

Studies on micropropagation of Moringa oleifera

蔡朋穎、余聰安

E-mail: 9417460@mail.dyu.edu.tw

ABSTRACT

The purpose of this study was to investigate Moringa oleifera micropropagation. Moringa oleifera was known as a multi-purpose, high potential, and high economical value plant. In this study, we established Moringa oleifera micropropagation system and prospectively to understand more elements about woody plant tissues culture. Shoot tip (>5 mm) were obtained from sterile F1 seeds which were culturing on MS medium and subcultured an interval of weeks. Although a high surviving rate of bud growing was observed in aseptic MS medium sowing, a failed development of multibud in addition to formation of enormous callus was found. In order to develop multibuds, we used nine various basic culture medium (1/2MS, MS, MSM, MSSh, 3/2MS, 3/2MSM, WPM, WPMm, WPMsh) mixed with different concentration of cytokinin (BA or Kin; 0, 0.1, 0.3, 0.5, 0.7, 0.9 mg·l⁻¹ respectively) and auxin (NAA or IAA 0.02, 0.04 mg·L⁻¹). The results showed that auxin caused foliar variation of explant and the optimum condition for formation of multibuds was 3/2MS medium with 0.3 mg·L⁻¹ BA. Each explant produced 5.56 buds with length of 57 mm and subculture period extended to 25 days. The optimum condition for rooting was proved in vitro rooting IBA 1000 mg·l⁻¹ (rooting rate 100%, stems: 6.5, domestication surviving rate 71%). The best efficiency (rooting rate 83%, stems: 9.1, foliar area 71 mm², domestication surviving rate 91%) was observed when combined with ventilation. In conclusion, this study successfully domesticated and produced plantlets, and built up a high efficacy and high quality Moringa oleifera micropropagation system.

Keywords : Moringa oleifera, micropropagation, multibuds, rooting, hardening.

Table of Contents

| | |
|--|--|
| 目錄 封面內頁 簽名頁 授權書----- | iii 中文摘要 |
| 要----- | iv 英文摘要 |
| 謝----- | v 誌 |
| 圖目錄----- | vii 目錄 |
| 圖目錄----- | x 表目錄 |
| 符號說明----- | xii 第一章 前言 1.1 辣木簡 |
| 介----- | 1 1.2 前人研究-----3 第二章 材 |
| 料和方法 2.1 實驗材料----- | 16 2.2 實驗方 |
| 法----- | 18 2.2.1 辣木的無菌播種-----18 2.2.2 辣木 |
| 叢生苗組織培養方法之建立----- | 19 2.2.2.1 不同濃度之不同生長調節劑對辣木芽體生長之影響-----19 2.2.2.2 不 |
| 同基本培養基組成對辣木芽體生長之影響----- | 20 2.2.2.3 不同張數通氣藥紙封瓶對辣木芽體生長之影響-----20 2.2.3 辣木之 |
| 發根處理----- | 20 2.2.3.1 不同濃度之不同生長調節劑對辣木瓶內發根之影-----20 2.2.3.2 高濃度之 |
| 不同生長調節劑對辣木瓶內發根之影----- | 21 2.2.3.3 不同張數通氣藥紙封瓶對辣木瓶內發根之影響-----21 2.2.3.4 高濃度之不 |
| 同生長調節劑對辣木瓶外發根之影----- | 21 2.2.4 組培苗的馴化-----22 2.2.5 溫室評 |
| 估----- | 22 第三章 結果 3.1 辣木叢生苗組織培養技術之建立-----23 |
| 3.1.1 不同濃度之不同生長調節劑對辣木芽體生長之影響----- | 23 3.1.2 不同基本培養基組成對辣木芽體生長之影響-----24 |
| 3.1.3 不同張數通氣藥紙封瓶對辣木芽體生長之影響----- | 25 3.2 辣木之發根處理-----25 |
| 3.2.1 不同濃度之不同生長調節劑對辣木瓶內發根之影----- | 25 3.2.2 高濃度之不同生長調節劑對辣木瓶內發根之影-----26 |
| 3.2.3 不同張數通氣藥紙封瓶對辣木瓶內發根之影----- | 26 3.2.4 高濃度之不同生長調節劑對辣木瓶外發根之影-----27 |
| 3.2.5 組培苗的馴化----- | 27 第四章 討論-----28 第 |
| 五章 結論 ----- | 33 參考文獻-----34 |
| 圖目錄 圖1在MS基本培養基添加不同濃度之不同生長調節劑，培養21天後，對辣木芽體生長之影 響----- | 圖2 MS培養基添加不同濃度之不同生長調節劑培養21天後對辣木芽體生長之結 果----- |
| xiii 圖3不同基本培養基組成對辣木芽體生長之影響----- | xiv 圖4在含有0.3 mg·l ⁻¹ BA之不同的基本培養基21天對辣木芽體生長之結果----- |
| xv 圖5不同張數通 氣藥紙封瓶對辣木芽體生長之影響，培養在添加0.3 mg·l ⁻¹ BA之3/2 MS培養基中21天----- | xvi 圖6以3/2MS培 養基添加0.3 mg·l ⁻¹ BA培養，辣木培植體在有無通氣處理下21天後之結果----- |
| xvii 圖7以1000 mg·l ⁻¹ 不同生長素浸漬3秒，再移入不含生長調節劑之1/2 MS培養基，21天後對辣木發根之影響。----- | xviii 圖8以1000 |

