Effect of Dipping Treatment in -PGA on Freshness and Quality of Tilapia during Refrigeration

邱欣潁、柯文慶;謝昌衛

E-mail: 9607830@mail.dyu.edu.tw

ABSTRACT

Live tilapia (Orechromis niloticus) was used as the material in this study. Based on the method of three piece filleting, tilapia was dissected into pieces and grouped into two categories, of which were raw fish fillets (with skin) and skinned fish steaks that had been resized into 3cm × 2cm × 1cm. Both groups of fish fillets and steaks were soaked with three kinds of solutions by different molecular mass and forms of 0.5% -PGA. The solutions used included high molecular weight (HM), low molecular weight (LM) and Na + form Hydrogel (Hy). In addition, fish fillets were also soaked with solutions of 0.5% sodium tripolyphosphate (P) and RO water (R) as the controls. The changes of appearance and freshness of each sample stored at 4 for 0-8 days were investigated. The results were shown as follows. 1. There was insignificantly change in skin color of tilapia fillets during storage, however, the skin of fillets soaked -PGA were moister than those with P and R. At the 8 – day storage, the meat color of R and HM fillets appeared to be brown, but that of P insignificantly changed. The result of odor analysis showed relative low scores in both groups in R and HM and the highest score in in P group. After 8-day storage, the losing weight of P fillet showed the highest, and that of LM fillet showed the lowest in this study. 2. Hunter b value of all the fish steaks increased with elongation of storage. At the 7-day storage, R and Hy steaks had the highest and lowest b values of -0.42 and -1.55 in this study, respectively. The results were similar to the changes of appearance and TBA. At the 8 – day storage, TBA value showed the lowest in HM steak (0.088) in this study, and that showed the lowest in R steak (0.147). The results showed that soaking with -PGA solution treatment could inhibit lipid oxidation of the fish steaks. The results of VBN values (mg/100g meat) showed that R and P steaks exceeded the hygienic standard at the 7-day storage., and Hy steak still fitted in the hygienic standard at the 8 – day storage. K values of the meats also increased with storage. The meats reached the putrid level at the 6-day and the 7-day storage for R and P steaks, respectively, while K value reached 83.67% for R steak and 73.99% for P steak. Moreover, K value reached 63.79 % for LM steak at the 8-day storage. Moisture levels of the meats decreased during the storage period. Both HM and LM had higher moisture contents. The levels of water-soluble proteins of the meat decreased with increasing the length of storage period. R (1.85)and HM (1.76)had significantly decreases at the 4-day storage, but for the LM (1.86) group the significant decrease happened at the 7-day storage. 3. In summary, dipping in -PGA solution is practicable for tilapia preservation based on the positive results in analysis of VBN, K value, TBA value, and the changes in weight loss and appearance. LM group and Hy group resulted in better quality in freshness of fish meats and are potential replacements for polyphosphate.

Keywords: -聚麩胺酸; 吳郭魚; 鮮度與品質

Table of Contents

目錄 封面內頁 簽名頁 授權書	iii 中文摘要	iv 英文摘要vi
誌謝viii 目錄	ix 圖目錄	xiii 表目
錄xvi 1. 緒言	12. 文獻回顧	3 2.1 吳郭
魚32.2 水產物鮮度鑑定法	4 2.2.1 感官鑑定法	4 2.2.2 化學鑑定
法5 2.2.3 物理鑑定法	10 2.2.4 微生物法	10 2.3 水產物的低溫保
鮮11 2.3.1 冷藏法	11 2.3.2 冷凍法	11 2.3.3 部分冷凍
法12 2.4 化學保鮮劑	12 2.4.1 亞硫酸類	12 2.4.2 聚合磷酸鹽
類13 2.4.3 維生素 C	14 2.4.4 五倍子酸	14 2.4.5 有機
酸15 2.5 生物保鮮劑	15 2.5.1 菌體二次代謝	產物保鮮15 2.5.2 多醣類物質
保鮮16 2.5.3 生物?保鮮	16 2.5.4 生物體自生的天	然成分提取物 16 2.6 聚麩胺酸的
介紹16 2.6.1 聚麩氨酸之發現與其	結構16 2.6.2 聚麩氨區	酸之製備17 2.6.3 聚麩氨酸
之特性20 2.6.4 聚麩氨酸之應用	23 3. 材料與方法	27 3.1 材
料27 3.1.1 吳郭魚	27 3.1.2 保鮮劑	27 3.2 試
藥30 3.3 儀器	31 3.4 實驗方法	33 3.4.1 實驗設計與流
程33 3.4.2 鮮度之測定	37 3.4.2.1 pH值之測定	

