

# 3OST3B與CDH3基因於台灣乳癌檢體組織之甲基化變異及基因功能分析

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## ABSTRACT

The death rate of breast cancer in Taiwan has obviously risen these years, which becomes a very important issue. Many risk factors such as aberrant DNA methylation of promoter region of tumor suppress gene and growth-related gene have been reported, besides family history of breast cancer, endocrine, and carcinogen. Therefore, this study was focused on the relation between aberrant DNA methylation in gene promoter region and breast cancer tumorigenesis. Tumor and normal tissues obtained from 76 breast cancer patients were used to analyze the methylation status of promoter region and gene expression in both of cell membrane protein gene - P-cadherin (CDH3) and regulatory extracellular matrix glycoprotein gene - heparan sulfate D-glucosaminyl 3-O-sulfotransferase (3OST3B) by methylation specific PCR(MS-PCR) and combined bisulfite restriction analysis (COBRA). Because there was no aberrant methylation observed in CDH3, only 3OST3B was chosen for further analyses including COBRA- sequencing, reverse transcription PCR (RT-PCR), and immunohistochemical stain (IHC). The distribution of methylated CpG site and the expression of mRNA and protein were analyzed. According to the methylation status of 3OST3B in the tumor tissues, we divided the 76 patients into three groups - 27 patients of hypomethylation, 20 patients of intermedium-methylation and 29 patients of hypermethylation. In 3OST3B promoter region, there were many CpG dinucleotides found in the binding sites of several transcription factors such as NF-B, E2F, and n-MYC. The methylation status of CpG dinucleotides might affect the expression of 3OST3B. The results of IHC showed that the expression of 3OST3B in tumor tissues with 3OST3B hypermethylation were lower than in normal tissues. After the treatment of demethylation agent, 5-azadC, in breast cancer cell line MDA-MB-231, the methylation level of 3OST3B decreased and the gene expression was restored. The association of 3OST3B methylation status with clinicopathological features of patients with breast cancer showed that 3OST3B with intermedium-methylation and hypermethylation was significantly correlated with non-metastasis (M0) and high p53 expression. Therefore, 3OST3B could be a valuable candidate gene as a predictor of breast cancer risk assessment.

Keywords : CDH3 ; 3OST3B ; DNA Methylation ; COBRA ; breast cancer

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## REFERENCES

- 行政院衛生署九十二年度癌症死亡統計。台灣 林天祐。 1984。 中華現代外科全書（八） - 一般外科學（下）。商務書局。台灣，台北。林怡君。 2004。台灣肝癌細胞位於第十六號染色體上腫瘤抑制基因群之核酸甲基化剖析。碩士論文。國立中興大學生物醫學研究所。郭守人。 2003。乳房醫學。彰化基督教醫院。台灣，彰化。Conrad, H. E. 1998. Heparin-Binding Proteins. Academic Press, San Diego. Huxtable, R. J. 1986. Biochemistry of sulfur. New York: Plenum Publishing Corp. McLaughlin, J. R. 2001 Canadian Cancer Statistics. Canadian Cancer Society, Canada. Mitchell, S. C. 1996 Biological interactions of sulfur compounds. London: Taylor and Francis Antequera, F. and Bird, A. 1993. Number of CpG islands and genes in human and mouse. PNAS 90: 11995 – 11999. Attwood, J. T., Yung, R. L. and Richardson, B. C. 2002. DNA methylation and the regulation of gene transcription. Cell Mol. Life Sci. 59: 241-57. Avner, P. and Heard, E. 2001. X-chromosome inactivation: counting, choice and initiation. Nat. Rev. Genet. 2: 59 – 67. Basilio, C. and Moscatelli, D. 1992. The FGF family of growth factors and oncogenes. Adv. Cancer Res. 59: 115 – 165. Berx, G., Cleton-Jansen, A. M., Nollet, F., de Leeuw, W.J., van de Vijver, M.,

