

Production of Recombinant Bovine Lactoferricin Trimer Peptide Using *Pichia pastoris*

陳澧樺、陳小玲

E-mail: 364887@mail.dyu.edu.tw

ABSTRACT

This study is using the generation of the recombinant yeast, *Pichia pastoris*, containing the bovine lactoferricin gene fragment of monomer, dimer and trimer and the feasibility of production. First, we selected 20-amino acid antibacterial corepeptide from bovine lactoferricin. The coding sequences of desired peptides are synthesized oligonucleotides sequences, and cloned the target gene into pGAPZ C yeast-expression vector. After confirmed by DNA sequencing, the recombinant vector BLFc_{in} / pGAPZ C monomer, dimer and trimer were transformed into *Pichia pastoris* GS115 by electroporation and the transformants were identified by expressing the biological antibacterial peptides. We could get high copy number strains selected by high multiple antibiotic, zeocin and culture B-LFc_{in} / pGAPZ C Trimer 29 in the growing medium by flasks. Collecting supernatant and analyzing by Tricine SDS-PAGE and Western blot. Data showed that a 10.8 kD predicted band was obtained in different culture periods. Therefore, we cultured recombinant B-LFc_{in} / pGAPZ C Trimer 29 by fermentor and supernatant could be collected. Fast protein liquid chromatography was utilized to purify the lactoferricin from cellular extracts of BLFc_{in} / pGAPZ C. The crude extracted protein was purified by Hitrap Heparin column and then was concentrated. The target protein showed that a 10.8 kD predicted band by Tricine SDS-PAGE. The condition of purification for lactoferricin needs further improvement.

Keywords : LFc_{in}、*Pichia pastoris*

Table of Contents

目錄 封面內頁 簽名頁 中文摘要	iii 英文摘要	
..... v 誌謝		
vi 目錄	vii 圖目錄	
..... xii 1. 緒言		
..... 1 2. 前言		
2 2.1 乳鐵蛋白	2 2.1.1 抗菌活性	
..... 3 2.1.2 抗真菌活性	3 2.1.3 抗寄生蟲活	
性	3 2.1.4 抗病毒活性	4 2.1.5 人體健康上的
影響	4 2.1.6 做為疾病的生物標記	5 2.2 乳鐵蛋白素
..... 6 2.2.1 抗菌活性	6 2.2.2 抗真菌和	
抗寄生蟲活性	7 2.2.3 抗病毒活性	7 2.2.4 抗腫瘤活性
..... 8 2.2.5 免疫調節作用	8 2.3 活性多?之表現	
系統	9 2.3.1 原核細胞表現系統	9 2.3.2 真核細胞表
現系統	9 3. 材料與方法	11 3.1 重組牛乳
鐵蛋白素於 pGAPZ C/ E-coli 之建構	11 3.1.1 牛乳鐵蛋白素之基因片段來源	11 3.1.1.1 合
成牛乳鐵蛋白素胜?片段	11 3.1.1.2 雙股去氧核醣核酸黏合作用	11 3.1.1.2 牛乳鐵蛋白素構
築於酵母菌表現型載體中	11 3.1.2.1 酵素截切	12 3.1.2.2 酵素截切 DNA 片段
回收	12 3.1.2.3 DNA 接合反應	13 3.1.2.4 勝任細胞的製備
..... 13 3.1.2.5 勝任細胞的轉型作用	14 3.1.2.6 快速膠體篩選法	
(Creacking gel)	14 3.1.2.7 菌落聚合?連鎖反應	14 3.1.2.8 DNA 定序分析
..... 15 3.2 酵母菌轉型株的篩選	15 3.2.1 酵母菌勝任細胞製	
備	16 3.2.2 電穿孔	16 3.2.3 高倍率抗生素篩選
..... 17 3.2.4 酵母菌染色體 DNA 萃取與 PCR 篩選	17 3.3 酵母菌轉型株 RNA 轉	
錄分析	18 3.3.1 酵母菌轉型株總 RNA 萃取	18 3.3.2 RNA 萃取過程
DNA 殘留檢測	19 3.3.3 去除殘留 DNA	19 3.3.4 反轉錄聚合?連鎖
反應	20 3.4 酵母菌轉型株的培養	20 3.5 酵母菌轉型株生長
曲線	20 3.6 酵母菌轉型株細胞內與細胞外蛋白質分析	21 3.6.1 酵母菌轉型
株蛋白質製備	21 3.6.1.1 酵母菌轉型株細胞內蛋白質萃取	21 3.6.1.2 酵母菌轉型株細

