

# Combine methylation sensitive electrophoresis and suppression subtractive hybridization to identify differential methylation

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## ABSTRACT

ABSTRACT Imprinting gene phenomenon may be taking place in fertilized period, according the allele to producing different epigenetic marks in this period, utilize these mark to distinguish paternal or maternal allele, have already known that DNA methylation may influence the nuclear chromatin structure or been involved in a certain protein and nucleic acid combination. And the nucleic acid methylation is one kind of the epigenetic mechanism, more evidences present that the DNA methylation controlled the expression of some basic genes and tissue specific genes. Igf2 and Snrpn are expressed from the paternal genome while H19, Igf2r and Mash2 are expressed from the maternal genome, at the time tracking the cell expression position of chimeria mouse. We combine methylation sensitive electrophoresis and suppression subtractive hybridization to identify differential methylation fragments between cortex and hypothalamus. Take the advantage of methylation sensitive restriction enzyme digestion and gel electrophoresis and suppression subtractive hybridization, we isolated a differential methylation DNA fragment between cortex and hypothalamus. The methylated site located in intron two of Zswim6. The differential methylation was confirmed by southern hybridization and methylation sensitive PCR. The expression level of Zswim6 mRNA is the highest in liver among 8 tissues, also the DNA methylation at the intron. Another short form of swim6 is also examination, the expression level is similar among tissues. Putting together, we propose the differential methylation make the post transcriptional machinery select different exon among tissues. Key Words : DNA methylation, Imprinted gene, methylation sensitive, suppression subtractive hybridization, methylation sensitive restriction enzyme, swim6

