

Agrobacterium-mediated Transformation of Commercial Muskmelon (Cucumis melo L., cv.Xing Hua)

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ABSTRACT

ABSTRACT Zucchini yellow mosaic virus (ZYMV) and Type W strain of Papaya ringspot virus (PRSV-W) are transmitted by aphids and cause serious economical loss to cucurbit cultivation in Taiwan. The major objective of this study was to establish a better micropropagated system and produce coat-protein-gene transgenic plants that are resistant to ZYMV and PRSV-W. Muskmelon multiple shoots exhibited hyperhydric appearance when cultured in vitro and poor survival rate under acclimatization. In this study, we tried to decrease the hyperhydric rate by adding different concentration of thiamine HCl (10 mg l⁻¹, 50 mg l⁻¹, 100 mg l⁻¹) to the MS basal medium. The results indicated that the rate of hyperhydric shoots was reduced when the shoot culture or the medium with 50 mg l⁻¹ thiamine HCl. In the transformation procedures, seed coats or mature seeds were removed and the seeds was sterilized as explants for transformation . Each fresh cotyledon was soaked on MS solution and cut into 4 pieces using a dull scalpel blade. A fresh overnight bacterial suspension was added 50 μ l and light sharked for 10 min. All explants were transferred to the co—culture medium for four days then transplanted to the regeneration medium containing 100 ppm kanamycin and 200 ppm carbenicillin or 200 ppm cefotaxime. The putative transformed buds formed from explants cultured for 4-6 weeks on the selective medium. The best rate of bud formation was 51.7 % on the regeneration medium (MS basal medium adding 0.5 mg l⁻¹ BA, 200 mg l⁻¹ carbenicillin and 100 mg l⁻¹ kanamycin). All the buds were removed and transplanted on multiple—elongation medium (0.01 mg l⁻¹ NAA & 0.1 mg l⁻¹ BA) containing 100 mg l⁻¹ kanamycin and 200 mg l⁻¹ carbenicillin. After several times subcultures the buds developed to normal-appearance multiple shoots. We produced more than 10 putative transgenic lines. All the putative transgenic lines were conformed by PCR using specific NPTII or CP gene primer. In our study, we describe the procedures for regeneration and Agrobacterium—mediated production of transgenic Muskmelon, an economically important cultivar of melon in Taiwan. Key Words : Muskmelon、tissue culture、transformation、regeneration、Agrobacterium

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Table of Contents

目錄 封面內頁 簽名頁 授權書 1.....	iii 授權書 2.....	iv 中文摘要.....	v 英文摘要.....
.....	vii 誌謝.....	ix 目錄.....	x 圖目錄.....
.....	xiii 表目錄.....	xiv 符號說明.....
.....	xv 第一章 前人研究 1.1 洋香瓜的特性及所面臨的問題.....	1 1.2 矮南瓜黃化嵌紋病毒之發生及特性.....	2 1.3 木瓜輪點病毒西瓜系統的發生及特性.....
.....	4 1.4 交互保護策略對抗病毒之研究.....	5 1.5 洋香瓜基因轉殖之研究.....	6 1.6 農桿菌的生理特性及基因轉殖機制.....
8 第二章 材料和方法 2.1實驗材料.....	10 2.2實驗方法.....	11 2.2.1網紋洋香瓜叢生苗組織培養方法之建立	11 2.2.2網紋洋香瓜的再生培養.....
13 2.2.3網紋洋香瓜基因轉殖-改良式的子葉切割法.....	14 2.2.4轉基因株系之分子分析.....	14 2.2.4.1 植物總 DNA 抽取法.....	15 2.2.4.2 聚合酵素連鎖反應... 15 2.2.4.3 南方點漬法
16 2.2.5轉基因植物之發根及馴化處理.....	18 2.2.6轉基因株系的溫室評估.....	18 2.2.6.1溫室評估.....	19 2.2.6.2自交留種.....
.....	20 第三章 結果 3.1 網紋洋香瓜叢生苗組織培養技術建立之探討	21 3.2 不同處理對網紋洋香瓜再生試驗...