定37 3.4.2.3 揮發性鹽基態氮之測定	37 3.4.2.4 總生菌數之測定		
定	42 3.4.2.7 氣味評估	42 3.4.3 肌肉特性之測	
定42 3.4.3.1 水份含量之測定	42 3.4.3.2 保水力	43 3.4.3.3 失重率之測	
定43 3.4.3.4 煮失率之測定	43 3.4.3.5 截切值之測定	44 3.4.3.6 蛋白質溶解度之	
測定44 3.4.3.7 肌動凝蛋白 Ca-ATPase 活性	之測定 46 4. 結果與討論	49 4.1 鮮度之測	
定49 4.1.1 pH 值	49 4.1.2 魚肉外觀	53 4.1.3 魚肉色	
澤56 4.1.4 揮發性鹽基態氮	62 4.1.5 總生菌數	62 4.1.6 K	
值	65 4.1.8 氣味評估	68 4.2 肌肉特	
性	率70 4.2.2 水分含量	70 4.2.3 保水	
力73 4.2.4 失重率	73 4.2.5 截切值	76 4.2.6 蛋白質溶解	
度76 4.2.7 肌動凝蛋白 Ca-ATPase 活性77 4.3 不同浸漬液處理之保鮮度評估 81 5. 結			
論83 參考文獻	84		