Cornelisse, C., and van Roy, F. 1995. E-cadherin is a tumour/invasion suppressor gene mutated in human lobularbreast cancers. *EMBO J.* 14: 6107 – 6115. Berx, G., Cleton-Jansen, A. M., Strumane, K., de Leeuw, W. J., Nollet, F., van Roy, F. and Cornelisse, C. 1996. E-cadherin is inactivated in a majority of invasive human lobular breast cancers by truncation mutations throughout its extracellular domain. *Oncogene* 13: 1919 – 1925. Bird, A. P. 1986. CpG-rich islands and the function of DNA methylation. *Nature* 321: 209-213. Blackhall, F. H., Merry, C. L., Davies, E. J. and Jayson, G. C. 2001. Heparan sulfate proteoglycans and cancer. *Br. J. Cancer* 85: 1094 – 1098. Blaschuk, O. W., Munro, S. B. and Farookhi, R. 1995. Cadherins, steroid and cancer. *Endocrine* 3: 83 – 89. Bowman, K. G. and Bertozzi, C. R. 1999. Carbohydrate sulfotransferases: mediators of extracellular communication. *Chem. Biol.* 6: R9 – R22. Bukholm, I. K., Nesland, J. M., Karesen, R., Jacobsen, U. and Borresen-Dale, A. L. 1998. E-cadherin and alpha-, beta-, and gamma-catenin protein expression in relation to metastasis in human breast carcinoma. *J. Pathol.* 185: 262 – 266. Bukholm, I. K., Nesland, J. M. and Borresen-Dale, A. L. 2000. Reexpression of E-cadherin, alpha-catenin and beta-catenin, but not of gamma-catenin, in metastatic tissue from breast cancer patients. *J. Pathol.* 190: 15 – 19. Burgess, W. H. and Maciag, T. 1989. The heparin-binding (fibroblast) growth factor family of proteins. *Annu. Rev. Biochem.* 58: 575 – 606. Carey, D. J. 1997. Syndecans: multifunctional cell-surface co-receptors. *Biochem. J.* 327:1 – 16. Catteau, A., Harris, W. H., Xu, C. F. and Solomon, E. 1999. Methylation of the BRCA1 promoter region in sporadic breast and ovarian cancer: correlation with disease characteristics. *Oncogene* 18: 1957 – 1965. Chen, C. M., Chen, H. L., Hsiau, T. H. C., Hsiau, A. H. A., Shi, H., Brock, G. J. R., Wei, S. H., Caldwell, C. W., Yan, P. S. and Huang, T. H. M. 2003. Methylation target array for rapid analysis of CpG island hypermethylation in multiple genomes. *Am. J. Pathol.* 163: 37-45. Clark, S. J., Harrison, J. and Molloy, P. L. 1997. Sp1 binding is inhibited by (m)Cp(m)CpG methylation. *Gene* 195: 67 – 71. Comb, M. and Goodman, H. M. 1990. CpG methylation inhibits proenkephalin gene expression and binding of the transcription factor AP-2. *Nucleic Acids Res.* 18: 3975 – 3982. Creusot, F., Acs, G. and Christman, J. K. 1982. Inhibition of DNA methyltransferase and induction of Friend erythroleukemia cell differentiation by 5-azacytidine and 5-aza-2'-deoxycytidine. *J. Biol. Chem.* 257: 2041-2048. Daniel, C. W., Strickland, P. and Friedmann, Y. 1995. Expression and functional role of E- and P-cadherins in mouse mammary ductal morphogenesis and growth. *Dev. Biol.* 169: 511 – 519. Davidson, S., Crowther, P., Radley, J. and Woodcock, D. 1992. Cytotoxicity of 5-aza-2'-deoxycytidine in a mammalian cell system. *Eur. J. Cancer* 28: 362-368. Davidson, N. E. 2000. Combined endocrine therapy for breast cancer – new life for an old idea? *Journal of the National Cancer Institute* 92: 859 – 860. Dixon J. M. 1985. Long-term survivors after breast cancer. *Br. J. Surge.