.....	24 3.3 網紋洋香瓜基因轉殖.....	25 3.4 網紋洋香瓜轉基因株系之分子分析.....	26 3.5 轉基因株系溫室抗病評估.....
.....	27 第四章 結論.....	29 參考文獻.....
.....	48 附錄一 台灣栽培之甜瓜(Cucumis melo L.)種類.....	57 附錄二 常用的基因轉殖方法.....	58 附錄三 基因轉殖作物.....
.....	59 附錄四 農桿菌之T-DNA轉殖模式.....	60 附錄五 矮南瓜黃化嵌紋病毒及木瓜輪點病毒西瓜系統鞘蛋白轉基因之構築.....	61 附錄六 植物總 DNA抽取流程圖.....
.....	62 附錄七 專一性引子設計序列.....	63 附錄八 南方點漬法(Southern blotting)裝置圖.....	64 附錄九 網紋洋香瓜發根及馴化處理流程圖.....
.....	65		

REFERENCES

- 參考文獻 余聰安。2001。木瓜微體繁殖與營養器官基因轉殖。中興大學植物學系博士論文。 林世敏。2002。不同成熟度轉殖木瓜輪點病毒鞘蛋白基因番木瓜果實其鞘蛋白基因表現之探討與其過敏原性評估。東海大學食品科學研究所碩士論文。 林詩舜。2001。台灣矮南瓜黃化嵌紋病毒株系遺傳變異之分析、基因組成之特性、具感染力轉錄載體之發展及具交互保護能力之輕症病毒株系之構築。國立中興大學農業生物科技學研究所博士論文。 陳冠君。2001。木瓜輪點病毒西瓜型生體外具感染力載體之構築及感染木瓜寄主專一性基因之分析。國立中興大學植物病理學系碩士論文。 蔡尚光。1995。設施洋香瓜與胡瓜的高品質生產。P14-23。淑馨出版社。 蔡竹固、陳瑞祥。2000。本省瓜類作物之重要病害及其管理。農業世界雜誌。200:12-19。 蔡竹固、童柏開、陳瑞祥。1999。甜瓜病害診斷及其防治。國立嘉義技術學院農業推廣委員會。20頁。 蘇宗振。1999。植物基因轉殖之研究。科學農業47(3,4):112-119。 Abel, P. P., Nelson, R. S., De, B., Hoffmann, N., Rogers, S. G., Fraley, R. T., and Beachy, R. N. 1986. Delay of disease development in transgenic plant that express the tobacco mosaic virus coat protein gene, *Science* 232 : 738-743. Akasaka - Kennedy, Y., Tomita, K. O. and Ezura, H. 2004. Efficient plant regeneration and *Agrobacterium* - mediated transformation via somatic embryogenesis in melon (*Cucumis melo* L.). *Plant Science* 166 : 763-769. Bevan, M. W., Masom, S. E., and Goelet, P., 1985. Expression of tobacco mosaic virus coat protein by cauliflower mosaic virus promoter in plants transformed by *Agrobacterium*, *EMBO J.* 4 : 1921-1926. Cabrera - Ponce, J. L., Vegas — Garcia, A. and Herrera - Estrella, L 1995. Herbicide resistant transgenic papaya plants produced by an efficient particle bombardment transformation method. *Plant Cell Rep* 15 : 1-7. Cai, W., Goncalves, C., Tennant, P., Fermin, G., Souza, M., Sarinud, N., Jan F. J., Zhu, H.Y. and Gonsalves, D.1999 A protocol for efficient transformation and regeneration of *Carica papaya* L. *In Vitro Cell. Dev. Biol.* 35 : 61-69. Chang, Y. M., Hsiao, C. H., Yang, W. Z., Hseu, S. H., Chao, Y. J., and Huang, C. H. 1987. The occurrence and distribution of five cucurbit viruses on melon and watermelon in Taiwan. *J. Agri. Res. China* 36 : 389-397. Chen, U.C., Shiau, Y.J., Lai, C.C and Tsay, H.S. 1998. Effects of mposition and vessel closure on the hyperhydricity and rooting of carnation in vitro culture. *Jour. Agric. Res. China.* 47 (4) : 364-376 Cost, A. S., and Muller, G. W. 1980. Tristeza control by cross protection: A U. S.-Brazil cooperative success. *Plant Dis.* 64 : 538-451. Davis, R. F. 1986. Partial characterization of zucchini yellow mosaic virus isolated from squash in Turkey. *Plant Dis.* 70 : 735-738. De Zoeten, G. A., and Fulton, R. W., 1975. Understanding generates possibilities. *Phytopathology* 65 : 221-222. Doyle, J.J. & Doyle, J.L., 1990. Isolation of plant DNA from fresh tissue. *Focus* 12 : 13-15. Ezura H. 2001. Genetic engineering of melon (*Cucumis melo* L.). *Plant Biotechnology.