

# Screening for the Oxidative Stress Signaling Factor in Arabidopsis by Functional Complementation of a Yeast Mutant

葉怡芳、洪淑嫻

E-mail: 9501063@mail.dyu.edu.tw

## ABSTRACT

In this study, we constructed an *Arabidopsis* cDNA library to search for the factors mediating oxidative signals by complementation of a *Yap1*-deficient strain. In order to improve the screening efficiency, We developed a high throughput system based on the co-cultivation of hundreds of transformants on a 96-well microplate. To evaluate the reasonable number of co-cultivated transformations, different ratios of the wild-type strains to the mutants were prepared to mimic the real situation. It is revealed that the H<sub>2</sub>O<sub>2</sub> tolerance of the wild-type strains (or trasformants) were not sheltered when the number of pooled population was below 200. According to this system, we isolated a colony showing somewhat of a resistance to hydrogen peroxide. However, further examination indicated that the H<sub>2</sub>O<sub>2</sub> tolerance may be due to the cell mutation instead of aquiring the *Arabidopsis* cDNA. Moreover, we also verified that freezing tolerance of *Arabidopsis* induced by hydrogen peroxide was also explored physiologically in this study. Seedling pretreated with hydrogen peroxide obtained freezing tolerance which is comparable to that induced by cold acclimation. However application of EGTA (a calcium chelator), La3+ (a calcium channel inhibitor) or W7 (a protein kinase inhibitor ) prior to the pretreatment of hydrogen peroxide will reverse the freezing tolerance induced by hydrogen peroxide. Taken together, seedlings pretreated with hydrogen peroxide a series of signaling cascade that leads to the anti-freezed gene expression. Furthermore, calcium and calcium dependent protein kinase may be involved in the signaling pathways.

Keywords : Arabidopsis : hydrogen peroxide : Yap1 : freezing tolerance

## Table of Contents

封面內頁 簽名頁 授權書	iii	中文摘要	iv	英文摘要	v	誌謝	vi	目錄	vii	圖目錄	xii	第一章 緒言	.....	1 第二章 文獻回顧	.....	2 2.1 逆境對植物造成的傷害	.....	2 2.2 植物的抗氧化機制	.....	2 2.3 氧化逆境與過氧化氫之關係	.....						
與相關基因抗氧化能力	.....	7 2.6 利用功能性互補法篩選動、植物相關基因	.....	8 第三章 材料與方法	.....	10 3.1 實驗架構	.....	10 3.2 實驗菌種	.....	11 3.3 植物材料與培養	.....	11 3.4 載體	.....	12 3.5 實驗藥品	.....	12 3.6 培養基	.....	13 3.7 緩衝液及試劑	.....	14 3.8 實驗中使用之套組	.....						
16 3.9 實驗方法	.....	17 3.9.1 細胞內過氧化氫含量測定	.....	17 3.9.2 蛋白質含量測定	.....	17 3.9.3 建構阿拉伯芥基因庫	.....	17 3.9.3.1 萃取total RNA	.....	18 3.9.3.2 甲醛變性膠體電泳	.....	18 3.9.3.3 mRNA之純化	.....	19 3.9.3.4 合成cDNA	.....	20 3.9.3.5 分離cDNA片段及adaptor片段	.....	21 3.9.3.6 非變性聚丙烯醯胺凝膠電泳	.....	22 3.9.3.7 以試劑組萃取大量質體DNA	.....	22 3.9.3.8 以試劑組萃取小量質體DNA	.....	23 3.9.3.9 質體DNA之小量製備	.....	24 3.9.3.10 限制酵素剪切	.....
25 3.9.3.11 DNA 片段的回收及純化	.....	25 3.8.7.12 cDNA的黏接反應	.....	26 3.9.4 酵母菌genomic DNA萃取	.....	26 3.9.5 引子設計	.....	27 3.9.6 聚合?連鎖反應	.....	27 3.9.7 瓊脂凝膠電泳	.....	28 3.9.8 大腸桿菌電勝任細胞之製備	.....	28 3.9.9 大腸桿菌電轉形操作流程	.....	29 2.9.10 酵母菌電勝任細胞之製備	.....	29 3.9.11 酵母菌轉形作用	.....	29 3.9.12 功能性選殖	.....	30 3.9.13 酵母菌質體回收	.....	31 3.9.14 DNA定序及序列之分析比對	.....		
31 3.9.15 過氧化氫前處理使植物提高低溫耐受性	.....	32 3.9.16 植物處理抑制劑之方法	.....	32 3.9.17 電解質滲漏率之測試	.....	33 第四章 結果與討論	.....	34 4.1 阿拉伯芥細胞內生性過氧化氫含量測定	.....	34 4.2 建構阿拉伯芥基因庫	.....	34 4.3 Yap1基因剔除酵母菌株鑑定	.....	40 4.4 篩選系統之建立	.....	44 3.4.1 野生株與突變株對過氧化氫耐受性之差異性	.....	44 3.4.2 集中培養之合理菌落數評估	.....	44 4.5 分析菌株抗過氧化氫之能力測	.....						