胞外蛋白質濃縮	21	3.6.2 蛋白質分析	22	3.6.2.1 三氫甲基甘氨酸-麥黃
酮-硫酸十二酯鈉聚丙醯胺 膠電泳			22	3.6.2.1.1 蛋白質前處理
.....		22 3.6.2.1.2 蛋白質變性電泳.....	23	3.6.2.1.3 Coomassie Blue 染色
.....	23	3.6.2.2 西方墨點法分析	23	3.6.2.2.1 轉漬作用
.....		24 3.6.2.2.2 西方墨點法	24	3.7 重組酵母菌牛乳鐵蛋白素基因大
量生產	25	3.7.1 重組酵母菌株活化	25	3.7.2 發酵槽前置設定與接種
.....		26 3.7.3 發酵槽饋料	27	3.7.4 發酵槽生長曲線與發酵液
收取	28	3.8 重組牛乳鐵蛋白素基因發酵液粗純化與濃縮	28	3.8.1 發酵液粗過濾 (0.45
μ M Hollow Fiber)	28	3.8.2 使用膜孔徑 100 kD 與 30 kD 卡匣過濾濃縮	29	3.8.3 發酵液以 3 kD Hollow
Fiber 進行濃縮	29	3.9 重組牛乳鐵蛋白素蛋白質含量測定法	30	3.10 利用快速蛋白質液相層
析系統純化重組牛乳鐵蛋白素 ...	30	3.10.1 上機前處理	31	3.10.2 高效能液相層析系統
.....		32 3.10.3 管柱清洗與保存	32	3.10.4 下機管線清洗與保存
.....		32 3.10.5 小量濃縮重組蛋白	33	3.11 以不同表現型載體產製重
組牛乳鐵蛋白素三元體	33	3.11.1 重組牛乳鐵蛋白素二元體酵素截切	34	3.11.2 重組牛乳鐵蛋白素二元
體基因片段回收	34	3.11.3 構築重組牛乳鐵蛋白素三元體於 pPICZ B 表現型載 體中		
.....		35 4. 結果與討論	36	
4.1 牛乳鐵蛋白素基因於酵母菌表現型載體之建構	36	4.2 重組酵母菌菌株之轉型與篩選		
.....		37 4.3 持續釋泌型酵母菌轉形菌株之生長測定	38	4.4 甲醇誘導型酵母菌轉形菌
株之生長測定	38	4.5 酵母菌轉型株蛋白質分析.....	39	4.6 重組牛乳鐵蛋白素初
步純化分析	39	4.7 重組酵母菌於發酵槽產製蛋白	40	4.7.1 B-LFcin/
pGAPZ C trimer 29 發酵產製	40	4.7.2 B-LFcin/pPICZ B trimer 37 發酵產製	41	4.7.3 利用切
向流濃縮透析系統	41	4.7.4 利用高效能液相層析系統純化	41	5. 結論
.....		43 參考文獻		

