, K., Fujino, M., Minamino, N. and Arimura, A. (1990). Isolation of a neuropeptide corresponding to the N-terminal 27 residues of the pituitary adenylyl cyclase activating polypeptide with 38 residues (PACAP38). *Biochem Biophys Res Commun*, 170(2): 643-648. Montpetit, C.J., Shahsavari, A. and Perry, S.F. (2003). Localisation of VIP-binding sites exhibiting properties of VPAC receptors in chromaffin cells of rainbow trout (*Oncorhynchus mykiss*). *J Exp Biol*, 206: 1917-1927. Morrow, J.A., Lutz, E.M., West, K.M., Fink, G. and Harmar, A.J. (1993). Molecular cloning and expression of a cDNA encoding a receptor for pituitary adenylyl cyclase activating polypeptide (PACAP). *FEBS Lett*, 329: 99-105. Nandha, K.A., Benito, O.M.A. and Smith, D.W. (1991). Action of pituitary adenylyl cyclase-activating polypeptide and vasoactive intestinal polypeptide on the rat vascular system: effects on blood pressure and receptor binding. *J Endocrinol*, 129: 69-73. Ogi, K., Kimura, C., Onda, H., Arimura, A. and Fujino, M. (1990). Molecular cloning and characterization of cDNA for the precursor of rat pituitary adenylyl cyclase activating polypeptide (PACAP). *Biochem Biophys Res Commun*, 173(3): 1271-1279. Ohkubo, S., Kimura, C. and Ogi, K. (1992). Primary structure and characterization of the precursor to human pituitary adenylyl cyclase activating polypeptide. *DNA Cell Biol*, 11(1): 21-30. Okazaki, K., Itoh, Y., Ogi, K., Ohkubo, S. and Onda H. (1995). Characterization of murine PACAP mRNA. *Peptides*, 16(7): 1295-1299. Pantaloni, C., Brabet, P., Bilanges, B., Dumuis, A., Houssami, S., Spengler, D., Bockaert, J. and Journot, J. (1996). Alternative splicing in the N-terminal extracellular domain of the pituitary adenylyl cyclase-activating polypeptide (PACAP) receptor modulates receptor selectivity and relative potencies of PACAP-27 and PACAP-38 in phospholipase C activation. *J Bio Chem*, 271(36): 22146-22151. Parker, D.B., Power, M.E., Swanson, P., Rivier, J. and Sherwood, N.M. (1997). Exon skipping in the gene encoding pituitary adenylyl cyclase-activating polypeptide in salmon alters the expression of two hormones that stimulate growth hormone release. *Endocrinology*, 138(1): 414-423. Pesce, M., Canipari, R., Ferri, G.L., Siracusa, G. and De Felici, M. (1996). Pituitary adenylyl cyclase-activating polypeptide (PACAP) stimulates adenylyl cyclase and promotes proliferation of mouse primordial germ cells. *Development*, 122: 215-221. Pisegna, J.R. and Wank, S.A. (1993). Molecular cloning and functional expression of the pituitary adenylyl cyclase activating polypeptide type I receptor. *Proc Natl Acad Sci USA*, 90(13): 6345-6349. Rawlings, S.R. and Hezareh, M. (1996). Pituitary adenylyl cyclase activating polypeptide (PACAP) and PACAP / vasoactive intestinal polypeptide receptors: action on the anterior pituitary gland. *Endocr Rev*, 17(1): 4-29. Sayasith, K., Brown, K.A. and Sirois, J. (2007). Gonadotropin-dependent regulation of bovine pituitary adenylyl cyclase-activating polypeptide in ovarian follicles prior to ovulation. *Reproduction*, 133(2): 441-453. Scaldaferri, L., Arora, K., Lee, S.H., Catt, K.J. and Moretti, C. (1996). Expression of PACAP and its type-I receptor isoforms in the rat ovary. *Mol Cell Endocrinol*, 117: 227-232. Schadlow, V.C., Barzilai, N. and Deutsch, P.T. (1992). Regulation of gene expression in PC12 cells via an activator of dual second messengers: pituitary adenylyl cyclase activating polypeptide. *Mol Biol Cell*, 