試.....	49	4.6 過氧化氫前處理使植物提高低溫耐受性.....	52	4.7 過氧化氫提高阿拉伯芥抗凍能力需透過細胞外鈣離子.....	58	4.8 以鈣離子通道抑制劑lanthanum chloride前處理植株使植株提高電解質滲漏率.....	59	4.9 過氧化氫提高阿拉伯芥抗凍能力需透過蛋白質激?.....	63	第五章 結論.....
— pYES2 vector map.....										67 附錄
										76

## REFERENCES

1. 李國維。1999。阿拉伯芥超氧歧化?調控之研究。國立台灣大學物研究所碩士論文。台灣，台北。
2. Allan, A.C., and Fluhr, R. 1997. Two distinct sources of elicited reactive oxygen species in tobacco epidermal cells. *Plant Cell* 9: 1559-1572.
3. Alvarez, M.E., Pennell, R.I., Meijer, P.J., Ishikawa, A., Dixon, R.A. and Lamb, C. 1998. Reactive oxygen intermediates mediate a systemic signal network in the establishment of plant immunity. *Cell* 92: 773-784.
4. Alvarez, S. M., Rufenacht, K., and Eggen R. L. 2000. The oxidative stress-sensitive yap1 null strain of *Saccharomyces cerevisiae* becomes resistant due to increased carotenoid levels upon the introduction of the 60S ribosomal protein L10a. *Biochem. and Biophys. Res. Commun.* 267: 953-959.
5. Ashworth, E. N. 1989. Freezing injury in deciduous fruit crops: opportunities for chemical manipulation. *Acta Hort.* 239: 175-186.
6. Azevedo, D., Delaunay, A., Pousada, C. R., and Toledano, M.B. 2003. Two redox centers within YAP1 for H<sub>2</sub>O<sub>2</sub> And thiol-reactive chemicals signaling. *Free Radic. Biol. Med.* 35: 889- 900.
7. Belles-Boix, E., Babiyuk, E., Montagu, M. V., Inze, D., and Kushnir, S. 2000. CEO1, a new protein from *Arabidopsis thaliana*, protects yeast against oxidative damage. *FEBS Lett.* 482: 19-24.
8. Belles-Boix, E., Babiyuk, E., Montagu, M. V., Inze, D., and Kushnir, S. 2000. CEF, a Sec24 homologue of *Arabidopsis thaliana*, enhances the survival of yeast under oxidative stress conditions. *J. Exp. Bot.* 51: 1761-1762.
9. Bowler, C., Marc, V.M., Dirk, I. 1992. Superoxide dismutase and stress tolerance. *Annu. Rev. Plant Physiol. Plant Mol. Biol.* 43: 83-116.
10. Bradford, M.M. 1976. A rapid and sensitive method for the quantitation of microgram Quan ties of protein utilizing the principle of protein-dye bindings. *Anal. Biochem.* 72: 248-254.
11. Browse, J., Xin, Z. 2001. Temperature sensing and cold acclimation. *Curr. Opin. Plant Bio.* 4: 241-246.
12. Browse, J., Xin, Z. 2001. Temperature sensing and cold acclimation. *Curr. Opin. Plant Bio.* 4: 241-246.
13. Cohen, B. A., Pilpel, Y., Mitra, R. D., and Church, G. M. 2002. Discrimination between paralogs using microarray analysis : application to the Yap1p and Yap2p transcriptional networks. *Mol. Biol. Cell* 13: 1608-1614.
14. Coleman, S. T., Epping, E. A., Steggerda, S. M., and Moye Rowley, W.S. 1999. Yap1p activates gene transcription in an oxidant specific fashion. *Mol. Cell. Biol.* 19: 8302-8313.
15. Delaunay, A., Isnard, A. D., and Toledano, M. B. 2000. H<sub>2</sub>O<sub>2</sub> sensing through oxidation of the Yap1 transcription factor. *EMBO J.* 19: 5157-5166.
16. Gidrol, X., Sabelli, P. A., Fern, Y. S., and Kush, A. K. 1996. Annexin-like protein from *Arabidopsis thaliana* rescues oxyR mutant of *Escherichia coli* from H<sub>2</sub>O<sub>2</sub> stress. *Proc. Natl. Acad. Sci. USA* 93: 11268-11273.
17. Gong, M., van der Luit, A. H., Knight, M. R., and Trewavas, A. J. 1998. Heat shock induced changes in intracellular Ca<sup>2+</sup> level in tobacco seedling in relation to thermotolerance. *Plant Physiol.* 116: 429-437.