* 18 : 1-6. Fang, G. and Grumet, R. 1990. *Agrobacterium tumefaciens* mediated transformation and regeneration of muskmelon plants. *Plant Cell Rep.* 9 : 160-164. Fitchen, J. H., and Beachy R. N. 1993. Genetically engineered protection against viruses in transgenic plants. *Annu. Rev. Microbiol.* 47 : 739-763. Fletcher, J. T. 1978. The use of avirulent virus strains to protect plants against the effects of virulent strains. *Ann. Appl. Biol* 89 : 110-114. Fromm, M. E., Taylor, L. P. and Walbot, V. 1986. Stable transformation of maize after gene transfer by electroporation. *Nature* 319 : 791-793. Fulton, T.M. Chunwongse J, and Tanksley SD. 1995. Microprep Protocol for Extraction of DNA from Tomato and other Herbaceous Plants. *Plant Molecular Biology Reporter* 13 : 207-209. Gamborg, O.L., Miller, R. A. and Ojima, K. 1968. Nutrient requirements of suspension cultures of soybean root cells. *Exp.Cell.Res.* 50 : 151-158. Gelvin, S.B. 2000. *Agrobacterium* and Plant Genes involved in T-DNA transfer and integration. *Annu.Rev.Plant Physiol.Plant Mol.Biol.* 51 : 223-256 Gibbs, A. 1969. Plant virus classification. *Adv. Virus Res.* 14: 263-328. Griesbach, R.J. 1983. Protoplast microinjection. *Plant Mol.Biol.Rep.* 1 : 32-37. Guerineau, F. 1995. Tools for expressing for eign genes in Plants. In : Jones H (Ed) *Methods in Molecular Biology*, vol. 49 : *Plant Gene Transfer and Expression Protocols.* (pp. 1-32). Humana Press Inc.,Totowa, N. J. Guis, M., Amor, M. B., Latche, A., Pech, J-C., and Roustan J-P. 2000. A reliable system for the transformation of cantaloupe charentais melon (*Cucumis melo* L. var. *cantalupensis*) leading to majority of diploid regenerants. *Scientia Horticulturae* 84 : 91-99. Heniknff, S. 1984. Unidirectional digestion with exonuclease III creates targeted break-points for DNA sequencing. *Gene* 28 : 351-359. Hollings, M., and Brunt, A.A. 1981. Potyvirus group. CMI/AAB Descriptions of plant viruses no.245. Kew, Surrey. Hseu, S. H., Wang, H. L., and Huang, C. H. 1985. Identification of a zucchini yellow mosaic virus from *Cucumis astivus*. *J. Agri. Res. China* 34 : 87-95. Hseu, S. H., Huang, C. H., Chang, C. A., Yang, W. Z., Chang, Y. M., and Hsiao, C. H. 1987. The occurrence of five viruses in six cucurbits in Taiwan. *Plant Prot. Bull. (Taiwan)* 29 : 233-244. Huang, C. H., Chang, L., and Tsai, J. H. 1993. The partial characterization of melon vein-banding mosaic virus, a newly recognized virus infecting cucurbits in Taiwan. *Plant Pathol.* 42 : 100-107. Kadota, M., Imizu, K and Hirano, T. 2001. Double-phase in vitro culture using sorbitol increases shoot proliferation and reduces hyperhydricity in Japanese pear. *Sci. Hort.* 89 : 207-215. Klein, T. M., Wolf, E.D., Wu, R. and Sanford, J.C.1987. High velocity microprojectiles for delivery of nucleic acids into living cell. *Nature* 327 : 70-73. Ku, H.M & Tsay, H.S. 1994a. Effect of medium composition on the vitrification of carnation plantlets cultured in vitro. *Jour.Agric.Res.China.