60

REFERENCES

- Artym, J., and Zimecki, M. (2007) The effects of lactoferrin on myelopoiesis: can we resolve the controversy? *Postepy Hig. Med. Dosw.* 61: 129 – 150.
- Ashby, B., Garrett, Q., and Willcox, M. (2011) Bovine lactoferrin structures promoting corneal epithelial wound healing in vitro. *Invest. Ophthalmol. Vis. Sci.* 52(5): 2719 – 2726.
- Aramaki, Y., Takano, S., Arima, H., and Tsuchiya, S. (2000) Induction of apoptosis in WEHI 231 cells by cationic liposomes. *Pharm. Res.* 17: 515 – 520.
- Andra, J., Lohner, K., Blondelle, S. E., Jerala, R., Moriyo, I., Koch, M. H. J., and Brandenburg, K. (2005) Enhancement of endotoxin neutralization by coupling of a C12-alkyl chain to a lactoferricin-derived peptide. *Biochem. J.* 385:135 – 143.
- Johanson, B. (1960) Isolation of an iron-containing red protein from human milk. *Acta Chem. Scand.* 14:510-512.
- Bellamy, W., Takase M., and Wakabayashi, H. (1992) Antibacterial spectrum of lactoferricin B, a potent bactericidal peptide derived from the N-terminal region of bovine lactoferrin. *J. Appl. Bacteriol.* 73(6):472 – 479.
- Bellamy, W., Wakabayashi, H., Takase, M., Kavase, K., Shinamura, S., and Tomita, M. (1993) Killing of *Candida albicans* by lactoferricin B, a potent antimicrobial peptide derived from the N-terminal region of bovine lactoferrin. *Med. Microbiol. Immunol.* 182:97 – 105.
- Bellamy, W., Yamauchi, K., Wakabayashi, H., Takase, M., Takakura, N., and Simamura, S. (1994) Antifungal properties of lactoferricin B, a peptide derived from the N-terminal region of bovine lactoferrin. *Lett. Appl. Microbiol.* 18: 230 – 233.
- Beutler, B. and Cerami, A. (1988) Tumor necrosis, cachexia, shock and inflammation: a common mediator. *Annu. Rev. Biochem.* 57: 505 – 518.
- Blais, A., Malet, A., Mikogami, T., Martin-Rouas, C., and Tome, D. (2009) Oral bovine lactoferrin improves bone status of ovariectomized mice. *Am. J. Physiol. Endocrinol. Metab.* 296(6): E1281 – E1288.
- Chu, B.C., Garcia-Herrero, A., Johanson, T.H., Krewulak, K.D., Lau, C.K., and Peacock, R.S. (2010) Siderophore uptake in bacteria and the battle for iron with the host; a bird's eye view. *Biometals* 23(4): 601 – 611.
- Cintra, W. M., Silva-Filho, F. C., and De Souza, W. (1986) The surface charge of *Toxoplasma gondii*: a cytochemical and electrophoretic study. *J. Submicrosc. Cytol.* 18:773 – 781.
- Cornish, J., Callon, K.E., Naot, D., Palmano, K.P., Banovic, T., and Bava, U. (2004) Lactoferrin is a potent regulator of bone cell activity and increases bone formation in vivo. *Endocrinology* 145(9): 4366 – 4374.
- Dionysius, D. A., and Milne, J. M. (1997) Antibacterial peptides of bovine lactoferrin: purification and characterization. *J. Dairy Sci.* 80: 667 – 674.
- Engelmayer, J., Blezinger, P., and Varadhachary, A. (2008) Lactoferrin stimulates wound healing with modulation of inflammation. *J. Surg. Res.* 149(2):278 – 286.
- Eliassen, L. T., Berge, G., Sveinbjornsson, B., Svendsen, J. S., Vorland, L. H. and Rekdal, O. (2002) Evidence for a direct anti-tumor mechanism of action of bovine lactoferricin. *Anticancer Res.* 22: 2703 – 2710.
- Fujihara, T., and Hayashi, K. (1995) Lactoferrin inhibits herpes simplex virus type-1 (HSV-1) infection to mouse cornea. *Arch. Virol.* 140:1469 – 1472.
- Gifford, J. L., Hunter, H. N., and Vogel, H. J. (2005) Lactoferricin: a lactoferrin-derived peptide with antimicrobial, antiviral, antitumor and immunological properties. *Cell Mol. Life Sci.* 62(22):2588 – 2598.
- Glimvall, P., Wickstrom, C., and Jansson, H. (2012) Elevated levels of salivary lactoferrin, a marker for chronic periodontitis? *J. Periodontal Res.*
- Grover, M., Giouzeppos, O., Schnagel, R. D., and May, J. T. (1997) Effect of human milk prostaglandins and lactoferrin on respiratory