* 72: 445. Dobrovic, A. and Simpfendorfer, D. 1997. Methylation of the BRCA1 gene in sporadic breast cancer. *Cancer Res.* 57: 3347 – 3350. Ehrlich, M. and Wang, R. Y. 1981. 5-Methylcytosine in eukaryotic DNA. *Science* 212: 1350 – 1357. Esteller, M. 2002. CpG island hypermethylation and tumor suppressor genes: a booming present, a brighter future. *Oncogene.* 21: 5427 – 5440. Esteller, M., Silva, J. M., Dominguez, G., Bonilla, F., Matias-Guiu, X., Lerma, E., Bussaglia, E., Prat, J., Harkes, I. C., Repasky, E. A., Gabrielson, E., Schutte, M., Baylin, S. B. and Herman, J. G. 2000 Promoter hypermethylation and BRCA1 inactivation in sporadic breast and ovarian tumors. *Journal of the National Cancer Institute* 92: 564 – 569. Ferguson, A. T., Lapidus, R. G. and Davidson, N. E. 1998. Demethylation of the progesterone receptor CpG island is not required for progesterone receptor gene expression. *Oncogene* 17: 577 – 583. Filmus, J. 2001. Glypicans in growth control and cancer. *Glycobiology* 11: 19R – 23R. Fishman, J., Osborne, M. P. and Telang, N. T. 1995. The role of estrogen in mammary carcinogenesis. *Annals of the New York Academy of Sciences* 768: 91 – 100. Flatau, E., Gonzales, F.. A., Michalowsky, L. A. and Jones, P. A. 1984. DNA methylation in 5-aza-2'-deoxycytidine-resistant variants of C3H 10T1/2 C18 cells. *Mol. Cell Biol.* 4: 2098-2102. Friedman, L. S., Ostermeyer, E. A., Szabo, C. I., Dowd, P., Lynch, E. D., Rowell, S. E. and King, M. C. 1994. Confirmation of BRCA1 by analysis of germline mutations linked to breast and ovarian cancer in ten families. *Nat. Genet.* 8: 399 – 404. Gallagher, J. T. 1994. Heparan sulphates as membrane receptors for the fibroblast growth factors. *Eur. J. Clin. Chem. Clin. Biochem.* 32: 239 – 247. Gamallo, C., Palacios, J. and Suarez, A. 1993. Correlation of Ecadherin expression with differentiation grade and histological type in breast carcinoma. *Am. J. Pathol.* 142: 987 – 993. Gardiner-Garden, M. and Formmer, M. 1987. CpG islands in vertebrate genomes. *J Mol Biol.* 196: 261-282. Gonzalez, M. A., Pinder, S. E., Wencyk, P. M., Bell, J. A., Elston, C. W., Nicholson, R. I., Robertson J. F. R., Blamey, R. W. and Ellis, I. O. 1999. An immunohistochemical examination of the expression of E- cadherin, - / - catenins, and 2- and 1- integrins in invasive breast cancer. *J. Pathol.* 187: 523-529. Graham, J. R., Huang, T. H. C., Chen, C. M. and Johnson, K. J. 2001. A novel technique for the identification of CpG islands exhibiting altered Methylation patterns ( ICEAMP ) . *Nucleic Acids Res.* 29: 123-130. Grunwald, G. B. 1993. The structural and functional analysis of cadherin calcium-dependent cell adhesion molecules. *Curr. Opin. Cell Biol.* 5: 797 – 805. Haaf, T. and Schmid, M. 1989. 5-Azadeoxycytidine induced undercondensation in the giant X chromosomes of *Microtus agrestis*.*Chromosoma* 98: 93-98. Han, A. C., Peralta-Soler, A., Knudsen, K. A., Wheelock, M. J., Johnson, K. R. and Salazar, H. 1997. Differential expression of N-cadherin in pleural mesotheliomas and E-cadherin in lung adenocarcinomas in formalin-fixed, paraffin-embedded tissues. *Hum. Pathol.* 28: 641 – 645. Han, A. C., Soler, A. P., Knudsen, K. A. and Salazar, H. 1999. Distinct cadherin profiles in special variant carcinomas and other tumors of the breast. *Hum. Pathol.* 30: 1035 – 1039. Han, A. C., Soler, A. P., Tang, C. K., Knudsen, K. A. and Salazar, H. 2000. Nuclear localization of E-cadherin expression in Merkel cell carcinoma. *Arch Pathol. Lab. Med.