* 43 (1) : 51-62. Ku, H.M & Tsay, H.S. 1994b. Influence of subculture generation on the vitrification of carnation plantlets culture in vitro. *Jour.Agric.Res.China.* 43 (3) : 308-319. Lecoq, H., Lisa, V., and Dellavalle, G. 1983. Serological identity of Muskmelon yellow stunt and Zucchini yellow mosaic viruses. *Plant Dis.*67 : 824-825. Lisa, V., and Lecoq, H. 1984. Zucchini yellow mosaic virus. CMI/AAB Description of Plant Virus, No. 282. Kew, Surrey. Lisa, V., Boccardo, G., D'Agostino, G., Dellavalle, G., and d'Aquilio, M. 1981. Characterization of a potyvirus that causes Zucchini yellow mosaic. *Phytopathology* 71 : 667-672. Lotfi, M., Alan, A. R., Henning, M. J., Jahn, M. M. and Earle, E. D. 2003. Production of haploid and doubled haploid plants of melon (*Cucumis melo* L.) for use in breeding for multiple virus resistance. *Plant Cell Reports* 21 : 1121-1128. Lovisolo, O. 1981. Virus and viroid disease of cucurbits. *Acta Horticulturae.* 88 : 33-82. Luo, Z. and Wu, R.1989. A simple method for the transformation of rice via the pollen-tube pathway. *Plant Mol.Biol.Rep.*7 : 69-77. Mathias, T. J. and Boyd, L. A. 1986. Cefotaxime stimulates callus growth embryogenesis and regeneration in hexaploid bread wheat (*Triticum aestivum* L EM. Thell). *Plant*

Sci. 46 : 217-233. Mathias, R. J. and Mukasa, C. 1987. The effect of cefotaxime in the growth and regeneration of callus from four varieties of barley (*Hordium vulgare* L.). *Plant Cell Rep.* 6 : 454-457. Mahgoub, H.A., Desbiez, C., Wipf-Scheibel, C., Dafalla, G., and Lecoq, H. 1997. Characterization and occurrence of zucchini yellow mosaic virus in Sudan. *Plant Pathol.* 46 : 800-805. Milne, K. S., Grogan, R. G. and Kimble, K. A. 1969. Identification of viruses infecting cucurbits in California. *Phytopathology* 59 : 819-828. Murashige, T. and Skoog, F. 1962. A revised medium for rapid growth and bioassays with tobacco tissue cultures. *Physiol. Plant.* 15 : 473-497. Nameth, S. T., Dodds, J. A., Paulus, A. O. and Laemmlen, F. F. 1986. Cucurbit viruses of California : An ever-changing problem. *Plant Dis.* 70 : 8-11. Nauerby, B., Billing, K. and Wyndaele, R. 1997. Influence of the antibiotic timentin on plant regeneration compared to carbenicillin and cefotaxime in concentrations suitable for elimination of *Agrobacterium tumefaciens*. *Plant Sci.* 123 : 169-177. Norelli, J. N. and Aldwinckle, H. S. 1993. The role of aminoglycoside Antibiotics in the regeneration and selection of neomycin phosphotransferase-transgenic apple tissue. *J. Amer. Soc. Hort. Sci.* 118 : 311-316. Okkels, F. T. and Pederson, M. G. 1988. The toxicity of plant tissue and to *Agrobacterium tumefaciens* of some antibiotics. *Acta Hort.* 225 : 199-207. Ponz, F., and Bruening, G. 1986. Mechanism of resistance to plant viruses. *Ann. Rev. Phytopathol.