REFERENCES

1.牟敦剛、江晃榮,2000,幾丁質在生技產品:醫療、食品及環保上之應用。 2.大森丘、重久保。1989。食肉???食肉製品??高壓利用。食 品?開發。24: 54-56。 3.太田靜行。1991。水產物?鮮度保持。筑波書局。東京。日本。 4.李嘉馨。2006。電子高壓誘導裝置貯藏對吳郭 魚鮮度與品質之影響。大葉大學大葉大學生物產業科技學系碩士論文。彰化。台灣。 5.劉蕙菁。2003。花腹鯖與虱目魚在不同溫度中生 物胺及鮮度品質之變化。國立台灣海洋大學食品科學研究所碩士論文。基隆。台灣。 6.林亮全、黃鑑宇、曾富元。2002。不同浸泡和 包裝處理對台灣商用土雞儲藏期間肉質之影響 1.對 pH 值、色澤、總生菌數、VBN 值、TBA 值、截切值之影響。台灣農業化學與食品 科學。41:176-188。 7.呂玟蒨。2006。聚麩胺酸鈉鹽製造微膠囊技術之研發與包覆納豆激?功效之評估。國立中興大學食品暨應用生物科 技學系。台中。台灣。 8.柯文慶、張?瑞、賴滋漢。2003。食品加工。第 33-35 頁。富林出版社。台灣。 9.胡興華。1996。拓漁台灣。第 33-35 頁。行政院農委會漁業署。台北。台灣。 10.齊藤恆內、內山均、梅本滋、河川俊治。1974。水產生物化學。食品學實驗書。p. 267-281。恆星社厚生閣。東京。日本。 11.邱萬敦。2002。漁獲物的保鮮與處理。第 95-98 頁。翠柏林企業股份公司。台中。台灣。 12. 須山三千山、鴻巢章二。1987。水產食品學。第 17-37 頁。恆星社厚生閣。日本。東京。 13.徐國強。1998。高壓常溫貯藏吳郭魚肌肉 之鮮度保持與加工適性。國立中興大學食品科學研究所碩士論文。台中。台灣。 14.新井健一。1977。多獲性赤身魚?有效利用。水產 學????35。恆星社厚生閣。日本。東京。 15.行政院衛生署。1998。食字第 87032655 號公告修正。行政院衛生署。台北。台灣。 16.行政 院農委會漁業署。2005。中華民國台灣地區漁業年報。台北。台灣。 17.中國國家標準。1982。冷凍魚類檢驗法。CNS 1451 N6029。經 濟部中央標準局。台北。台灣。 18.中國國家標準。1996。食品微生物之檢驗法—生菌數檢驗。CNS10890。經濟部中央標準局。台北。 台灣。 19.張為憲。2001。食品化學。華香園出版社。台北。 20.陳建州。2003。聚麩胺酸之生物絮凝性質的研究。大葉大學食品工程學 系碩士論文。彰化。台灣。 21.陳文騰。1999。生鮮吳郭魚在流通期間之品質變化與控制。國立中興大學食品科學研究所。台中。台灣 。 22.陳憶馨。2005。以聚麩胺酸水溶膠吸附 Hydralazine HCI 進行釋放之研究。大葉大學生物產業科技學系碩士論文。彰化。台灣。 23.陳雅玲, 2003, 聚麩胺酸的物性與化妝品應用之研究。靜宜大學應用化學系碩士論文。台中。台灣。 24.邵奕遠。 2003。以批次醱酵 槽生產聚麩胺酸及其抗凍性之研究。大葉大學論文。彰化。台灣。 25.周榮吉、林高塚、吳建平。1995。酸液浸漬及包冰處理對凍藏雞 胸肉品質之影響。嘉義農專學報。40:37-49. 26.邵廣昭。1996。台灣常見魚介貝類圖說(下)-魚類。第 174-175 頁台灣省漁業局。 27.曾明 義。2005。吳郭魚出口加工產業現況分析。國立台灣海洋大學水產養殖研究所碩士論文。基隆。台灣。 28.蔡佳玲。2006。收穫後處理 與包裝對海鱺、吳郭魚與鱸魚品質與 5 儲藏期限之影響。國立台灣海洋大學食品科學研究所碩士論文。基隆。台灣。 29.蘇遠志 。2003。納豆菌代謝產物的開發與應用。生物產業14(2):17-30。 30.吳熊清、邱思魁。1996。水產食品學。國立編譯館。台北。台灣。 31.吳淑靜、柯文慶、賴滋漢。2003。食品添加物。第 106-160 頁。富林出版社。台中。台灣。 32.AOAC. 1984. Official Methods of Analysis. 14th ed. Association of Official Analytical Chemists, Washing, D.C. USA. 33. Bhattacharya, M. and Hanna M. A. 1989. Kinetics of drip loss, cooking loss and color degradation in frozen ground beef during storage. J. Food Eng. 9: 83-96. 34. Bhattacharyya, D., Hestekin, J.A., Brushaber, P., Cullen, L.G. and Sikdar, S. K. 1998. Novel poly-glutamic acid functionalized microfiltration membranes for sorption of heavy metal at high capacity. J. Membrane 141: 121-135. 35.Bovarnick, M. 1942. The formation of extracellular D-glutamic acid polypeptide by Bacillus subtilis. J. Bio. Chem. 145: 415-424 36.Bramsnaes, F. 1981. Maintaining the quality of frozen foods during distribution. Food Technol. 35: 38. 37. Chen, H. C., Moody, M. W., and Jiang, S. T. 1990. Changes in biochemical and bacteriological Quality of grass pawn during transportation by icing and oxygenating. J. Food Sci. 55: 670-673. 38. Davies, J. R., Bardsly, R. G. and Ledward, D. A. 1988. Myosin Thermal Stability in fish Muscle. J. Food Sci. 45: 61-68. 39. Dickson, J. S. and Anderson, M. E. 1992. Microbiological decontamination food animal carcasses by washing and sanitizing systems: A review. J. Food Prot. 55: 133-140. 40. Dyer, W. J., French, H. V., and Snow, J. M. 1950. Protein in fish muscle. 1. Extraction of protein fraction in flesh fish. J. Fish. Res. Board Can. 7: 585-593. 41. Fey, M. S. and Regenstein, J. M. 1982. Extending shelf life of fresh wet red hake and salmon using CO2-O2 modified atmosphere and potassium sorbate ice at 1 . J. Food Sci. 47: 1048-1054. 42. Fleming, S. E., Sosulski, R. W., Kilara, A. and Humbert, E. S. 1974. Viscosity and water absorption characteristics of slurries of sunflower and soybeen flours, concentrates and isolates. J. Food Sci. 39:188-191. 43. Florene, G., Touraille, C., Oual, A., Renerre, M. and Moni, G. 1994. Relationships between postmortem pH change and some traits of sensory quality in veal. Meat Sci. 37: 315-325. 44.Fujii, H. 1963. On the formation of mucilage by

