The Studies of the G 11 Pseudogene in Leukemia Cell Line K562 Differentiation

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ABSTRACT

Heterotrimeric guanine nucleotide-binding proteins (G-protein) have been demonstrated to play integral role in the transduction of extracellular signals from cell membrane receptor (G-protein-coupled receptors, GPCR) to intracellular effectors proteins. G proteins regulate critical processes such as cell growth, differentiation and development. Chronic myeloid leukemia (CML) is a clonal myeloproliferative disorder of hematopoietic stem cells, that has acquired a Philadelphia (Ph) chromosome encoding the BCR - ABL oncogenic fusion protein, which has lost its differentiation activity. Therefore, in recent years, some scholars proposed differentiation therapy, by treating appropriate inducers to induced advanced or aggressive malignant cells maturation and differentiation into mature cells. K562 is the first human immortalized myelogenous leukaemia cell line and belonging undifferentiated pluripotent hematopoietic progenitor cells. K562 cell can be differentiated into erythrocytic or megakaryocytic lineages upon different inducers treatments and was used as a model cell line to study the relationship between the blood cell differentiation and signal transduction. In this report, three different inducers, huangqi (Astragalus membranaceus) and chemicals Hemin and HMBA were used to induce K562 cell differentiation. Two erythroid markers, -globin or -globin, and two megakaryocyte markers CD41 and CD61 are used to monitor the differentiation process. In former report, the G 11 pseudogene was induced by the huangqi administration. In this study, in promoter activity assay the pseudogene promoter activity increased by two fold when the presence of 1.5 mg / ml of huanggi extract. However, under the influence of the three-inducing agent, the G 11 pseudogene were up-regulated in the HMBA-induced K562 cell, but we failed to detect significant changing of cell differentiation markers. It may be the consequence of fetal bovine serum used in cell culture which led to change cell characters and low transfection efficiency of the genes. Currently, re-transfection with higher amount of expression plasmid in K562 cells, and select stable clones are on going. I expect the performance of the proper G proteins function will change cell fate. In conclusion, understand the G protein function in cell differentiation, can provide the information need for future differentiation therapy of leukemia.

