

利用莖頂培養與農桿菌基因轉殖方式發展轉花色基因之菊花

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摘要

由於植物莖頂培養具有高度的分化全能性，遺傳性狀穩定且可使培植體快速增殖，因此本實驗乃利用莖頂培養技術生產無變異之菊花種苗，來提供基因轉殖材料。另外亦將CHI及DFR花色基因轉殖至菊花中，以觀察其對菊花花色表現的影響。由田間採得之菊花植株，經次氯酸鈉消毒後，切取其莖頂進行紙橋培養，約15天至2個月左右可得菊花芽體，再移植至固態MS培養基中進行大量繁殖。由於所切莖頂大小會影響存活率，本實驗共得158無菌菊花苗，存活率為39%。此外，由田間採集來及莖頂培養所得之菊花植株，利用反轉錄聚合酶連鎖反應檢測菊花矮化類病毒（Chrysanthemum stunt viroid, CSVd），均發現有CSVd的感染。菊花轉殖前的再生培養基，以MS加入1 mg/l BA及0.5 mg/l NAA的條件對黃秀芳及白東陽兩種品種的菊花葉片的再生能力最佳，28個黃秀芳培植體均可全部再生，平均每個培植體可再生出6.3株不定芽；48個白東陽培植體中有36個培植體有再生能力，平均每個培植體可再生出2.7株不定芽。而在菊花對hygromycine最低致死濃度的測試中發現，10 mg/l的濃度即可使葉片停止再生，並於第28天全部死亡，因此目前所進行之轉殖實驗即以基礎MS培養基含1 mg/l BA及0.5 mg/l NAA及10 mg/l之hygromycine進行篩選。而轉殖過程中菊花培植體與農桿菌的共培養時間，則以四天為最佳。農桿菌的轉殖試驗中，共取109個菊花培植體進行CHI基因轉殖，經篩選後得25個再生芽體，以CHI專一引子進行PCR分析，有9個芽體可偵測到CHI基因，其大小為541 bp，此PCR產物再以南方墨點法分析，更進一步證實其為轉基因植物。另外取83個培植體進行DFR基因轉殖，經篩選後共得13個再生芽體，經PCR分析，有9個再生芽體可偵測到相對的DFR基因，其大小為586 bp，以南方墨點法分析，也均可偵測到正反應。為了進一步了解轉基因併入染色體的數目，因此再進行植物基因體南方點漬法，發現所轉殖的轉DFR基因菊花，其基因體均可偵測到DFR基因的存在，而且均只有單一個插入位置，目前轉基因菊花已形成花苞，於溫室中觀察其開花情形。