bacillus natto. Part IIIchemical constitutions of mucilage in natto (1). Nippon Nogeikagaku Kaishi 37: 407-411. 45. Giulivi, C. and Cadenas, E. 1993. The reaction of ascorbic acid with different heme iron redox states of myoglobin. Antioxidant and prooxidant aspects. FEBS Lett. 332: 287-290. 46. Gorman, B. M., J. N. Sofos, J. B. Morgan, G. R. Schmidt and G. C. Smith. 1995. Evaluation of hard-trimming, various sanitizing agent and hot water spraying – washing as decontamination interventions for beef brisket adipose tissue. J. Food Prot. 58: 899-907. 47. Gornall. A.G., Bardawill, C.T., and David, M.M. 1949. Determination of serum proteins by means of the biuret reactions. J. Biol. Chem. 177: 715-766. 48.Gram, L. 1991. Inhibition of mesophilic spoilage Aeromonas spp. On fish by salt, potassium sorbate, liguid smoke, and chilling. J. Food Prot. 54: 436-441. 49. Hollender, R., F. G. Bender, R. K. Jenkins, and C. L. Black. 1993. Research note: Consumer evaluation of chicken treated with a trisodium phosphate application during processing. Poultry Sci. 72:755-759. 50. Honikel, K. O. 1987. The water binding of meat. Fleischwirtsch. 67: 1098-1100. 51. Hultin, H. O. 1985. Characteristics of muscle tissue. In: O. R. Fennema (Ed.). Food chemistry, 2nd ed. Marcel Dekker, Inc., Madison Ave., New York.USA.723-789. 52. Ikigai, H., Nakae, T., Hara, Y. and Shimamura, T. 1993. Bactericidal catechins damage the lipid bilayer. Biochimica. Biophysica Acta 1147:132-136. 53. Ito, Y. 1996. Glutamic acid independent production of Poly (-glutamic acid) by Bacillus Subtilis TMA-4. J. Biosci. Biotechnol Biochem. 60: 1239-1242. 54. Jayasingh, P. and Cornforth, D. P. 2003. Comparison of antioxidant effects of milk mineral, butylated hydroxytoluene and sodium tripolyphosphate in raw and cooked ground pork. Meat Sci. 66:83-89. 55. Jiang, S. T., Wang, F. J., and Chen, C. S. 1989. Properties of actin and stability of the actomyosin reconstituted from milkfish (Chanos chanos) actin and myosin. J. Agric. Food Chem. 37: 1232-1235. 56. Johnson, L. N., Phillips, D. C. and Rupley, J. A. 1968. The activity of lysozyme: An Interim review of crystall ographic and chemical evidence. Brookhaven Symp. Biol. 21: 120~138. 57.Ko, W. C. 1996. Efect of high pressure on gelation of meat paste and inactivation of actomyosin Ca-ATPase prepared from milkfish. Fisheries Sci. 62: 101-104. 58. Katoh, N., Nozaki, H., Komatsu, L. and Arai, K. 1979. A new method for evaluation of the quality of frozen surimi from Alaska pollack relationship between myofibrillar ATPase activity and kamaboko forming ability of frozen surimi. Bull. Japan. Soc. Sci. Fish. 45: 1027-1032. 59. Kunno, A., Taguchi T. and Yamaguchi, T. 1998. Bakery products and noodles containing polyglutamic acid. US Patent 4: 888,193. 60. Kutota, H., Nambu, Y., Takeda, H. and Endo, T. 1992. Poly-gamma- glutamic acid ester and shaped boby hereof. US Patent 5: 118,784. 61. Kutota, H., Nambu, Y. and Endo, T. 1993. Convenient and quantitative esterification of poly(-gutamic acid) produced by microorganism. J. Polym. Sci. Part A Polym Chem. 31: 2877-2878. 62.Labuza, T. P. 1985. An integrated approach to food chemistry. In "Food Chemistry", Fennema, O. R. Ed., p. 766-772. Dekker, New York. 63.LeBlance, E. L., Leblance, R. J., and Gill. T. A. 1987. Effect of pressure processing on frozen stored muscle protein of Atlantic cod (Gadus mohua)fillets. J. Food Prot. 11:209-235. 64.Martin, J. B. and Doty, D. M. 1949. Determination of inorganic phosphate. Anal. Chem. 21: 965. 65.Matsuda, Y. 1979. Influence of sucrose on the protein denaturation of lyophilized carp myofibrils during storage. Bull. Jap. Soc. Sci. Fish. 45: 573-579. 