mgl-1 IBA浸漬3秒後，移入有無通氣處理之1/2 MS培養基21天後辣木培植體之結果-----xx 圖9馴化90天
後移入土壤盆栽的辣木組培苗-----xxi 表目錄 表1不同濃度之不同生長素對辣木瓶內發根之影響-----xxi 表2不同高濃度之生長素對辣木發根之影響-----xxii 表3不同張數通氣藥紙封瓶對辣木瓶內發根之影響-----xxiii 表4高濃度之不同生長素對辣木試管外發根之影響-----xx

REFERENCES

參考文獻 邢世岩。1989。木本植物組織培養玻璃化的原因和控制。國外作物的組織培養。24:88~94。陳正華。1986。木本植物組織培養及應用。北京:高等教育出版社。陳雪貞。1983。本植物組織培養之滅菌法，理工學報第二十期129-136。馮莉真。2002。聖誕紅單節培養及經由體胚之誘變育種。國立中興大學園藝系研究所碩士論文。黃曉慧。2003。網紋洋香瓜基因轉殖。大葉大學分子生物科學系研究所碩士論文。廖明芳。2003。酪梨組織培養之研究。國立中興大學園藝系研究所碩士論文。賴卓群。1996。通氣促進木瓜組織培養叢生芽之生長。國立中興大學植物學系研究所碩士論文。廖哲正。2003。台灣紅豆杉之組織培養及二次代謝物之研究。長庚大學化工與材料工程研究所碩士論文。周佳瑩、莊秉憲、陳惠民。2003。奇蹟之樹 辣木。科學發展371(11):41-45。Broin, M., Santaella, C., Cuine, S., Kokou, K., Peltier and G., Joet, T. 2002 Flocculent activity of a recombinant protein from *Moringa oleifera* Lam. Seeds, Appl. Microbiol. Biotechnol 60: 114-110. Caceres, A., Cabrera, O., Morales, O., Mollinedo. P., and Mendia, P. 1991. Pharmacological properties of *Moringa oleifera*. 1: Preliminary screening for antimicrobial activity, Journal of Ethnopharmacology 33: 213-216. Caceres, A., Cabrera, O., Morales, O., Mollinedo. P., and Mendia, P. 1991 Pharmacological properties of MORINGA OLEIFERA. 3. Effect of seed extracts in the treatment of experimental pyoderma, Fitoterapia Volume LXII 5: 449-450. Cassellati-Sforzolini G., Villarini L.M., Moretti LM, Marcarelli LM, Pasquini R, Fatigoni C, Kaur LS, Kumar S, and Grover IS. 1999 Antigenotoxic properties of Terminalia arjuna bark extracts, J Environ Pathol Toxicol Oncol.18:119-125. Dsouza, J., and Kulkarni, A.R., 1993 Comparative studies on nutritive values of tender foliage of seedlings and mature plants of *Moringa oleifera* Lam. J.Econ.Tax.Bot., 17 (2) pp479-485. Green Wood MS. 1995 Juvenility and maturation in conifers: current concepts, Tree Physiol.15:433-438. Jahn,S.A.A., 1989, *Moringa oleifera* for food and water purification - selection of clones and growing of annual short stem. Entwicklung + Landlicher Raum, 23 (4) pp22-25. Jahn,S.A.A., Musnad,H.A. and Burgstaller,H., 1986, The tree that purifies water; Cultivating multipurpose Moringaceae in the Sudan. Unasylva, 38, pp23-28. Li, L. 1990 Regeneration of bark of Chinese medicinal plants after large area girdling.Zhongguo Zhong Yao Za Zhi.15:387-389. Martin, K.P. 2003 Rapid in vitro multiplication and ex vitro rooting of *Rotula aquatica* Lour., a rare rheophytic woody medicinal plant. Plant Cell Rep. 2003: 415-420. Martin, K.P., Beena, M.R., and Joseph, D. 2003 High frequency axillary bud multiplication and ex vitro rooting of *Wedelia chinensis* (Osbeck) Merr.