3(8): 941-951. Schafer, H., Schwarzhö, R. and Creutzfeldt, W. (1991). Characterization of a guanosine-nucleotide-binding-coupled receptor for pituitary adenylyl cyclase activating polypeptide on plasma membranes from rat brain. *Eur J Biochem*, 202(3): 951-958. Sherwood, N.M., Krueckl, S.L. and McRory, J.E. (2000). The origin and function of the pituitary adenylyl cyclase-activating polypeptide (PACAP)/glucagon superfamily. *Endocr Rev*, 21(6): 619-670. Sherwood, N.M. and Sheng, W. (2005). Developmental role of GnRH and PACAP in a zebrafish model. *Gen Comp Endocrinol*, 142(1-2): 74-80. Sherwood, N.M., Adams, B.A., Isaac, E.R., Wu, S. and Fradinger, E.A. (2007). Knocked down and out: PACAP in development, reproduction and feeding. *Peptide*, 28: 1680-1687. Small, B.C. and Nonneman, D. (2001). Sequence and expression of a cDNA encoding both pituitary adenylyl cyclase activating polypeptide and growth hormone-releasing hormone-like peptide in channel catfish (*Ictalurus punctatus*). *Gen Comp Endocrinol*, 122(3): 354-363. Spengler, D., Waeber, C., Pantaloni, C., Holsboer, F., Bockaert, J., Seeburg, P.H. and Journot, L. (1993). Differential signal transduction by five splice variants of the PACAP receptor. *Nature*, 365(6442): 170-175. Sundler, F., Ebklad, E., Absood, A., Hakanson, R., Koves, K. and Arimura, A. (1992). Pituitary adenylyl cyclase activating peptide: a novel vasoactive intestinal peptide-like neuropeptide in the gut. *Neuroscience*, 46(2): 439-54. Svoboda, M., Tastenoy, M., Van Rampelbergh, J., Goossens, J.F., De Neef, P., Waelbroeck, M. and Robberecht, P. (1994). Molecular cloning and functional characterization of a human VIP receptor from SUP-T1 lymphoblasts. *Biochem Biophys Res Commun*, 205: 1617-1624. Uddman, R., Luts, A., Arimura, A. and Sundler, F. (1991). Pituitary adenylyl cyclase activating peptide (PACAP), a new vasoactive intestinal peptide (VIP)-like peptide in the respiratory tract. *Cell Tissue Res*, 265: 197-201. Wang, Y., Wong, O.L. and Ge, W. (2003). Cloning, regulation of messenger ribonucleic acid expression, and function of a new isoform of pituitary adenylyl cyclase-activating polypeptide in the zebrafish ovary. *Endocrinology*, 144(11): 4799-4810. Watanabe, T., Nakamachi, T., Matsuno, R., Hayashi, D., Nakamura, M., Kiruyama, S., Nakajo, S. and Shioda, S. (2007). Localization, characterization and function of pituitary adenylyl cyclase-activating polypeptide during brain development. *Peptide*, 28(9): 1713-1719. Watanabe, T., Shimamoto, N. and Takakashi, A. (1995). PACAP stimulates catecholamine release from adrenal medulla: a novel noncholinergic secretagogue. *Am J Physiol*, 269: E903-909. Wong, A.O., Leung, M.Y., Shea, W.L., Tse, L.Y., Chang, J.P. and Chow, B.K. (1998). Hypophysiotropic action of pituitary adenylyl cyclase-activating polypeptide (PACAP) in the goldfish: immunohistochemical demonstration of PACAP in the pituitary, PACAP stimulation of growth hormone release from pituitary cells, and molecular cloning of pituitary type I PACAP receptor. *Endocrinology*, 139(8): 3465-3479. Yada, T., Sakurada, M., Filipsson, K., Kikuchi, M. and Ahren, B. (2000). Intraperitoneal PACAP administration decreases blood glucose in GK rats and in normal and high-fat diet mice. *Ann NY Acad Sci*, 321: 259-263. Zia, F., Fagarasan, M. and Bitar, K. (1995). Pituitary adenylyl cyclase activating peptide receptors regulate the growth of non-small cell lung cancer cells. *Cancer Res*, 55(21): 4886-4891.