66. Matsumoto, J. J. 1979. Denaturation of fish muscle proteins during frozen storage. In "Protein at Low Temperature", (Ed.), p. 205-224. By O. Fennema, ACS. Washington D.C. 67.Ohnishi, T., Gall, R. S., and Mayer, M. L. 1975. An improved assay of inorganic phosphate in the presence of extractable phosphate compound: application to the ATPase assay in the presence of phosphocreatine. Anal. Biochem. 69: 261-267. 68. Owusu-Anshah. Y. O. and Hultin, H. O. 1986. Chemical and physical changes in red hake fillets during frozen storage. J. Food. Sci. 51: 1402-1406. 69. Perez-Camero, G., Congregado, F., and Bou, J. 1999. Biosynthesis and ultra-sonic degradation of bacterial poly -glutamic acid. Biotechnol. bioeng. 63: 110-115. 70. Perez-villarreal, B. and Pozo, R. 1990. Chemical composition and ice spoilage of albacore (Thunnus alalunga). J. Food, Sci. 55: 678, 71. Post, L. S., Lee, D. A., Solberg, M., Furgang, D., Speecchio, J. and Garham, C. 1985, Development of botulinal toxin and sensory deterioration during storage of vacuum and modified atmosphere packaged fish fillets. J. Food. Sci. 50: 990. 72. Price, R. J., Melvin, E. F. and Bell, J. W. 1991. Postmortem changes in chilled round, bled and dessed albacoe. J J. Food. Sci. 73. Ritchie, S. M. C., Bachas, L. G., Olin, T., Sikdar, S. K. and Bhattacharyya, D. 1999. Surface modification of silica and cellulose-based microfilitration membranes with functional polyamino acids for heavy metal sorption. Langmuir. 15:6346-6357. 74. Ryder, J. M. 1985. Determination of adenosine triphosphate and its breakdown products in fish muscle by high-performance liquid chromatography. J. Agric Food Chem. 33: 678-680. 75. Sathivel, S. 2005. Chitosan and Protein Coatings Affect Yield, Moisture Loss, and Lipid Oxidation of Pink Salmon (Oncorhynchus gorbuscha) Fillets During Frozen Storage. J. Food Sci. 70:455-459. 76.Saito, T. and Arai. K. 1959. A new method for estimating the freshness of fish. Bull. Jap. Soc. Sci. Fish. 24: 749~750. 77.Sanda, F. Fujiyama, T, and Endo, T. 2001. Chemical synthesis of polygamma glutamic acid by polycondesation of gamma glutamic acid methyester. J. Polym Science Part A-1. 39: 732-741. 78. Scannell, A. G., Ross, R. P., Hill, C. 2000. An effective lacticin biopreservative in fresh pork sausage. J. Food Prot. 63: 370-375 79. Shewan, J.M., MacIntosh, R.G., Tucker, C.G. and Ehrenberg, A.S.C., 1953. The development of a numerical scoring system for the sensory assessment of the spoilage of wet white fish stored in ice. J. Sci. Food. Agric. 4: 283 – 298. 80. Shih, I. L., Van, Y. T., Yeh, L. C., Lin, H. G. and Chang, Y. N. 2001. Production of a biopolymer flocculant from Bacillus licheniformis and its flocculation properties. Bioresour Technol. 78: 267-271. 81. Siger, J. W., De, Vries, P. and Bhatt, R. 2000. Conjugation of camptothesins to poly(L-glutamic acid). Ann NY Acas Sci. 922: 136-150. 82. Sigholt, T., Erikson, U., Rustad, T., Johansen, S., Nordtvedt, T. S. and Seland, A. 1997. Handling stress and storage temperature affect meat quality of farmed-raised Atlantic salmon (Slmo salar) J. Food Sci. 62: 898-905 83. Sigurgisladottir, S., Hafsteinsson, H., Jonsson, A., Nortvedt, R., Thomassen, M. and Torrissen, O. 1999. Textural Properties of Raw Salmon Fillets as Related to Sampling Method. J. Food Sci. 64: 99-104. 84.Sikorski, Z. E., Olley, J., and Kostuch, S. 1976. Protein changes in frozen fish. CRC Crit. Rev. Food Sci. Nutr. 8: 97-129. 85.Smulders, F. J. M. 1987. Preospectives for microbial decontamination of meat and poultry by organic acid with special reference to lactic acid Elimination of pathogenic organisms from meat and poultry.319-344. Elsevier. 86. Srikar, L.N. and Reddy, G.V.S. 1991. Protein solubility and emulsifying capacity in frozen stored fish mince. J. Sci. Food Agric. 55:447-453. 87. Stanley, D. W. 1983. Relation of structure to physical properties of animal