Keywords: K562 cell、Huangqi、Hemin、HMBA、G q、G 11、G 11 pseudogene

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REFERENCES

1.周成功。2005。1994年生理醫學桂冠,G蛋白檔案 (諾貝爾的榮耀·生理醫學桂冠,林榮崧)。天下遠見出版股份有限公司。台北市,台 灣。2.涂慧珠。2006。GNAS、Gi、Gq蛋白經由Huangqi、Hemin和HMBA誘導K562細胞分化中所扮演之角色。大葉大學。分子生物科 技學系。彰化。台灣。3.來國鈞。2008。不同型G蛋白在藥劑誘導K562細胞分化下所扮演調合者角色。大葉大學。分子生物科技學系。 彰化。台灣。4.Andersson, Nilsson, Gahmberg, K562--a human erythroleukemic cell line. International journal of cancer, Journal international du cancer 23, 143-147 (1979).5. Astesano, A. et al. Cellular and Subcellular Expression of Golf/Gs and Gg/G11 Subunits in Rat Pancreatic Endocrine Cells. Journal of Histochemistry & Cytochemistry 47, 289-302, (1999).6. Berestetskaya, Regulation of Apoptosis by alpha -Subunits of G12 and G13 Proteins via Apoptosis Signal-regulating Kinase-1. Journal of Biological Chemistry 273, 27816-27823, (1998).7. Brass, Manning, Cichowski, Abrams. Signaling through G proteins in platelets: to the integrins and beyond. Thrombosis and haemostasis 78, 581-589, (1997).8. Brooks, Regulation of fibroblast cell cycle by serum. Nature 260, 248-250, (1976).9. Chang, Kan. Correction of the sickle cell mutation in embryonic stem cells. Proceedings of the National Academy of Sciences of the United States of America 103, 1036-1040, (2006).10. Christie P. Thomas, Rafael Mattera, Ca2+ signalling in K562 human erythroleukaemia cells: effect of dimethyl sulphoxide and role of G-proteins in thrombin- and thromboxane A2-activated pathways. (1995).11.Costa, G., Kouskoff, V. & Lacaud, G. Origin of blood cells and HSC production in the embryo. Trends in immunology 33, 215-223, (2012).12. Crouthamel, M. et al. An N-terminal polybasic motif of Galphag is required for signaling and influences membrane nanodomain distribution. Molecular pharmacology 78, 767-777, (2010).13.Daley, Van Etten, Baltimore, D. Induction of chronic myelogenous leukemia in mice by the P210bcr/abl gene of the Philadelphia chromosome. Science 247, 824-830, (1990).14. Debili, N. Different expression of CD41 on human lymphoid and myeloid progenitors from adults and neonates. Blood 97, 2023-2030, (2001).15. Downes, N. G. The G protein subunit gene families. (1999).16. Fraser, Berridge. Induction of B-lymphocyte antigens on the chronic myeloid leukemic cell line K562 using sodium butyrate. Experimental hematology 15, 406-413, (1987).17.Gabriela J. Greif, Bruce P. Bean, Eva J. Neer, And Ulrike Mende. Altered regulation of potassium and calcium channels by GABA(B) and adenosine receptors in hippocampal neurons from mice lacking Galpha(o). (2000).18.Gauthami Jalagadugula, Koneti Rao. Phorbol 12-myristate 13-acetate (PMA) responsive sequence in Galphag promoter during megakaryocytic differentiation. Regulation by EGR-1 and MAP kinase pathway. (2008).19.Gerhard J. Johnson, Specificity of G alpha q and G alpha 11 gene expression in platelets and erythrocytes. Expressions of cellular differentiation and species differences. (1996).20.Gocek, Marcinkowska, E. Differentiation Therapy of Acute Myeloid Leukemia. Cancers 3, 2402-2420, (2011).21.Gunsilius, Gastl, Petzer. Hematopoietic stem cells. Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie 55, 186-194, (2001).22. Higgs, D. R. Gene regulation in hematopoiesis new lessons from thalassemia. (2004).23. Jacquel, A. et al. A survey of the signaling pathways involved in megakaryocytic differentiation of the human K562 leukemia cell line by molecular and c-DNA array analysis. Oncogene 25, 781-794, (2006).24. Jianguo Jin. Coactivation of two different G protein-coupled receptors is essential for ADP-induced platelet aggregation. (1998).25. Knudsen, S. Promoter 2.0 for the recognition of PolII promoter sequences. (1999).26. Koeffler, Golde. Human myeloid leukemia cell lines: a review. Blood 56, 344-350, (1980).27. Kucukkaya, B., Arslan, D. O. & Kan, B. Role of G proteins and ERK activation in hemin-induced erythroid differentiation of K562 cells. Life sciences 78, 1217-1224, (2006).28.Liu, J. The Stimulatory G Protein -Subunit Gs Is Imprinted in Human Thyroid Glands: Implications for Thyroid Function in Pseudohypoparathyroidism Types 1A and 1B, Journal of Clinical Endocrinology & Metabolism 88, 4336-4341, (2003).29.Livia CioA, Howard R. Hubbell, Pacifico Meo, Peter Curtis, Giovanni Povera. Differential expression of the globin genes in human leukemia K562. (1981).30.Lozzio, C.B. and Lozzio, B.B. Human Chronic M yelogenous Leukemia Cell-Line With Positive Philadelphia Chromosome. (1975).31. Malbon, C.C. G proteins in development. Nature reviews. Molecular cell biology 6, 689-701, (2005).32.Marie JP, I.C., Civin CI, Mirro J, McCulloch EA. The presence within single K-562 cells of erythropoietic and granulopoietic differentiation markers. (1981).33.Mark G. Davis, Y.K., Ifeanyi J. Arinze. Involvement of Gi 2 in sodium butyrate-induced erythroblastic differentiation of K562 cells. (2000).34.Marks, P.A., Richon, V.M. & Rifkind, R.A. Induced differentiation of cancer cells: second generation potent hybrid polar compounds target cell cycle regulators. Eur J Cancer Prev 5 Suppl 2, 75-77 (1996).35. Mary Lynn Benka, M.L., Guang-Rong Wang, ShaAvhree Buckman. The thrombin receptor in human platelets is coupled to a GTP binding protein of the G g family. (1995).36.Mikkola, H.K. Orkin, S.H. The journey of developing hematopoietic stem cells. Development 133, 3733-3744, (2006).37.Milligan, G. Kostenis, E. Heterotrimeric G-proteins: a short history. British journal of pharmacology 147 Suppl 1, S46-55, (2006).38. Milligan. Agonist-induced transfer of the alpha subunits of the quanine-nucleotide-binding regulatory proteins G alpha q and G alpha 11 and of muscarinic ml acetylcholine receptors from plasma membranes to a light-vesicular membrane fraction. (1994).39. Mironneau, J. Macrez, N. Specificity of G(q) and G(11) Protein Signaling in Vascular Myocytes. Trends in cardiovascular medicine 8, 157-162, (1998).40. Moss, J. Vaughan, M. ADP-ribosylation of guanyl nucleotide-binding regulatory proteins by bacterial toxins. Advances in enzymology and related areas of molecular biology 61, 303-379, (1988).41.N Monplaisir, G.M., C Poyart, M D Rhoda, C Craescu, M Vidaud, F Galacteros, Y Blouquit, and J Rosa. Hemoglobin S Antilles a variant with lower solubility than hemoglobin S and producing sickle cell disease in heterozygotes. (1986).42. Nana Kawasaki, Kazushige Morimoto, Takao Hayakawa. Control of Hemoglobin Synthesis in Erythroid Differentiating K562 Cells. (1996).43. Olefsky, J.M. Nuclear receptor minireview series. The Journal of biological chemistry 276, 36863-36864, (2001).44.Park, D. The Role of Carboxyl-terminal Basic Amino Acids in Ggalpha -dependent Activation, Particulate Association, and Nuclear Localization of Phospholipase C-beta 1. Journal of Biological Chemistry 271, 21187-21192, (1996).45. Paul R. Albert, L.R. G protein specificity traffic direction required. (2002).46.Philippe. Structure, localization and transcriptional properties.