關鍵詞：莖頂培養，菊花矮化類病毒，反轉錄聚合酶連鎖反應，農桿菌轉殖，共培養，南方點漬法。

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參考文獻

1. 王強生, 2002年, 利用基因工程技術圓一個古老的夢(創造新花色), 科學發展期刊, 351期。
2. 行政院農業委員會, 2001年, 農業統計年報, 行政院農業委員會。
3. 邱輝龍、范明仁 1998 花青素與花色之表現.中國園藝44:102-105。
4. 張淑芬, 菊花微體繁殖之研究, 國立中興大學園藝學研究所碩士論文, 中華民國83年6月。
5. 黃敏展. 1990. 花卉園藝栽培技術. 行政院青年輔導委員會PP:70-82。
6. 藤野守弘、藤岡作太郎、藤村良. 1973. 莖頂培養之增殖研究報告. 兵庫縣立農業試驗所試驗 研究報告 第20號別刷:125-131。
7. Aida, R., Kishimoto, S., Tanaka, Y., and Shibata, M. 2000. Modification of flower colour in torenia (*Torenia fournieri* Lind.) by genetic transformation. *Plant Sci* 153: 33-42.
8. Ben, J. J., and Langhans, R. W. 1972. Rapid multiplication of Chrysanthemum plant by stem-tip proliferation. *Hort. Sci* 7(3):289-290.
9. Broertjes, C. and Keen, A. 1980. Adventitious shoots: Do they develop from one cell. *Euphytica* 29: 73-87.
10. Bush, S. R., Earle, E. D. and Langhans, R. W. 1976. Plantlets from petal segments, petal epidermis, and shoot tips of the pericinal chimera, chrysanthemum morifolium " indianapolis." *Amer. J. Bot* 63(6):729-737.
11. Dixon, R.A., and Steele, Y.A. 1999. The phytoalexin response : Elicitation, signaling, and control of host gene expression. *Biol. Rev* 61: 239-291.
12. Earle, E. D., and Langhans, R. W. 1974. Propagation of Chrysanthemum in vitro: . Production, growth. And flowering of plantlets from tissue cultures. *J. Amer. Soc. Hort. Sci* 99(4):352-358.
13. Evans, D. A., Willan, R. S., and Cristopher, E. F. 1981. Growth and behavior of cell culture. In " Plant tissue culture method and application in Agriculture ." Academic press, New York. pp 45-113.
14. Evans, D. A. and Bravo, J. E. 1986. Phenotypic and genotypic stability of tissue cultured plants. In " Tissue Culture as a Plant Propagation System for Horticultural crops," etided by Zimmerman, R. H. , Griesbach, R. J., Hammerschlag F. A., and Lawson, R. H. Martinus Nijhoff Publishers. pp 73-94.
15. Faust, J. E. and Heins, R. D. 1992. High night temperatures do not cause poor lateral branching of chrysanthemum. *Hort Sci* 27(9):981-982.
16. Fulton, T. M., Chunwongse, J., and Tanksley, S. D. 1995 Microprep Protocol for Extrac- tion of DNA from Tomato and other Herbaceous Plants. *Plant Molecular Biology Reporter* 13: 207-209.
17. George, E. F. 1993. Plant propagation by tissue culture. *Preslia, Praha* 51: 213-237.
18. George, E. F., and Sherrington, P. D. 1984. Plant propagation by tissue culture. Exegetics Ltd., Eversley, Basinstoke, Heres 709p.
19. Hartmann, H. T. . Kester, D. E and Davies, F. T. 1990. Propagation of selected annuals and perennials used as ornamentals. In:Plant Propagation pp:609-633.
20. Helariutta, Y., Elomaa, P., Kotilainen, M., Seppanen, P., and Teerl, T. H. 1993. Cloning of cDNA coding for dihydroflavonol-4-reductase (DFR) and characterization of dfr expression in the corollase of *Gerbera hybrida* war. Regina (composites) . *Plant Mol. Biol* 22: 183-193.
21. Hirabayashi, T.T., Moriguchi, I., Kozaki, Y., and Yamamoto, T. 1987. In vitro pro- pagation of Pyrus shoot tips. *Bull. Fruit Tree Res. Stn. Japan A.* 14: 9-16.
22. Hollings, M. and Stone, O. M. 1970. Attempts to eliminate Chrysanthemum stunt from chrysanthemum by meristem-tip culture after heat treatment. *Ann. Appl. Biol* 65: 311-315.
23. Horst, R. K. and Cohen, D. 1980. Amantadine supplemented tissue culture medium: A method for obtaining chrysanthemum free of Chrysanthemum stunt viroid. *Acta Hort* 110: 315-320.
24. Khalid, N., Davey, M. R., and Power, J. B. 1989. An assesment of somaconal variation in chrysanthemum moriflium: the generation of plants of potential commerical value. *Scientia Hort.* 38: 287-294.
25. Kofranek, A. M. 1992. Cut chrysanthemum. Introduction to Floriculture, Larson, R. A. Aademic Press, New York. pp:3-35.
26. Kudo, S., Shibata, N., Kanno, Y., and Suzuki, M. 2002. Transformation of chrysanthemum (*Dendranthema grandiflorum* (Ramat.) kitamura) via *Agrobacterium tumefaciens*. *Acta Hort* 572: 139-147.
27. Lu, C. Y., Nugent, T., and Dalling, J. M. 1991. *Agrobacterium*-mediated trans- formation of carnation (*Dianthus caryophyllus* L.) *Biotechnology* 9: 864-868.
28. Mercuri, A., Sacchetti, A., Benedetti, D. L., Schiva, T., and Alberti, S. 2002 Green fluorescent flower. *Plant Sci* 162:647-654.
29. Meyer, P., Heidmann, I., Forkmann, G., Saedler, H. 1987. A new petunia flower colour generated by transformation of a mutant with a maize gene. *Nature* 330: 677-678.
30. Mitiouchkina, T. Yu., Ivanova. E. P., Taran, S. A. and Dolgov, S. V. 2000. Chalcone synthase gene from *antirrhinum majus* in antisense orientation successfully suppressed the petals pigmentation of chrysanthemum. *Acta Hort* 508: 215-217.
31. Mol, J., Grotewold, E. and Koes, R. 1998. How genes paint flowers and seeds. *Trends Plant Sci* 3: 212-217.
32. Napoli, C., Lemieux, C., and Jorgensen, R. 1990. Introduction of a Chimeric Chalcone Synthase Gene into Petunia Results in Reversible Co-Suppression of Homologous Genes in trans. *Plant Cell* 2: 279-289.
33. Paludan, N. 1985. Inactivation of viroids in chrysanthemum by low-temperature treatment and meristem-tip culture. *Acta Hort* 164: 181-186.
34. Renou, P. J., Brochard, P., and Jalouzot, R. 1993. Recovery of transgenic chrysanthemum (*Dendranthema grandiflora* Tzvelev) after hygromycin resistance selection. *Plant sci* 89:185-197.
35. Sangwan, R. S. and Harada, H. 1977. Cellular totipotency in chrysanthemum tissue cultured in vitro. *Acta Hort.* 78: 237-242.
36. Swartz, H. J. 1991. Post culture behavior: genetic and epigenetic effects and related problems. In: micropropagation. Edited by Debergh, P. C. and Zimmerman, R. H. pp 95-121.
37. Tanaka, Y., Fukul, Y., Fukuchi, M., M., Holton, T. , Higgins, E., and Kusumi, T. 1995. Molecular cloning and characterization of *Rosa Hybrida* dihydroflavonol 4-reductase gene. *Plant Cell Physiol* 36: 1025-1031.
38. Yang, C.L., Su, J. C., and Wu, J. L. 1977. Purification and propertie of sucrose synthase from the shoot of bamboo *Leleba oldhami*. *Plant Physiol* 60: 17-21.
39. Yepes, L. M., Mittak, V., Pang, S. Z., Gonsalves, C., Slightom, J. L., and Gonsalves, D. 1995. Biolistic transformation of chrysanthemum with the nucleocapsid gene of tomato spotted wilt virus. *Plant Cell Reports* 14: 694-698.