* 124: 1147 – 1151. Hazan, R. B., Phillips, G. R., Qiao, R. F., Norton, L. and Aaronson, S.A. 2000. Exogenous expression of N-cadherin in breast cancer cells induces cell migration, invasion, and metastasis. *J. Cell Biol.* 148: 779 – 790. Heard, E., Clerc, P. and Avner, P. 1997. X-chromosome inactivation in mammals. *Annu. Rev. Genet.* 31: 571 – 610. Hiraguri, S., Godfrey, T., Nakamura, H., Graff, J., Collins, C. and Shayesteh, L. 1998. Mechanisms of inactivation of E-cadherin in breast cancer cell lines. *Cancer Res.* 58: 1972 – 1977. Holland, R., Peterse, J. L., Millis, R. R., Eusebi, V., Faverly, D., van de Vijver, M. J. and Zafrani, B. 1994. Ductal carcinoma in situ: a proposal for a new classification. *Semin Diagn Pathol.* 11: 167 – 180. Holliday, R. and Pugh, J. E. 1975. DNA modification

mechanisms and gene activity during development. *Science*. 187: 226–232. Hooper, L. V., Manzella, S. M. and Baenziger, J. U. 1996. From legumes to leukocytes: biological roles for sulfated carbohydrates. *FASEB J.* 10: 1137 – 1146. Hortobagyi, G. N. 1998. Treatment of breast cancer. *New England J. of Med.* 339: 974 – 984. Iguchi-Ariga, S. M. and Schaffner, W. 1989. CpG methylation of the cAMP-responsive enhancer/promoter sequence TGACGTCA abolishes specific factor binding as well as transcriptional activation. *Genes Dev.* 3: 612 – 619. Jablonka, E., Goitein, R., Marcus, M. and Cedar, H. 1985. DNA hypomethylation causes an increase in DNase-I sensitivity and an advance in the time of replication of the entire inactive X chromosome. *Chromosoma* 93: 152-156. Jaenisch, R., Schnieke, A. and Harbers, K. 1985. Treatment of mice with 5-azacytidine efficiently activates silent retroviral genomes in different tissues. *Proc. Natl. Acad. Sci. USA*. 82: 1451-1455. Jarrard, D. F., Paul, R., van Bokhoven, A., Nguyen, S. H., Bova, G. S. and Wheelock, M. J. 1997. P-cadherin is a basal cell-specific epithelial marker that is not expressed in prostate cancer. *Clin. Cancer Res.* 3: 221 – 228. Jayson, G. C. et al. 1998. Heparan sulfate undergoes specific structural changes during the progression from human colon adenoma to carcinoma in vitro. *J. Biol. Chem.* 273: 51 – 57. Joana p., Fernanda, M., Leda, V., Isabel, A. And Fernando, S. 2002. P- cadherin expression is associate with high-grade ductal carcinoma in situ of the breast. *Virch. Arch* 440: 16-21. Jones, P. A., Taylor, S. M., Mohandas, T. and Shapiro, L. J. 1982. Cell cycle-specific reactivation of an inactive X-chromosome locus by 5-azadeoxycytidine. *Proc. Natl. Acad. Sci. USA*. 79:1215-1219. Jones, P. A., Taylor, S. M. and Wilson, V. 1983. DNA modification, differentiation, and transformation. *J. Exp. Zool.* 228: 287-295. Jones, P. A. 1985. Altering gene expression with 5-azacytidine. *Cell*. 40: 485-6. Jones, P. A. and Laird, P. W. 1999. Cancer epigenetics comes of age. *Nat. Genet.* 21: 163-167. Knudsen, K. A., Myers, L. and McElwee, S. A. 1990. A role for the Ca<sup>2+</sup>-dependent adhesion molecule, N-cadherin, in myoblast interaction during myogenesis. *Exp. Cell Res.* 188: 175 – 184. Kobayashi, S., Morimoto, K., Shimizu, T., Takahashi, M., Kurosawa, H. and Shirasawa, T. 2000. Association of EXT1 and EXT2, hereditary multiple exostoses gene products, in Golgi apparatus. *Biochem. Biophys. Res. Commun.* 268: 860-867 Kolset, S. O. and Salmivirta, M. 1999. Cell surface heparan sulfate proteoglycans and lipoprotein metabolism. *Cell Mol. Life Sci.