Keywords : methylation

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## REFERENCES

- 參考文獻 鄭又璋，2004，利用甲基化核糖核酸多形分析法找尋父母源標記基因，私立大葉大學分子生物科技研究所，碩士論文。
- Antequera, F. & Bird, A. (1993). Number of CpG islands and genes in human and mouse. *Proc. Natl. Acad. Sci. U S A* 90, 11995-11999.
- Bellefroid, E.J., Lecocq, P.J., Benhida, A., Poncelet, D.A., Belayew, A. & Martial, J.A. (1989). The human genome contains hundreds of genes coding for finger proteins of the Kruppel type. *Dna* 8, 377-387.
- Bestor, T.H. (2000). The DNA methyltransferases of mammals. *Hum. Mol. Genet.* 9, 2395-2402.
- Bestor, T.H. (1992). Activation of mammalian DNA methyltransferase by cleavage of a Zn binding regulatory domain. *EMBO J.* 11, 2611-2617.
- Bestor, T.H., Laudano, A.P., Mattaliano, R. & Ingram, V.M. (1988). Cloning and sequencing of a cDNA encoding DNA methyltransferase of mouse cells. The carboxyl-terminal domain of the mammalian enzymes is related to bacterial restriction methyltransferases. *J. Mol. Biol.* 203, 971-983.
- Bird, A.P. & Southern, E.M. (1978). Use of restriction enzymes to study eukaryotic DNA methylation: I. The methylation pattern in ribosomal DNA from *Xenopus laevis*. *J. Mol. Biol.* 118, 27-47.
- Byers, R.J., Hoyland, J.A., Dixon, J. & Freemont, A.J. (2000). Subtractive hybridization-genetic takeaways and the search for meaning. *Int. J. Exp. Pathol.* 81, 391-404.
- Choo, Y. & Klug, A. (1994). Selection of DNA binding sites for zinc fingers using rationally randomized DNA reveals coded interactions. *Proc. Natl. Acad. Sci. USA* 91, 11168-11172.
- Constancia, M., Pickard, B., Kelsey, G. & Reik, W. (1998). Imprinting mechanisms. *Genome Res.* 8, 881-900.
- Denissenko, M.F., Chen, J.X., Tang, M.S. & Pfeifer, G.P. (1997). Cytosine methylation determines hot spots of DNA damage in the human P53 gene. *Proc. Natl. Acad. Sci. U S A* 94, 3893-3898.
- Diatchenko, L., Lau, Y.F., Campbell, A.P., Chenchik, A., Moqadam, F., Huang, B., Lukyanov, S., Lukyanov, K., Gurskaya, N., Sverdlov, E.D. & Siebert, P.D. (1996). Suppression subtractive hybridization: a method for generating differentially regulated or tissue-specific cDNA probes and libraries. *Proc. Natl. Acad. Sci. U S A* 93, 6025-6030.
- Filippova, G.N., Qi, C.F., Ulmer, J.E., Moore, J.M., Ward, M.D., Hu, Y.J., Loukinov, D.I., Pugacheva, E.M., Klenova, E.M., Grundy, P.E., Feinberg, A.P., AM C.J., Moerland, E.W., Cornelisse, C.J., Suzuki, H., Komiya, A., Lindblom, A., F D.B., Neiman, P.E., Morse, H.C. 3rd., Collins, S.J. & Lobanenkov, V.V. (2002). Tumor-associated zinc finger mutations in the CTCF transcription factor selectively alter tts DNA-binding specificity. *Cancer Res.* 62, 48-52.
- Hagemann, C. & Blank, J.L. (2001). The ups and downs of MEK kinase interactions. *Cell Signal* 13, 863-875.
- Holliday, R. & Pugh, J.E. (1975). DNA modification mechanisms and gene activity during development. *Science* 87, 226-232.
- Hoovers, J.M., Mannens, M., John, R., Blied, J., van Heyningen, V., Poreeous, D.J., Leschot, N.J., Westerveld, A. & Little, P.F. (1992). High-resolution localization of 69 potential human zinc finger protein gene: a number are clustered. *Genomics* 12, 254-263.
- Imamura, T., Ohgane, J., Ito, S., Ogawa, T., Hattori, N., Tanaka, S. & Shiota, K. (2001). CpG island of rat sphingosine kinase-1 gene: tissue-dependent DNA methylation status and multiple alternative first exons. *Genomics* 76, 117-125.
- Issa, J.P., Vertino, P.M., Wu, J., Sazawal, S., Celano, P., Nelkin, B.D., Hamilton, S.R. & Baylin, S.B. (1993). Increased cytosine DNA-methyltransferase activity during colon cancer progression. *J. Nalt. Cancer Inst.* 85, 1235-1240.
- Jacobs, G. H. (1992). Determination of the base recognition position of zinc fingers from sequence analysis. *EMBO J.* 11, 4507-4517.
- Jeffrey, R.M. (2001). Imprinting in the germ line. *Stem Cells* 19, 287-294.
- Jeltsch, A. (2002). Beyond Watson and Crick: DNA methylation and molecular enzymology of DNA methyltransferases.