--a medicinal plant. Indian J Exp Biol.: 262-266. Makonnen, E., Hunde, A., and Damecha, G. 1997 Hypoglycaemic effect of *Moringa stenopetala* aqueous extract in rabbits, Phytotherapy research 11: 147-148. Nautiyal,B.P. and Venkataraman,K.G. 1987, *Moringa* (drumstick) - An ideal tree for social forestry: Growing conditions and uses - Part 1. Myforest, 23 (1) pp53-58. Nair, L.G., and Seenii, S. 2001 Rapid in vitro multiplication and restoration of *Celastrus paniculatus* Willd. sub sp. *paniculatus* (Celastraceae), a medicinal woody climber. Indian J Exp Biol.39:697-704. Nwosu, M.O., and Okafor, J.I. 1995 Preliminary studies of the antifungal activities of some medicinal plants against Basidiobolus and some other pathogenic fungi, Mycoses 38: 191-195. Saroj, K. Pal, Pulok, K. Mukherjee and B. P. Saha. 1995 Studies on the antiulcer activity of *Moringa oleifera* leaf extract on gastric ulcer models in rats, Phytotherapy research 9: 463-465. Tabuti, J.R., Lye, K.A., and Dhillon S.S. 2003 Traditional herbal drugs of Bulamogi, Uganda: plants,useandadministration.J Ethnopharmacol 88:19-44 Urkovic. J. D.2003 Regeneration of *Acer caudatifolium* Hayata plantlets from juvenile explant, Plant Cell Rep.;21:1060-1064. Valia,R.Z., Patil,V.K., Patel, Z.N. and Kapadia ,P.K.1993, Physiological responses of Drumstick (*Moringa oleifera* Lam.) to varying levels of ESP. Indian J. Plant Physiol., 36 (4) pp261-262. Went, F. W. and Thimann, K. V. 1937. Root formation In:Phytohormones (pp,183-206). MacMillan, New York. Went, F. W. 1939. The dual effect of auxin on root formation. Am J Bot 26: 24-29. Yu, T. A., Yeh, S. D., Cheng, Y. H. and Yang, J. S. 2000. Efficient rooting establishment of papaya plantlets by micropropagation. Plant Cell Tiss.Org. Cult. 61: 29-35 Yan, X. 2003 Ecological protection of medicinal woody plants. Ying Yong Sheng Tai Xue Bao.14:1561-1564. Zhiri, A., Jaziri, M., Guo, Y., Vanhaelen-Fastre, R., and Vanhaelen, M., Homes, J., Yoshimatsu, K., Shimomura. K.,1995 Tissue cultures of *Taxus baccata* as a source of 10-deacetylbaicatin III, a precursor for the hemisynthesis of taxol.Biol Chem Hoppe Seyler.376:583-6.