* 56: 857 – 870 Kreuger, J., Salmivirta, M., Sturiale, L., Gimenez-Gallego, G. and Lindahl, U. 2001. Sequence analysis of heparan sulfate epitopes with graded affinities for fibroblast growth factors 1 and 2. *J. Biol. Chem.* 276: 30744 – 30752. Lapidus, R. G., Nass, S. J., Butash, K. A., Parl, F. F., Weitzman, S. A., Graff, J. G., Herman, J. G. and Davidson, N. E. 1998. Mapping of ER gene CpG island methylation-speci.c polymerase chain reaction. *Cancer Res.* 58: 2515 – 2519. Larsen, F., Gundersen, G., Lopez, R. and Prydz, H. 1992. CpG islands as gene markers in the human genome. *Genomics* 13:1095-1107. Larue, L., Antos, C., Butz, S., Huber, O., Delmas, V., Dominis, M. and Kemler, R. 1996. A role for cadherins in tissue formation. *Dev.* 122: 3185 – 3194. Leal, C. B., Schmitt, F. C., Bento, M. J., Maia, N. C. and Lopes, C. S. 1995. Ductal carcinoma in situ of the breast. *Cancer* 75: 2123 – 2131. Lee, J. T. and Jaenisch, R. 1997. The (epi)genetic control of mammalian X-chromosome inactivation. *Curr. Opin. Genet. Dev.* 7: 274 – 280. Li, E. 2002. Chromatin modification and epigenetic reprogramming in mammalian development. *Nat. Genet.* 3: 662-673. Lindahl, U., Kusche-Gullberg, M. and Kjellen, L. 1998. Regulated diversity of heparan sulfate. *J. Biol. Chem.* 273: 24979-24982. Liu, D., Shriver, Z., Qi, Y., Venkataraman, G. and Sasisekharan, R. 2002. Dynamic regulation of tumor growth and metastasis by heparan sulfate glycosaminoglycans. *Semin. Thromb. Hemost.* 28: 67 – 78. Loo, B. M., Kreuger, J., Jalkanen, M., Lindahl, U. and Salmivirta, M. 2001. Binding of heparin/heparan sulfate to fibroblast growth factor receptor 4. *J. Biol. Chem.* 276: 16868 – 16876. Magdinier, F., Billard, L. M., Wittmann, G., Frappart, L., Benchaib, M., Lenoir, G. M., Guerin, J. F. and Dante, R. 2000. Regional methylation of the 5' end CpG island of BRCA1 is associated with reduced gene expression in human somatic cells. *FASEB J.* 14: 1585 – 1594. Marino, M., Andrews, D., and McCluskey, R. T. 2000. Binding of rat thyroglobulin to heparan sulfate proteoglycans. *Thyroid*. 10: 551 – 559. McCormick, C., Duncan, G., Goutsos, K. T. and Tufaro, F. 2000. The putative tumor suppressors EXT1 and EXT2 form a stable complex that accumulates in the Golgi apparatus and catalyzes the synthesis of heparan sulfate. *Proc. Natl. Acad. Sci. USA* 97: 668-673. Merajver, S. D., Pham, T. M., Caduff, R. F., Chen, M., Poy, E. L., Cooney, K. A., Weber, B. L., Collins, F. S., Johnston, C. and Frank, T. S. 1995. Somatic mutations in the BRCA1 gene in sporadic ovarian tumours. *Nat. Genet.* 9: 439 – 443. Michalowsky, L. A. and Jones, P. A. 1987. Differential nuclear protein binding to 5-azacytosine-containing DNA as a potential mechanism for 5-aza-2'-deoxycytidine resistance. *Mol. Cell Biol.* 7: 3076-3083. Michalowsky, L. A. and Jones, P. A. 1987. Differential nuclear protein binding to 5-azacytosine-containing DNA as a potential mechanism for 5-aza-2'-deoxycytidine resistance. *Mol. Cell Biol.* 7: 3076-3083. Moll, R., Mitze, M., Frixen, U. H. and Birchmeier, W. 1993. Differential loss of E-cadherin expression in infiltrating ductal and lobular breast carcinomas. *Am. J. Pathol.* 143: 1731 – 1742. Momparler, R. L., Bouchard, J. and Samson, J. 1985. Induction of differentiation and inhibition of DNA methylation in HL-60 myeloid leukemic cells by 5-AZA-2'-deoxycytidine. *Leuk. Res.