material. In "Physical Properties of Foods." M. Peleg, and EB. Bagley(Ed). p.157-206. AVI Publishing Compony Inc. Westport, CT. USA. 88. Taguchi, T., Kikuchi, K., Oguni, M., Tanaka, M., and Suzuki., K. 1978. Heat change of myosin B Mg-ATPase and "Setting" of fish meat paste. Bull. Japan. Soc. Sci. Fish. 44: 1363-1368. 89. Tanimoto, H. H., Kuuraishi, C. Kido, K. and Seguto, K. 1995. High absorption mineral-containing composition and food. US patent US Patent US5447: 732. 90. Watabe, S., Kamal, M. and Hashimoto, K. 1991. Postmortem changes in ATP, creatine phosphate, and lactate in sardine muscle. J. Food Sci. 56: 151-153. 91. Watanabe, A., Tsuneishi, E., and Takimoto. Y. 1989. Analysis of ATP and its breakdown products in beef by reversed-phase HPLC. J. Food Sci. 54: 1169-1172. 92. Zhang, Y., Lu, H., Levin, R. E. 2003. Enhanced storage-life of fresh haddock fillets with stabilized sodium chlorite in ice. Food Microbiology 20: 87-90. 93. Yokoi, H., Natsuda, O., Hirose, J., Hayashi, S. and Takasaki, Y. 2001. Characteristics of a biopolymer flocculant produced by Bacillus sp. PY-90. J. Ferment Bioeng. 79: 378-380.