18. Grant, C. M., Collinson, L. P., Roe, J-H., and Dawes, I. W. 1996. Yeast glutathione reductase is required for protection against oxidative stress and is a target gene for YAP-1 transcriptional regulation. *Mol. Microbiol.* 21: 171-179.
19. Hariyadi, P., and Parkin, K.L. 1993. Chilling-induced oxidative stress in cucumber (*Cucumis sativus* L. cv. Calypso) seedlings. *J. Plant Physiol.* 141: 733-738.
20. Inoue, Y., Matsuda, T., Sugiyama, K., Izawa, S., and Kimura, A. 1999. Genetic analysis of glutathione peroxidase in oxidative stress reponse of *Saccharomyces cerevisiae*. *J Biol Chem* 274: 27002-27009.
21. Jena, S and Choudhuri, M.A. 1981. Glycolate metabolism of three submerged aquatic angiosperms during aging. *Aquat. Bot.* 12: 345-354.
22. Kendall, A. C., Keys, A. J., Turner, J. C., Lea, P. J., Miflin, B. J. 1983. The isolation and characterization of a catalase-deficient mutant of barley (*Hordeum vulgare* L.). *Planta* 159: 505-511.
23. Knight, H., Trewavas, A. J., and Knight, M. R. 1996. Cold calcium signaling in *Arabidopsis* involves two cellular pools and a change in calcium signature after acclimation. *Plant Cell* 8: 489-503.
24. Larkindale, J., and Knight, M. R. 2002. Protection against heat stress induced oxidative damage in *Arabidopsis* involves calcium, abscisic acid, ethylene, and salicylic acid. *Plant Physiol.* 128: 682-695.
25. Levitt, J. 1980. Response of plants to environmental stress. Vol. 1. Chilling, freezing and high temperature stresses. 2nd ed. New York, Academic Press.
26. Liu, H. T., Li, B., Shang, Z. L., Li, X. Z., Mu, R. L., Sun, D. Y., and Zhou, R.G. 2003. Calmodulin is involved in heat shock signal transduction in wheat. *Plant Physiol.* 132: 1186-1195.
27. Monroy, A. F., Fathey Sarhan, and Dhindsa, R. S. 1993. Cold induced changes in freezing tolerance, protein phosphorylation, and gene expression. *Plant Physiol.* 102: 1227-1235.
28. Monroy, A. F., and Dhindsa, R. S. 1995. Low-temperature signal transduction: induction of cold acclimation-specific genes of Alfalfa by calcium at 25°C. *Plant Cell* 7: 321-331.
29. Morgan, B. A., Banks, G. R., Toone, W. M., Raitt, D., Kuge, S., and Johnston, L. H. 1997. The Skn7 response regulator controls gene expression in the oxidative stress response of budding yeast *Saccharomyces cerevisiae*. *EMBO J.* 16: 1035-1044.
30. Neuenschwander, U., Vernooij, B., Friedrich, L., Uknnes, S., Kessmann, H., and Ryals, J. 1995. Is hydrogen peroxide a second messenger of salicylic acid in systemic acquired resistance? *Plant J.* 8: 227-233.
31. O'Kane, D., Gill, V., Boyd, P., and Burdon, R. 1996. Chilling, oxidative stress and antioxidant responses in *Arabidopsis thaliana* callus. *Planta* 198: 371-377.
32. Okuda, T., Matsuda, Y., Yamanaka, A., and Sagisaka, S. 1991. Abrupt increase in the level of hydrogen peroxide in leaves of winter wheat is caused by cold treatment. *Plant Physiol.* 97: 1265-1267.
33. Orozco-Cardenas, M., Narvaez-Vasquez, J., and Ryan, C. 2001. Hydrogen peroxide acts as a second messenger for the induction of defense genes in tomato plants in response to wounding, systemin, and methyl jasmonate. *Plant Cell* 13: 179-191.
34. Palate J. P., Levitt J., Stadelman E. J. 1977. Freezing injury in onion bulb cells. I. Evaluation of the conductivity method