* 24: 336-381. Provvidenti, R. 1986. Viral disease of cucurbits and sources of resistance. Food & Fertilizer Technology Center. Technical bulletin. No. 93. Provvidenti, R., Gonsalves, D., and Humaydan, H.s. 1984. Occurrence of zucchini yellow mosaic virus in cucurbits from Connecticut, New-York, Florida, and California. *Plant Dis.* 68 : 443-446. Purcifull, D. E., Edwardson, J. R., Hiebert, E., and Gonaslves, D. 1984. Papaya ringspot virus. CMI/AAB Description of Plant Virus. No. 292. Restrepo, M. A., Feed, D.D., and Carrington, J.C. 1990. Nuclear transport of plant potyviral proteins. *Plant Cell* 2 : 987-998. Sambrook, J., Fritsch, E.F., and Maniatis, T. 1989. Analysis and cloning of eukaryotic genomic DNA.. In *Molecular cloning*. 2 nd.vol. 2 : 9.34-9.45 Cold Spring Harbor Laboratory Press. Schenk, R. U. and A. C. Hildebrandt, 1972, Medium and Techniques for Induction and Growth of Monocotyledonous and Dicotyledonous Plant Cell Cultures, *Can. J. Bot.* 50 : 199-204. Tabei, Y., T. Kanno, T. & Nishio, T. (1991): Regulation of organogenesis and somatic embryogenesis by auxin in melon, *Cucumis melo* L. *Plant Cell Rep.* 10 : 225-229. Taravira, N. and Kintzios, S. 1997. Effect of genotype and light intensity on somatic embryogenesis and plant regeneration in melon (*Cucumis melo* L.) *Plant Breed.* 116 : 359-362. Tepfer, M. 1993. Viral genes and transgenic plants. *Bio / Technology* 11 : 1125-1132. Thomas, P., Mythili, J. B. and Shivashankara, K. S. 2000. Explant, medium and vessel aeration affect the incidence of hyperhydricity and recovery of normal plantlets in triploid watermelon. *Journal of Horticultural Science and Biotechnology* 75 (1) : 19-25. Tomlinson, J. A. 1987. Epidemiology and control of virus disease of vegetables. *Ann. Appl. Biol.* 110 : 661-681. Tzfira, T & Citovsky, V. 2002. Partners-in-infection: host proteins involved in the transformation of plant cells by *Agrobacterium*. *Trends in Cell Biology* 12 (3) : 121-129. Valles, M.P. & Lasa, J. M. 1994. *Agrobacterium*-mediated transformation of commercial melon (*Cucumis melo* L., Amarillo Oro). *Plant Cell Rep.* 13 : 145-148. Vandenmoortele, J. L. 1999. A procedure to prevent hyperhydricity in cauliflower axillary shoots. *Plant Cell Tiss. Org. Cult.* 56 : 85-88. Yashida, K., Goto, T., Nemoto, M. and Tsuchizaki, T. 1980. Rive viruses isolated from melon (*Cucumis melo* L.) in Hokkaido. *Ann. Phytopath. Soc. Japan.* 46 : 339-343. Yeh, S.D., Jan, F. J., Chiang, C.H., Doong, T. J., Chen, M. C., Chung, P. H., and Bau, H. J. 1992. Complete nucleotide sequence and genetic organization of Papaya ringspot virus RNA. *J. Gen. Virol.* 73 : 2531-2541. Yu, T. A., Yeh, S. D., and Yang, J. S. 2000. Comparison of the effects of kanamycin and geneticin on regeneration of papaya from root tissue. *Plant Cell Tiss. Org. Cult.* 74 : 169-178. Zupan, J.R. and Zambryski, P. 1995. Transfer of T-DNA *Agrobacterium* to the plant cell. *Plant Physiol.* 107 : 1041-1047.