## The Study on Brewing Sorghum Spirits by Amylo Process

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#### **ABSTRACT**

Amylo is a method used to liquefy grains with water and hydrochloric acid by cooking in high temperature and high pressure, and after cooling of the cooked grains, fungus and yeasts are inoculating to make koji cultivated, starch liquefied and saccharified, yeasts inoculating and fermentation performed at the same time in a large fermentation tank, that saveed the skill to cultivate separately and production space. In this thesis, the amylo method was used to prepare sorghum spirits. This thesis can be divided into five parts: In the first part of this thesis, the best condition for the hydrolysis and the fermentation of sorghum grains were investigated. In the second part of this thesis, the optimum addition concentration and time of hexanoic acid into sorghum paste during fermentation to get better acceptance of the sorghum spirits were investigated, and to compare the difference of components analysis between before and after storage. In the third part of this thesis, the changes of volatile compounds during storage in the sorghum spirits with or without hexanoic acid added during fermentation were investigated. In the fourth part of this thesis, the optimum addition ratio and time of mixed acid that the chosen concentration of hexanoic acid confection with acetate and butyric acid in the same concentration into sorghum paste during fermentation to get better acceptance of the sorghum spirits were investigated, and also to compare the difference of components analysis between before and after storage. In the fifth part of this thesis, the changes of volatile compounds during storage in the sorghum spirits with different ratio of mixed acid added during fermentation were investigated. The results showed that the most acceptable fermentation condition to prepare spirits is that each kilogram red sorghum milled with 3L water, then adjusting pH to 4.5 with citric acid, followed by adding 2mL liquefaction enzyme (SpezymeR Fred -amylase) and reacted for 90 minutes at 95 , then after cooling to 50 of the sorghum paste, 3mL saccharification enzyme (OptimaxR HP7525) was added and reacted for another 72 hrs. Finally, 10 grams of commercial yeast (Red Star Distiller 's Active Dry Yeast) was added in the paste fermented 72 hrs at 26 ± 2 . The sorghum spirits obtained from the fermented paste with 4mL of 2.5% hexanoic acid ethanol solution added at 36 hrs ' fermentation duration was found to be more acceptable. The yield of sorghum spirits was found to decrease with the increasing of the concentrantion of hexanoic acid added. During storage, the content of total acid and total ester were found to increase with the increasing of the storage time. The major volatile compounds found in the sorghum spirits with or without hexanoic acid added during fermentation were n-propanol, butanol, isobutanol, 2-butanol, pentanol, isoamyl alcohol, 2-phenyl ethanol, acetal, acetaldehyde, acetic acid, propanoic acid, butyric acid, isobutyric acid, valeric acid, caproic acid, ethyl acetate, ethyl propionate, ethyl butyrate, ethyl pentanoate, ethyl caproate, ethyl myristate, ethyl palmitate, ethyl stearate, ethyl oleate, ethyl linoleate, ethyl linolenate, and ethyl lactate. After storage the content of alcohols, aldehydes and acids in the sorghum spirits with or without hexanoic acid added during fermentation were found to increase with the increasing of the storage time, whereas that of esters were found to decrease with the increasing of the storage time. The sorghum spirits obtained from the fermented paste with 4mL of 2.5% mixed acid (acetic acid: butyric acid: hexanoic acid= 2:1:5) ethanol solution added at 36 hrs ' fermentation duration was also found to be more acceptable. The yield of sorghum spirits was found to decrease with the decreasing of the ratio of hexanoic acid added. During storage, the content of total acid, total ester and total volatile compounds in the sorghum with mixed acid added during fermentation were found to increase with the increasing of the storage time.