Chem.Bio.Chem. 3, 274-293. Kaffer, R., Gringerg, A. & Pfeifer, K. (2001). Regulatory mechanisms at the mouse *Igf2/H19* locus. *Mol. Cell Biol.* 21, 8189-8196. Kautiainen, T.L. & Jones, P.A. (1986). DNA methyltransferase levels in tumorigenic and nontumorigenic cells in culture. *J. Biol. Chem.* 261, 1594-1598. Keverne, E.B., Fundele, R., Narasimha, M., Barton, S.C. & Surani, M.A. (1996). Genomic imprinting and the differential roles of parental genomes in brain development. *Devel. Brain Res.* 92, 91-100. Laird, P.W. & Jaenish, R. (1996). The role of DNA methylation in cancer genetics and epigenetics. *Ann. Rev. Genet* 30, 441-464. Laity, J.H., Lee, B.M. & Wright, P.E. (2001). Zinc finger proteins: new insights into structural and functional diversity. *Curr. Opin. Struct. Biol.* 11, 39-46. Lei, H., Oh, S.P., Okano, M., Juttermann, R., Goss, K.A., Jaenisch, R. & Li, E. (1996). De novo DNA cytosine methyltransferase activities in mouse embryonic stem cells. *Development* 122, 3195-3205. Lui, K., Wang, Y., Cantemir, C. & Muller, T. (2003). Endogenous assay of DNA methyltransferase: evidence for differential activities of DNMT1, DNMT2, and DNMT3 in mammalian cells in vivo. *Mol. Cell Biol.* 23, 2709-2719. Luo, X., Budihardjo, I., Zou, H., Slaughter, C. & Wang, X. (1998). Cytosolic translocation required for MEKK-1 induced apoptosis. *Cell* 94, 481-490. Macleod, D., Ali, R.R. & Bird, A. (1998). An alternative promoter in the mouse major histocompatibility complex class II I-Abeta gene: implications for the origin of CpG islands. *Mol. Cell Biol.* 18, 4433-4443. Makarova, K.S., Aravind, L. & Koonin, E.V. (2002). SWIM, a novel Zn-chelating domain present in bacteria, archaea and eukaryotes. *Trends Biochem Sci.* 27, 384-386. Mann, J.R. (2001). Imprinting in the germ line. *Stem Cells* 19, 287-294. McGrath, J. & Solter, D. (1984). Completion of mouse embryo genesis requires both the maternal and paternal genomes. *Cell* 37, 179-183. Okano, M., Bell, D. W., Haber, D. A. & Li, E. (1999). DNA methyltransferases *Dnmt3a* and *Dnmt3b* are essential for de novo methylation and mammalian development. *Cell* 99, 247-257. Okano, M., Xie, S. & Li, E. (1998). Cloning and characterization of a family of novel mammalian DNA(cytosine-5)methyltransferases. *Nat. Genet.* 19, 219-220. Pfeifer, K. (2000). Mechanisms of genomic imprinting. *Am. J. Hum. Genet* 67, 777-787. Reik, W. & Walter, J. (2001). Genomic imprinting: parental influence on the genome. *Nature* 2, 21-32. Reinhart, B., Eljanne, M. & Richard, J. (2002). Shared role for differentially methylated domains of imprinted gene. *Mol. Cell Biol.* 22, 2089-2098. Rhee, I., Bachman, K.E., Park, B.H., Jair, K.W., Yen, R.W., Schuebel, K.E., Cui, H., Feinberg, A.P., Langeur, C., Kinzler, K.W., Baylin, S.B. & Vogelstein, B. (2002). DNMT1 and DNMT3b cooperate to silence genes in human cancer cell. *Nature* 416, 552-556. Rice, J.C., Massey-Brown, K.S. & Futscher, B.W. (1998). Aberrant methylation of the BRCA1 CpG island promoter is associated with decreased BRCA1 mRNA in sporadic breast cancer cells. *Oncogene* 17, 1807-1812. Schlesinger, T.K., Bonvin, C., Jarpe, M.B., Fanger, G.R., Cardinaux, J.R., Johnson, G.L. & Widmann, C. (2002). Apoptosis stimulated by the 91kDa caspase cleavage MEKK1 fragment requires translocation to soluble cellular compartments. *J. Biol. Chem.* 277, 10283-10291. Singer-Sam, J., Grant, M., LeBon, J.M., Okuyama, K., Chapman, V., Monk, M. & Riggs, A.D. (1990). Use of a HpaII-polymerase chain reaction assay to study DNA methylation in P<sub>gk</sub>-1 CpG island of mouse embryos at the time of X-chromosome inactivation. *Mol. Cell Biol.* 10, 4987-4989. Srivastava, M., Hsien, A. & Pfeifer, K. (2000). H19 and *Igf2* monoallelic expression is regulated in two distinct way by a shared cis acting element. *Genes* 14, 1186-1195. Stein, R., Gruenbaum, Y., Pollack, Y., Razin, A. & Cedar, H. (1982). Clonal inheritance of the pattern of DNA methylation in mouse cells. *Proc. Natl. Acad. Sci. U S A* 79, 61-65. Vanaja, D.K., Cheville, J.C., Iturria, S.J. & Young, C.Y. (2003). Transcriptional silencing of zinc finger protein 185 identified by expression profiling is associated with prostate cancer progression. *Cancer Res.* 63, 3877-82. Wigler, M., Levy, D. & Peruchio, M. (1981). The somatic replication of DNA methylation. *Cell* 24, 33-40. Yoder, J.A., Walsh, C.P. & Bestor, T.H. (1997). Cytosine methylation and the ecology of intragenomic parasites. *Trends Genet.* 13, 335-340.