* 9: 1361-1366. Mundhenke, C., Meyer, K., Drew, S. and Friedl, A. 2002. Heparan sulfate proteoglycans as regulators of fibroblast growth factor-2 receptor binding in breast carcinomas. *Am. J. Pathol.* 160: 185 – 194. Muramatsu, T. 2000. Essential roles of carbohydrate signals in development, immune response and tissue functions, as revealed by gene targeting. *J. Biochem.* 127: 171 – 176 Nan, X., H. H. Ng, C. A. Johnson, C. D. Laherty, B. M. Turner, R. N. Eisenman, and A. Bird. 1998. Transcriptional repression by the methyl-CpG-binding protein MeCP2 involves a histone deacetylase complex. *Nature*. 329: 451-454. Ng, H. H. and A. Bird. 2000. Histone deacetylases: silencers for hire. *Trends Biochem. Sci.* 25: 121-126. Nieman, M. T., Prudoff, R. S., Johnson, K. R. and Wheelock, M. J. 1999. N-cadherin promotes motility in human breast cancer cells regardless of their E-cadherin expression. *J. Cell Biol.* 147: 631 – 643. Ottaviano, Y. L., Issa, J. P., Parl, F. F., Smith, H. S., Baylin, S. B. and Davidson, N. E. 1994. Methylation of the estrogen receptor gene CpG island marks loss of estrogen receptor expression in human breast cancer cells. *Cancer Res.* 54: 2552 – 2555. Palacios, J., Benito, N., Pizarro, A., Suarez, A., Espada, J., Cano, A. and Gamallo, C. 1995. Anomalous expression of P-cadherin in breast carcinoma. Correlation with E-cadherin expression and pathological features. *Am. J. Pathol.* 146: 605 – 612. Paulsen, M., El-Maarri, O., Engemann, S., Strodicke, M., Franck, O., Davies, K., Reinhardt,

R., Reik, W. and Walter, J. 2000. Sequence conservation and variability of imprinting in the Beckwith-Wiedemann syndrome gene cluster in human and mouse. *Hum. Mol. Genet.* 9: 1829 – 1841. Peralta Soler, A., Knudsen, K. A., Jaurand, M. C., Johnson, K. R., Wheelock, M. J., Klein-Szanto, A. J. and Salazar, H. 1995. The differential expression of N-cadherin and E-cadherin distinguishes pleural mesotheliomas from lung adenocarcinomas. *Hum. Pathol.* 26: 1363 – 1369. Peralta Soler, A., Harner, G. D., Knudsen, K. A., McBearty, F. X., Grujic, E. and Salazar, H. 1997. The expression of P-cadherin identifies PSA negative cells in epithelial tissues of male sexual accessory organs and in prostatic carcinomas. Implications for prostate cancer biology. *Am. J. Pathol.* 151: 471 – 478. Peralta Soler, A., Knudsen, K. A., Salazar, H., Han, A. C. and Keshgegian, A. A. 1999. P-cadherin expression in breast carcinoma indicates poor survival. *Cancer* 86: 1263 – 1272. Pereira, L. V. and Vasques, L. R. 2000. X-chromosome inactivation: lessons from transgenic mice. *Gene* 255: 363 – 337. Perrimon, N. and Bernfield, M. 2000. Specificities of heparin sulphate proteoglycans in developmental processes. *Nature* 404: 725 – 728. Pinto, A. and Zagonel, V. 1993. 5-Aza-2'-deoxycytidine (Decitabine) and 5-azacytidine in the treatment of acute myeloid leukemias and myelodysplastic syndromes: past, present and future trends. *Leukemia* 1: 51-60. Pizarro, A., Gamallo, C., Benito, N., Palacios, J., Quintanilla, M. and Cano, A. 1995. Differential patterns of placental and epithelial cadherin expression in basal cell carcinoma and in the epidermis overlying tumors. *Br. J. Cancer* 72: 327 – 332. Plotnikov, A. N., Schlessinger, J., Hubbard, S. R. and Mohammadi, M. 1999. Structural basis for FGF receptor dimerization and activation. *Cell* 98:641 – 650 Prendergast, G. C. and Ziff, E. B. 1991. Methylation-sensitive sequence-specific DNA binding by the c-Myc basic region. *Science* 251: 186 – 189. Radice, G. L., Ferreira-Cornwell, M. C., Robinson, S. D., Rayburn, H., Chodosh, L. A., Takeichi, M. 1997. Precocious mammary gland development in P-cadherin-deficient mice. *J. Cell Biol.