Keywords: sorghum spirit; amylo; hexanoic acid; mixed acid; volatile compounds

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#### **REFERENCES**

1. 小崎道雄、????.?? ?、片岡二郎、山中茂、吉田集而。1998。?????餅麴(Marcha),酒(Jaanr)?蒸餾酒(Raksi)。釀協 95(2):115-122。 2. 王三郎。2000。應用微生物。高立圖書有限公司。台北,臺灣。3.王光輝。1998。清酒醪中酸度高的可能原因。製酒科技專論彙編 20:58-58。 4. 臺灣農家要覽編輯委員會。1995。臺灣農家要覽。第81-86頁。豐年社。台北,臺灣。 5. 江妍慧。2005。米酒的改造運動 。新臺灣新聞週刊 508。 6. 江金標。1980。高粱麴中微生物之特性簡介。製酒科技專論彙編 2:100-104。 7. 江茂煇。1999。氣相層析法 分析不同香型白酒之香氣成分。酒類試驗所88年度研究年報,87-99。 8. 西村驥一。1975。香料。112,45。 9. 吳鳴鈴。。2002。米酒 及穀類酒製造之簡介。食品工業月刊 34(1):8-13。 10. 沈怡方。1998。白酒生產技術全書。第124、248頁。中國輕工業出版社。北京 ,中華人民共和國。 11. 沈怡方和李大和。1996。低度白酒生產技術。中國輕工業出版社。北京,中華人民共和國。 12. 周婉萍。1997 。微生物及酵素在澱粉工業上之應用。食品工業月刊 29(4):33-43。 13. 林俊杰。1992。液態發酵高粱酒之研製(一)高粱澱粉之酵素 水解。酒類試驗所81年度研究年報,135-145。14. 林俊杰。1996a。酵母之高濃度酒精發酵。製酒科技專論彙編(18):63-85。15. 林俊 杰。1996b。釀酒有關之酵素。製酒科技專論彙編(18):158-168。 16. 林源義、黃玉蓮。1992。利用質譜檢測器鑑定蒸餾酒中之香氣成 分(一)高粱酒類香氣成分之鑑定。酒類試驗所81年度研究年報,51-62。 17. 林源義。1993。利用質譜檢測器鑑定蒸餾酒中之香氣成分 (二)不同類型高粱酒風味成分之比較。酒類試驗所82年度研究年報,213-227。18.相田浩、高橋甫、上田清基、櫪倉辰六郎、上田誠 之助。1997。新版應用微生物物學。朝倉書店。東京,日本。 19. 胡鳳綬。1988。酒類中之香氣成分。製酒科技專論彙編,139-174。 20. 胡鳳綬。1992。蒸餾酒在熟成中香氣成分之變化。製酒科技專論彙編,303-310。 21. 胡鳳綬。1994。酒中之酚類成分。製酒科技專 論彙編(16):299-304。 22. 胡鳳綬。1995。白酒的後處理。製酒科技專論彙編(17):45-47。 23. 倪德全。1982。酵母菌有機酸生成及 利用。製酒科技專論彙編 4:78-87。 24. 張弘儒。2005。山藥酒製程開發:106-108。私立大葉大學生物產業科技研究所碩士論文。彰化, 臺灣。 25. 淺野中男和黑瀨直孝。2000。清酒酵母?有機酸生成????。釀協 95(4):227-234。 26. 野白喜久雄、吉澤淑、鎌田耕造、水沼 武二。1988。釀造?事典。朝倉書店。東京,日本。 27. 陳昭安。2005。以酵素替代傳統酒麴製造清酒之研究:130-132。私立大葉大學生 物產業科技研究所碩士論文。彰化,臺灣。28.陳漢根。2002。中式白酒簡介。食品工業月刊34(1):1-7。29.陸壽鵬。1996。白酒工 藝學。中國輕工業出版社。北京,中華人民共和國。 30. 程竹青。1997。酵素在食品工業之應用與發展。食品工業月刊 29 ( 7 ) :9-20。 31. 黃及時。1994。糖化後發酵之條件探討。製酒科技專論彙編 16:299-304。 32. 黃正財。1981。酵母菌高濃度酒精生產性及耐性之生成 機構。製酒科技專論彙編 3:94-103。 33. 黃村能。1990。蒸餾酒泛論。製酒科技專論彙編,87-100。 34. 黃季芳。1983a。脂肪分解酵素 與其在釀酒上之應用。製酒科技專論彙編 25(9):26-33。 35. 黃季芳。1983b。脂肪分解酵素與其在釀酒上之應用。製酒科技專論彙編 5:62-67。 36. 黃癸林。1987。以高濃度液體發酵方式試製高粱蒸餾酒(一)。酒類試驗所76年度研究年報,125-136。 37. 黃癸林 。1988a。以高濃度液體發酵方式試製高粱蒸餾酒(二)。酒類試驗所77年度研究年報,137-146。 38. 黃淑媛。1988b。釀造酒與蒸餾酒 中風味化合物之形成。製酒科技專論彙編,175-190。39.黃燕君。2003。高粱酒釀造過程噴酸處理對高粱酒品質及風味之影響:132-136 。私立大葉大學生物產業科技研究所碩士論文。彰化,台灣。 40. 黃靜宜。2005。發酵中添加複合揮發有機酸對液態發酵液態蒸餾高粱 酒品質及風味之影響。117-118。私立大葉大學生物產業科技研究所碩士論文。彰化 , 臺灣。 41. 經濟部標準檢驗局。2004a。酒類檢驗 法 - 總酸度及揮發性酸度之測定。標準編號:14850。 42. 經濟部標準檢驗局。2004b。酒類檢驗法 - 總酯之測定。標準編號:14851。 43. 詹淑惠。2003。不同市售麴及噴酸處理對液態發酵液態蒸餾高粱酒品質之影響。私立大葉大學生物產業科技研究所碩士論文。彰化,台 灣。 44. 劉益善。1981。澱粉液化酵素及其在製酒上之應用。製酒科技專論彙編,71-79。 45. 劉益善。1993。中國傳統酒精飲料製造技 術之特性。製酒科技專論彙編 15:71-79。 46. 劉國棟、呂來福、李肇基、黃純真。1976。以阿米洛法由高粱試製米酒之研究。酒廠65年 度研究年報,169-179。47.劉懿慧。2005。發酵時添加己酸對液態發酵液態蒸餾五糧溢品質及風味之影響:121。私立大葉大學生物產業 科技研究所碩士論文。彰化,台灣。 48. 蔡正輝。1974。高粱、玉米、珍珠米以阿米洛法製酒試驗。酒類試驗所63年度研究年報 , 161-166。 49. 羅境鏜、鄭明鐘、徐松山。1991。液化酵素使用最佳條件之探討。第27屆蒸餾酒釀造技術研討會。Baker R. A. 1979. Clarifying properties of pectin fractions separated by ester content. J Agric Food Chem. 27(6):1387-1389. Beveridge T. 1997. Haze and cloud in apple juices. Crit Rev Food Sci Nutr. 37(1):75-91. 50. Burns J. K. 1991. Pectinesterase. In: Walter R. H., editor. The chemistry and technology of pectin. Academic Press. 165-188. 51. David Andrens 1987. Brewer's Guardian. 116(1):14-15,18-21. 52. Fischer, E. H. and Stein, E. A. 1954. Arch. Sci. (Geneva)7, 131. 53. Fischer, E. H. and Stein, E. A. 1961. Biochem. Preps., 8, 27. 54. Fogarty, W. M. 1983. Microbial amylase in "Microbial enzymes and biotechnology" Fogarty, W. M ed. Applied Science Publishers, 1-92. Hyun, H. H. and Zeikus, J. G. 1985a. General biochemical characterization of thermostable extracellular -amylase from Clostridium thermosulfurogenes. Appl. Environ. Microbiol. 49(5):1162-1167. 55. Hyun, H. H. and Zeikus, J. G. 1985b. Simultaneous and enhanced production of thermostable amylase and ethanol from starch by coculture of Clostridium thermosulfurogenes and Clostridium thermohydrosulfaricum. Appl. Environ. Microbiol. 49(5):1174-1181. 56. Labib A. A. S., and El-Ashwash F. A. 1995. Heat-inactivation of mango pectinesterase and polygalacturonase. Food Chem. 53(2):137-142. 57. Lafon-Lafourcade, S., Geneix, C. and Ribereau-gayon, P. 1984. Inhibition of alcoholic fermentation of grap must by fatty acids produced by yeasts and their elimination by yeat ghosts. Appl. Environ. Microbiol. 47(6):1246-1249. 58. Lehtonen, M. and Suomalainen, H. 1977. Rum. Economic microbiology.