(1992).47. Prestridge, D.S. Predicting Pol II Promoter Sequences Using Transcription Factor Binding Sites. (1995).48. Ren, R. Mechanisms of BCR-ABL in the pathogenesis of chronic myelogenous leukaemia. Nature reviews. Cancer 5, 172-183, (2005).49.Roberta C. Reuben, R.L. W., Ronald Breslow, Richard A. Rifkind, And Paul A. Marks. A new group of potent inducers of differentiation in murine erythroleukemia. (1976).50.Rutherford, T.R., Clegg, J. B. Weatherall, D.J. K562 human leukaemic cells synthesise embryonic haemoglobin in response to haemin. Nature 280, 164-165 (1979).51. Sachs, L. Hematopoietic growth and differentiation factors and the reversibility of malignancy: cell differentiation and by-passing of genetic defects in leukemia. Medical oncology and tumor pharmacotherapy 3, 165-176 (1986).52. Sawyers, C.L. Chronic myeloid leukemia. The New England journal of medicine 340, 1330-1340, (1999).53.Simon, M.S. A.M. G protein diversity A distinct class of a subunits is present in vertebrates and invertebrates. (1990).54.Strader, C.D., Fong, T.M., Graziano, M.P. Tota, M.R. The family of G-protein-coupled receptors. FASEB journal: official publication of the Federation of American Societies for Experimental Biology 9, 745-754, (1995).55. Takeshi Yagami, Julio Cesar Padovan, Brian T. Chait, Anthony M. Popowicz, James M. Manning. N-terminal contributions of the gamma-subunit of fetal hemoglobin to its tetramer strength: Remote effects at subunit contacts. (2002).56. Tim Rutherford, Higgs, R.W. Jones, J. Thompson. Embryonic erythroid differentiation in the human leukemic cell line K562. (1981).57.Vogelstein, B. Kinzler, K.W. Cancer genes and the pathways they control. Nature medicine 10, 789-799, (2004).58. Wang, M., Wang, L., Pan, X. J. Zhang, H. Monocytic differentiation of K562 cells induced by proanthocyanidins from grape seeds. Archives of pharmacal research 35, 129-135, (2012).59. Weatherall, D.J. Thalassaemia and malaria, revisited. Annals of tropical medicine and parasitology 91, 885-890, (1997).60. Xiao-Dong Cheng et al. Effects of Huanggi (Hex) on Inducing Cell Differentiation and Cell Death in K562 and HEL Cells. (2004).61. Yonish-Rouach, E. et al., Wild-type p53 induces apoptosis of myeloid leukaemic cells that is inhibited by interleukin-6. Nature 352, 345-347, (1991).62. Yueh-Lun Lee1, Fu-Hwa Liu3, Yu-Wen Huang, Huei-Mei Huang. Aclacinomycin A Sensitizes K562 Chronic Myeloid Leukemia Cells to Imatinib through p38MAPK-Mediated Erythroid Differentiation. (2013).63.Zhi-Xiang Shen, Jian-Hua Ni, Xiu-Shong Li, Shu-Min Xiong, Qian-Yao Qiu, Jun Zhu, Zhang, Sai-Juan Chen, Zhu Chen and Zhen-Yi Wang. Use of Arsenic Trioxide (As2O3) in the Treatment of Acute Promyelocytic Leukemia (APL). (1997).