

Cloning and Expression of the para-Hydroxybenzoate Hydroxylase Gene from *Pseudomonas aeruginosa* and ...

謝佩芬、簡宏堅

E-mail: 9701206@mail.dyu.edu.tw

ABSTRACT

L-DOPA is a medicine to prevent Parkinson's disease getting worse, therefore para-hydroxybenzoate hydroxylase (HBHD) from *Pseudomonas aeruginosa* PAO1 was chosen to substitute for tyrosine hydroxylase to transform L-tyrosine become L-DOPA to achieve the treatment. So hbhd gene was used as template to design primer pair. Chromosome DNA of *P. aeruginosa* PAO1 was used as template to amplify hbhd gene by PCR. Cloning of hbhd gene into *Escherichia coli* and expression of enzymatic activities were performed. The length of hbhd performed open reading frame (ORF) was 1,185 bp. The translation HBHD product was 45 kDa of molecular weight. The *P. aeruginosa* hbhd gene was cloned into pQE30 expression vector and transformed to *E. coli* Nova Blue. Finally, Ni-NTA column was used to purify the enzyme. With the para-hydroxybenzoate as substrate, enzyme activity was maximal at 80 °C, the optimum pH 9.0 assayed by high performance liquid chromatography, Fe²⁺ ion promoted enzyme activity. When the concentration of para-hydroxybenzoate reached to 100 mM, enzyme activity was measured without any substrate inhibition. With the L-tyrosine as substrate, enzyme activity was maximal at 30 °C, the optimum pH 3.5 characterized by HPLC. Without metal ion, the HBHD is still active. When the concentrations of L-tyrosine and L-DOPA reached to 80 mM and 3.8 mM, the HBHD activity was reduced to 50 % with the substrate and product inhibition, respectively.

Keywords : L-DOPA ; gene cloning ; high performance liquid chromatography

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REFERENCES

- Boonyapiwat B, Forbes B, Steventon GB. (2004) Phenylalanine hydroxylase: possible involvement in the S-oxidation of S-carboxymethyl-L-cysteine. *Anal Biochem.* 1;335(1):91-7.
- Costa RM, Lin SC, Sotnikova TD, Cyr M, Gainetdinov RR, Caron MG, Nicoletti MA. (2006) Rapid alterations in corticostriatal ensemble coordination during acute dopamine-dependent motor dysfunction. *Neuron* 52(2):359-69.
- Daubner SC, Fitzpatrick PF. (1998) Mutation to phenylalanine of tyrosine 371 in tyrosine hydroxylase increases the affinity for phenylalanine. *Biochemistry.* 37(46):16440-4.
- Entsch B, Palfey BA, Ballou DP, Massey V. (1991) Catalytic function of tyrosine residues in para-hydroxybenzoate hydroxylase as determined by the study of site-directed mutants. *J Biol Chem.* 266(26):17341-9.
- Eppink MH, Schreuder HA, van Berkel WJ. (1998) Lys42 and Ser42 variants of p-hydroxybenzoate hydroxylase from *Pseudomonas fluorescens* reveal that Arg42 is essential for NADPH binding. *Eur J Biochem.* 253(1):194-201.
- Fujii T, Kaneda T. (1985) Purification and properties of NADH/NADPH-dependent p-hydroxybenzoate hydroxylase from *Corynebacterium cyclohexanicum*. *Eur J Biochem.* 147(1):97-104.
- Ginns EI, Rehavi M, Martin BM, Weller M, O'Malley KL, LaMarca ME, McAllister CG, Paul SM. (1988) Expression of human tyrosine hydroxylase cDNA in invertebrate cells

using a baculovirus vector. *J Biol Chem.* 263(15):7406-10. 8. King EO, Ward MK, Raney DE (1970) Two simple media for the demonstration of pyocyanin and fluorescin. *J Lab Clin Med.* 44 (2) :301-7. 9. Mulholland AJ and Ridder L. (2003) Caught in the act: modelling how a biological catalyst works. *CSAR Focus* 10,12-13. 10. Nishimura M, Kumamoto Y, Shibuya A, Hirose T, Tsukamoto T, Ohya S. (1990) An in vitro study on the treatment of complicated cystitis using an automatic simulator. *Kansenshogaku Zasshi.* 64(8):1004-12. 11. Randhir, T.O. (2003) Watershed-scale effects of urbanization on sediment export: Assessment and policy. *Water Resour Res.* 39(6):1-13. 12. Saiki I, Murata J, Iida J, Sakurai T, Nishi N, Matsuno K, Azuma I..(1989) Antimetastatic effects of synthetic polypeptides containing repeated structures of the cell adhesive Arg-Gly-Asp (RGD) and Tyr-Ile-Gly-Ser-Arg (YIGSR) sequences. *Br J Cancer.* 60(5):722-8. 13. Seibold B, Matthes M, Eppink MH, Lingens F, Van Berkel WJ, Muller R. (1996) 4-Hydroxybenzoate hydroxylase from *Pseudomonas* sp. CBS3. Purification, characterization, gene cloning, sequence analysis and assignment of structural features determining the coenzyme specificity. *Eur J Biochem.* 239(2):469-78. 14. Van Berkel W, Westphal A, Eschrich K, Eppink M, de Kok A. (1992) Substitution of Arg214 at the substrate-binding site of p-hydroxybenzoate hydroxylase from *Pseudomonas fluorescens*. *Eur J Biochem.* 210(2):411-9. 15. Vervoort J, Rietjens IM, van Berkel WJ, Veeger C. (1992) Frontier orbital study on the 4-hydroxybenzoate-3-hydroxylase-dependent activity with benzoate derivatives. *Eur J Biochem.* 206(2):479-84. 16. Wijnands RA, Muller F. (1982) A study of p-hydroxybenzoate hydroxylase from *Pseudomonas fluorescens*: chemical modification of histidine residues. *Biochemistry.* 21(26):6639-46.