1:595-633. 59. Masami Onishi, Guymon, J. F. and Csowell, E. A. 1977. Am. J. Enol, Vitic, Vol, 28, No.3, 152. 60. Morton, I. D. and Macleod, A. J. 1984. Food flavors Part B. The Flavor of beverages (Elsecier Science Publisher, B. V.), 272. 61. Robyt, J. F. and Whelan, W. J. 1968. In "Starch and Its Derivatives" Redley, J. A. ed. 4th ed. 430-476. 62. Schwimmer, S. and Balls, A. K. 1949. J. Biol. Chem. 179, 1063. 63. Shomer I. 1991. Protein coagulation cloud in citrus fruit extract. 1. Formation of coagulates and their bound pectin and neutral sugar. J Agric Food Chem. 39(12):2263-2266. 64. Taylor, G. T. and Kirsop, B. H. 1977. The origin of the medium chain length fatty acids present in beer. J. Inst. Brew (83):241-243. 65. Versteeg C., Rombous F. M., Spanaansen C. H. and Pilnik W. 1980. Thermostability and orange juice cloud destabilizing properties of multiple pectinesterase from orange. J Food Sci. 45:969-971. 66. Viegas, C. A., Rosa, M. F., Sa-Correia, I. and Novais, J. M. 1989. Inhibition of yeast growth by octanoic and decanoic acids produced during ethanolic fermentation. Appl. Environ. Microbiol. 55(1):21-28.