

Development of Flower Pigment Transgenic Chrysanthemum by Meristem-tip Culture and Agrobacterium Transformation

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ABSTRACT

Since meristem-tip culture has the fully ability to differentiate into plants without changing the original inherency and can produce a large amount of explants in a short time, it has been widely used for plant micropropagation. In this study, meristem-tip culture were used to obtain chrysanthemum explants for gene transformation. In order to observe the influence on the expression of flower color, two pigment genes, CHI and DFR, were used for transformation of chrysanthemum explants. Chrysanthemum plants from field were first sterilized and meristem-tips were cut and cultured in appropriate media for fifteen days to two months. Since the size of the cutting affect the survival rate, only 39% in 410 meristem-tip cultures were obtained and used for mass production. In addition, chrysanthemum from the field and meristem-tip culture were detected by RT-PCR using primers specific to Chrysanthemum stunt viroid, it was found that none of them was viroid-free. In the regeneration experiments, two chrysanthemum varieties, Huang-shin-Fang and Bai-Dong-Yang, showed that the best regeneration rate was in the MS medium containing 1 mg/l BA and 0.5 mg/l NAA. For Huang-shin-Fang, all the explants had the ability to regenerate into adventitious shoots, and the average regeneration number of each explant was 6.3. For Bai-Dong-Yang, thirty six out of forty eight explants were able to regenerate and each explant produced about 2.7 adventitious shoots. Hygromycin was used for the selection of transformed plants. From the hygromycin sensitivity test on non-transgenic chrysanthemum, it was found that 10 mg/l of hygromycin was high enough to stop the regeneration of chrysanthemum. Therefore, the transformation condition was performed by using MS medium with 1 mg/l BA, 0.5 mg/l NAA, and 10 mg/l of hygromycin. The suitable co-culture time for chrysanthemum and Agrobacterium is four days. There were 25 putative CHI transgenic chrysanthemum obtained from 109 explants. When the putative CHI transgenic plants were further analyzed by PCR amplification coupled with Southern blotting, nine of them showed positive reaction. Similarly, thirteen out of eighty three explants were able to regenerate from selection medium after DFR gene transformation, and 9 of them were further verified by PCR amplification coupled with Southern blotting. To further study the insertion number of the DFR transgenic plants, genomic Southern blotting were carried out. It was found that the DFR transgenic chrysanthemum all contained one transgene insertion. The transgenic lines are now kept in greenhouse for the observation of the flower pigment expression.

Keywords : meristem-tip culture, Chrysanthemum stunt viroid, RT-PCR, Agrobacterium transformation, co-culture, Southern blot.