and analysis of ion and sugar efflux from injured cells. *Plant Physiol.* 60: 393-397. 35. Potikha, T. S., Collins, C. C., Johnson, D. I., Delmer, D. P., and Levine, A. 1999. The involvement of hydrogen peroxide in the Differentiation of secondary walls in cotton fibers. *Plant Physiol.* 119: 849-858. 36. Prasad, T. K., Anderson, M. D., Martin, B. A., and Stewart, C. R. 1994. Evidence for chilling induced oxidative stress in maize seedling and a regulatory role for hydrogen peroxide. *Plant Cell* 6: 65-74. 37. Prasad, T.K. 1996. Mechanisms of chilling-induced oxidative stress injury and tolerance in developing maize seedlings: changes in antioxidant system, oxidation of proteins and lipids, and protease activities. *Plant J.* 10: 1017-1026. 38. Reichheld, J.P., Vernoux, T., Lardon, F., Montagu, M. V. , and Inze, D. 1999. Specific checkpoints regulate plant cell cycle progression in response to oxidative stress. *Plant J.* 17: 647-656. 39. Ren, D., Yang, H., and Zhang, S. 2002. Cell death mediated by MAPK is associated with hydrogen peroxide production in Arabidopsis. *J.Biol. Chem.* 277: 559-565. 40. Rentel, M. C., and Knight, M. R. 2004. Oxidative stress induced calcium signaling in Arabidopsis. *Plant Physiol.* 135: 1471-1479 41. Sambrook J, and Russell DW. 2001. Molecular Cloning: a laboratory manual, third edition. Cold Spring Harbor Laboratory Press, New York. 42. Tahtiharju, S., Sangwan, V., Monroy, A. F., Dhindsa, R. S., Borg, M. 1997. The induction of Kin genes in cold acclimating Arabidopsis thaliana. Evidence of a role for calcium. *Planta* 203: 442-447. 43. Uchida, A., Jagendorf, A. T., Hibino, T., Takabe, T., and Takabe. T. 2002. Effects of hydrogen peroxide and nitric oxide on both salt and heat stress tolerance in rice. *Plant Sci.* 163: 515-523. 44. Wanner, L. A and Junntila, O. 1999. Cold induced freezing tolerance in Arabidopsis. *Plant Physiol.* 120: 391-399. 45. White, P.J., Bowen, H. C., Demidchik, V., Nichols, C., Davies, J. M. 2002. Genes for calcium permeable channels in the plasma membrane of plant root cells. *Biochim. Biophys. Acta* 1564: 299-309. 46. Wingsle, G. and Hallgren, J.E. 1993. Influence of SO<sub>2</sub> and NO<sub>2</sub> exposure on glutathione, superoxide dismutase and glutathione reductase activities in Scots pine needles. *J. Exp. Bot.* 44: 463-470. 48. Yu, C. W., Murphy T. M., Sung, W. W, and Lin C. H. 2002. H<sub>2</sub>O<sub>2</sub> treatment induces glutathione accumulation and chilling tolerance in mung bean. *Funct. Plant Biol.* 29: 1081-1087. 49. Yu, C. W., Murphy T. M., and Lin C. H. 2003. Hydrogen peroxide induced chilling tolerance in mung beans mediated through ABA independent glutathione accumulation. *Funct. Plant Biol.* 30: 995-963. 50. Zhang, L., Onda, K., Imai, R., and Fukuda, R. 2003. Growth temperature downshift induces antioxidant response in *Saccharomyces cerevisiae*. *Biochem and Biophl Res Commun.* 307: 308-314.