* 139: 1025 – 1032. Rasbridge, S. A., Gillett, C. E., Sampson, S. A., Walsh, F. S. and Millis, R. R. 1993. Epithelial (E-) and placental (P-) cadherin cell adhesion molecule expression in breast carcinoma. *J. Pathol.* 169: 245 – 250. Razin A. and Riggs A. D. 1980. DNA methylation and gene function. *Science*. 210: 604 – 610. Redies, C., Engelhart, K. and Takeichi, M. 1993. Differential expression of N- and R-cadherin in functional neuronal systems and other structures of the developing chicken brain. *J. Comp. Neurol.* 333: 398 – 416. Richel, D. J., Colly, L. P., Lurvink, E. and Willemze, R. 1988. Comparison of the antileukaemic activity of 5 aza-2-deoxycytidine and arabinofuranosyl-cytosine in rats with myelocytic leukaemia. *Br. J. Cancer* 58: 730-733. Riggs, A. D. 1975. X inactivation, differentiation, and DNA methylation. *Cytogenet Cell Genet.* 14: 9-25. Ringwald, M., Baribault, H., Schmidt, C. and Kemler, R. 1991. The structure of the gene coding for the mouse cell adhesion molecule uvomorulin. *Nucleic Acids Res.* 19: 6533 – 6539. Rosenberg, R. D., Shworak, N. W., Liu, J., Schwartz, J. J. and Zhang, L. 1997. Heparan sulfate proteoglycans of the cardiovascular system. Specific structures emerge but how is synthesis regulated? *J. Clin. Invest.* 100:S67-S75. Ruiz-Cabello, J., Berghmans, K., Kaplan, O., Lippman, M. E., Clarke, R. and Cohen, J. S. 1995. Hormone dependence of breast cancer cells and the effects of tamoxifen and estrogen: 31P NMR studies. *Breast Cancer Res. and Treat.* 33: 209 – 217. Sanders, D. S. A., Perry, I., Hardy, R. and Jankowski, J. 2000. Aberrant P-cadherin expression is a feature of clonal expansion in the gastrointestinal tract associated with repair and neoplasia. *J. Pathol.* 190: 526 – 530. Sanderson, R. D. 2001. Heparan sulfate proteoglycans in invasion and metastasis. *Semin. Cell Dev. Biol.* 12: 89 – 98. Salmivirta, M., Lidholt, K. and Lindahl, U. 1996 Heparan sulfate: a piece of information. *FASEB J.* 10: 1270 – 1279 Schmitt, F. C., Figueiredo, P. and Lacerda, M. 1995. Expression of cerbB- 2 protein and DNA ploidy in breast carcinogenesis. *Arch. Pathol. Lab. Med.* 119: 815 – 820. Schmutte, C. and Fishel, R. 1999. Genomic instability:first step to carcinogenesis. *Anticancer Res.* 19: 4665 – 4696. Shimoyama, Y., Hirohashi, S., Hirano, S., Noguchi, M., Shimosato, Y., Takeichi, M. and Abe, O. 1989. Cadherin cell-adhesion molecules in human epithelial tissues and carcinomas. *Cancer Res.* 49: 2128 – 2133. Shimoyama, Y. and Hiroshasi, S. 1991. Expression of E- and P-cadherin in gastric carcinomas. *Cancer Res.* 51: 2185 – 2192. Shworak, N. W., Liu, J., Petros, L. M., Zhang, L., Kobayashi, M., Copeland, N. G., Jenkins, N. A. and Rosenberg, R. D. 1999. Multiple isoforms of heparan sulfate D-glucosaminyl 3-O-sulfotransferase. Isolation, characterization, and expression of human cdnas and identification of distinct genomic loci. *J. Biol. Chem.* 274: 5170 – 5184. Siitonen, S. M., Kononen, J. T., Helin, H. J., Rantala, I. S., Holli, K. A. and Isola, J. J. 1996. Reduced E-cadherin expression is associated with invasiveness and unfavorable prognosis in breast cancer. *Am. J. Clin. Pathol.