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REFERENCES

- 王強生, 2002年, 利用基因工程技術圓一個古老的夢(創造新花色), 科學發展期刊, 351期。
- 行政院農業委員會, 2001年, 農業統計年報, 行政院農業委員會。
- 邱輝龍、范明仁 1998 花青素與花色之表現.中國園藝44:102-105。
- 張淑芬, 菊花微體繁殖之研究, 國立中興大學園藝學研究所碩士論文, 中華民國83年6月。
- 黃敏展. 1990. 花卉園藝栽培技術. 行政院青年輔導委員會PP:70-82。
- 藤野守弘、藤岡作太郎、藤村良. 1973. 莖頂培養之增殖研究報告. 兵庫縣立農業試驗所試驗 研究報告 第20號別刷:125-131。
- Aida, R., Kishimoto, S., Tanaka, Y., and Shibata, M. 2000. Modification of flower colour in torenia (*Torenia fournieri* Lind.) by genetic transformation. *Plant Sci* 153: 33-42.
- Ben, J. J., and Langhans, R. W. 1972. Rapid multiplication of Chrysanthemum plant by stem-tip proliferation. *Hort. Sci* 7(3):289-290.
- Broertjes, C. and Keen, A. 1980. Adventitious shoots: Do they develop from one cell. *Euphytica* 29: 73-87.
- Bush, S. R., Earle, E. D. and Langhans, R. W. 1976. Plantlets from petal segments, petal epidermis, and shoot tips of the pericinal chimera, chrysanthemum morifolium " indianapolis ." *Amer. J. Bot* 63(6):729-737.
- Dixon, R.A., and Steele, Y.A. 1999. The phytoalexin response : Elicitation, signaling, and control of host gene expression. *Biol. Rev* 61: 239-291.
- Earle, E. D., and Langhans, R. W. 1974. Propagation of Chrysanthemum in vitro: . Production, growth. And flowering of plantlets from tissue cultures. *J. Amer. Soc. Hort. Sci* 99(4):352-358.
- Evans, D. A., Willan, R. S., and Cristopher, E. F. 1981. Growth and behavior of cell culture. In " Plant tissue culture method and application in Agriculture ." Academic press, New York. pp 45-113.
- Evans, D. A. and Bravo, J. E. 1986. Phenotypic and genotypic stability of tissue cultured plants. In " Tissue Culture as a Plant Propagation System for Horticultural crops," edited by Zimmerman, R. H. , Griesbach, R. J., Hammerschlag F. A., and Lawson, R. H. Martinus Nijhoff Publishers. pp 73-94.
- Faust, J. E. and Heins, R. D. 1992. High night temperatures do not cause poor lateral branching of chrysanthemum. *Hort Sci* 27(9):981-982.
- Fulton, T. M., Chunwongse, J., and Tanksley, S. D. 1995 Microprep Protocol for Extraction of DNA from Tomato and other Herbaceous Plants. *Plant Molecular Biology Reporter* 13: 207-209.
- George, E. F. 1993. Plant propagation by tissue culture. *Preslia, Praha* 51: 213-237.
- George, E. F., and Sherrington, P. D. 1984. Plant propagation by tissue culture. Exegetics Ltd., Eversley, Basinstoke, Heres 709p.
- Hartmann, H. T. . Kester, D. E and Davies, F. T. 1990. Propagation of selected annuals and perennials used as ornamentals. In:Plant Propagation pp:609-633.
- Helariutta, Y., Elomaa, P., Kotilainen, M., Seppanen, P., and Teerl, T. H. 1993. Cloning of cDNA coding for dihydroflavonol-4-reductase (DFR) and characterization of dfr expression in the corollase of *Gerbera hybrida* war. Regina (composites) . *Plant Mol. Biol* 22: 183-193.
- Hirabayashi, T. T., Moriguchi, I., Kozaki, Y., and Yamamoto, T. 1987. In vitro propagation of Pyrus shoot tips. *Bull. Fruit Tree Res. Stn. Japan A.* 14: 9-16.
- Hollings, M. and Stone, O. M. 1970. Attempts to eliminate Chrysanthemum stunt from chrysanthemum by meristem-tip culture after heat treatment. *Ann. Appl. Biol* 65: 311-315.
- Horst, R. K. and Cohen, D. 1980. Amantadine supplemented tissue culture medium: A method for obtaining chrysanthemum free of Chrysanthemum stunt viroid. *Acta Hort* 110: 315-320.
- Khalid, N., Davey, M. R., and Power, J. B. 1989. An assesment of somaconal variation in chrysanthemum morifolium: the generation of plants of potential commerical value. *Scientia Hort.* 38: 287-294.
- Kofranek, A. M. 1992. Cut chrysanthemum. Introduction to Floriculture, Larson, R. A. Academic Press, New York. pp:3-35.
- Kudo, S., Shibata, N., Kanno, Y., and Suzuki, M. 2002. Transformation of chrysanthemum (*Dendranthema grandiflorum* (Ramat.) kitamura) via *Agrobacterium tumefaciens*. *Acta Hort* 572: 139-147.
- Lu, C. Y., Nugent, T., and Dalling, J. M. 1991. *Agrobacterium*-mediated transformation of carnation (*Dianthus caryophyllus* L.) *Biotechnology* 9: 864-868.
- Mercuri, A., Sacchetti, A., Benedetti, D. L., Schiva, T., and Alberti, S. 2002 Green fluorescent flower. *Plant Sci* 162:647-654.
- Meyer, P., Heidmann, I., Forkmann, G., Saedler, H. 1987. A new petunia flower colour generated by transformation of a mutant with a maize gene. *Nature* 330: 677-678.
- Mitiouchkina, T. Yu., Ivanova. E. P., Taran, S. A. and Dolgov, S. V. 2000. Chalcone synthase gene from *antirrhinum majus* in antisense orientation successfully suppressed the petals pigmentation of chrysanthemum. *Acta Hort* 508: 215-217.
- Mol, J., Grotewold, E. and Koes, R. 1998. How genes paint flowers and seeds. *Trends Plant Sci* 3: 212-217.
- Napoli, C., Lemieux, C., and Jorgensen, R. 1990. Introduction of a Chimeric Chalcone Synthase Gene into Petunia Results in Reversible Co-Suppression of Homologous Genes in trans. *Plant Cell* 2: 279-289.
- Paludan, N. 1985. Inactivation of viroids in chrysanthemum by low-temperature treatment and meristem-tip culture. *Acta Hort* 164: 181-186.
- Renou, P. J., Brochard, P., and Jalouzot, R. 1993. Recovery of transgenic chrysanthemum (*Dendranthema grandiflora* Tzvelev) after hygromycin resistance selection. *Plant sci* 89:185-197.
- Sangwan, R. S. and Harada, H. 1977. Cellular totipotency in chrysanthemum tissue cultured in vitro. *Acta Hort.* 78: 237-242.
- Swartz, H. J. 1991. Post culture behavior: genetic and epigenetic effects and related problems. In: micropropagation. Edited by Debergh, P. C. and Zimmerman, R. H. pp 95-121.
- Tanaka, Y., Fukui, Y., Fukuchi, M., M., Holton, T. , Higgins, E., and Kusumi, T. 1995. Molecular cloning and characterization of *Rosa Hybrida* dihydroflavonol 4-reductase gene. *Plant Cell Physiol* 36: 1025-1031.
- Yang, C.L., Su, J. C., and Wu, J. L. 1977. Purification and properties of sucrose synthase from the shoot of bamboo *Leba oldhami*. *Plant Physiol* 60: 17-21.
- Yepes, L. M., Mittak, V., Pang, S. Z., Gonsalves, C., Slightom, J. L., and Gonsalves, D. 1995. Biolistic

transformation of chrysanthemum with the nucleocapsid gene of tomato spotted wilt virus. *Plant Cell Reports* 14: 694-698.