* 105: 394 – 402. Silverstein, M. J., Lagios, M. D., Craig, P. H., Waisman, J. R., Lewinsky, B. S., Colburn, W. J. and Poller, D. N. 1996. A prognostic index for ductal carcinoma in situ of the breast. *Cancer* 77: 2267 – 2274. Soler, A. P. and Knudsen, K. A. 1994. N-cadherin involvement in cardiac myocyte interaction and myofibrillogenesis. *Dev. Biol.* 162: 9 – 17. Soler, A. P., Knudsen, K.A., Tecson-Miguel, A., McBrearty, F. X., Han, A. C. and Salazar, H. 1997. Expression of E-cadherin and Ncadherin in surface epithelial-stromal tumors of the ovary distinguishes mucinous from serous and endometrioid tumors. *Hum. Pathol.* 28: 734 – 739. Takeichi, M. 1990. Cadherins: a molecular family important in selective cell-cell adhesion. *Annu. Rev. Biochem.* 59: 237 – 252. Tan, D. S., Potts, H. W., Leong, A. C., Gillett, C. E., Skilton, D., Harris, W. H., Liebmann, R. D. and Hanby, A. M. 1999. The biological and prognostic significance of cell polarity and E-cadherin in grade I infiltrating ductal carcinoma of the breast. *J. Pathol.* 189: 20 – 27. Ullrich, A. and Schlessinger, J. 1990. Signal transduction by receptors with tyrosine kinase activity. *Cell* 61:203 – 212 Varki, N. M. and Varki, A. 2002. Heparin inhibition of selectinmediated interactions during the hematogenous phase of carcinoma metastasis: rationale for clinical studies in humans. *Semin. Thromb. Hemost.* 28, 53 – 66. Vlodavsky, I., Miao, H. Q., Medalion, B., Danagher, P. and Ron, D. 1996. Involvement of heparan sulfate and related molecules in sequestration and growth promoting activity of fibroblast growth factor. *Cancer Metastasis Rev.* 15:177 – 186 Wei, G., Bai, X., Bame, K. J., Koshy, T. I., Spear, P. G. and Esko, J. D. 2000. Location of the glucuronosyltransferase domain in the heparan sulfate copolymerase EXT1 by analysis of Chinese hamster ovary cell mutants. *J. Biol. Chem* 275:27733-27740. Wheelock, M. J. and Knudsen, K. A. 1991. Cadherins and associated proteins. In. *Vivo.* 505 – 513. Wilson, V. L., Jones, P. A. and Momparler, R. L. 1983. Inhibition of DNA methylation in L1210 leukemic cells by 5-aza-2'-deoxycytidine as a possible mechanism of chemotherapeutic action. *Cancer Res.* 43:

3493-3496. Wollnerl, D. A., Krzeminski, K. A. and Nelson, W. J. 1992. Remodelling the cell surface distribution of membrane proteins during the development of epithelial cell polarity. *J. Cell Biol.* 116: 889 – 899. Woodward, W. A., Strom, E. A., Tucker, S. L., McNeese, M. D., Perkins, G. H., Schechter, N. R., Singletary, S. E., Theriault, R. L., Hortobagyi, G. N., Hunt, K. K. and Buchholz, T. A. 2003. Changes in the 2003 American Joint Committee on cancer staging for breast cancer dramatically affect stage-specific survival. *J. Clin. Oncology* 17: 3244-3248. Xiong, Z. and Laird, P. W. 1997. COBRA: a sensitive and quantitative DNA Methylation assay. *Nucleic Acids Res.* 25: 2532-2534. Yasui, Y., Sano, T., Nishimura, Y., Kitadi, Y., Ji, Z. Q. and Yokozaki, H. 1993. Expression of P-cadherin in gastric carcinoma and its reduction in tumor progression. *Int. J. Cancer* 54: 49 – 52. Yu, F., Thiesen, J. and Stratling, W. H. 2000. Histone deacetylase-independent transcriptional repression by methyl-CpG binding protein 2. *Nucleic Acids Res.* 28: 2201 – 2206. Zimmermann, P. and David, G. 1999. The syndecans, tuners of transmembrane signaling. *FASEB J.* 13: S91 – S10