syncytial virus and rotavirus *Acta Paediatr* 86:315 – 316. 21. Hara, K., Ikeda, M., and Saito, S. (2002) Lactoferrin inhibits hepatitis B virus infection in cultured human hepatocytes. *Hepatol. Res.* 24(3):228. 22. Harmsen, M. C., Swart, P. J., and De Bethune, M. P. (1995) Antiviral effects of plasma and milk proteins: lactoferrin shows potent activity against both human immunodeficiency virus and human cytomegalovirus replication in vitro. *J. Infect. Dis.* 172:380 – 388. 23. Hasegawa, K., Motosuchi, W., Tanaka, S., and Dosako, S. (1994) Inhibition with lactoferrin of in vitro infection with human herpes virus. *Jpn. J. Med. Sci. Biol.* 47:73 – 85. 24. He, J., and Furmanski, P. (1995) Sequence specificity and transcriptional activation in the binding of lactoferrin to DNA. *Nature* 373: 721 – 724. 25. Montreuil, J., Tonnelat, J., and Mullet, S. (1960) Preparation and properties of lacto- transferrin of human milk. *Biochim. Biophys. Acta.* 45:413 – 421. 26. Kullberg, B. J., Netea, M. G., Vonk, A. G. and van der Meer, J. W. (1999) Modulation of neutrophil functions in host defense against disseminated *Candida albicans* infection in mice. *FEMS Immunol. Med. Microbiol.* 26: 299 – 307. 27. Ikeda, M., Sugiyama, K., Tanaka, T., Tanaka, K., Sekihara, H., Shimotohno, K., and Kato, N. (1998) Lactoferrin markedly inhibits hepatitis C virus infection in cultured human hepatocytes. *Biochem. Biophys Res. Commun.* 245:549 – 553. 28. Leon-Sicairos, N., Martinez-Pardo, L., Sanchez-Hernandez, B., de la Garza, M., and Carrero, C. (2012) Oral lactoferrin treatment resolves amoebic intracecal infection in C3H/ HeJ mice. *Biochem. Cell Biol.* 90(3):435 – 41. 29. Lupetti, A., Paulusma-Annema, A., Welling, M. W., Senesi, S., van Dissel, J. T. and Nibbering, P. (2000) Candidacidal activities of human lactoferrin peptides derived from the N-terminus. *Antimicrob. Agents Chemother.* 44: 3257 – 3263. 30. Groves, M.L. (1960) The isolation of a red protein from milk. *J. Amer. Chem. Soc.* 82:3345 – 3350. 31. Malet, A., Bournaud, E., Lan, A., Mikogami, T., Tome, D., and Blais, A. (2011) Bovine lactoferrin improves bone status of ovariectomized mice via immune function modulation. *Bone.* 48(5): 1028 – 1035. 32. Mattsby-Baltzer, I., Roseanu, A., Motas, C., Elverfors, J., Engberg, I. and Hanson, L. A. (1996) Lactoferrin or a fragment thereof inhibits the endotoxin-induced interleukin-6 response in human monocytic cells. *Pediatr. Res.* 40: 257 – 262. 33. Murphy, M. E., Kariwa, H., Mizutani, T., Yoshimatsu, K., Arikawa, J., and Takashima, I. (2000) In vitro antiviral activity of lactoferrin and ribavirin upon hantavirus. *Arch Virol* 145, 1571 – 1582. 34. Naot, D., Grey, A., Reid, I.R., and Cornish, J. (2005) Lactoferrin – a novel bone growth factor. *Clin. Med. Res.* 3(2): 93 – 101. 35. Omata, Y., Satake, M., and Maeda, R. (2001) Reduction of the infectivity of *Toxoplasma gondii* and *Eimeria stiedii* sporozoites by treatment with bovine lactoferricin. *J Vet Med Sci* 63:187 – 190. 36. Orsi, N. (2004) The antimicrobial activity of lactoferrin: current status and perspectives. *Biometals* 17(3):189 – 196. 37. Ortiz-Estrada, G., Luna-Castro, S., Pina-Vazquez, C., Samaniego-Barron, L., Leon-Sicairos, N., Serrano-Luna, J., and de la Garza, M. (2012) Iron-saturated lactoferrin and pathogenic protozoa: could this protein be an iron source for their parasitic style of life? *Future Microbiol.* 7(1): 149 – 164. 38. Masson, P. L., Heremans, J. F., and Dive, C. (1966) An iron-binding protein common to many external secretions, *Clin. Chim. Acta* 14: 735 – 739. 39. Samuelsen, O., Haukland, H. H., Ulvatne, H., and Vorland, L. H. (2004) Anti-complement effects of lactoferrin-derived peptides. *FEMS Immunol. Med. Microbiol.* 41: 141 – 148. 40. Shin, K., Yamauchi, K., Teraguchi, S., Hayasawa, H., Tomita, M., and Otsuka, Y. (1998) Antibacterial activity of bovine lactoferrin and its peptides against enterohaemorrhagic *Escherichia coli* O157:H7. *Lett. Appl. Microbiol.* 26:407 – 411. 41. Tanaka, T., Omata, Y., and Narisawa, M. (1997) Growth inhibitory effect of bovine lactoferrin on *Toxoplasma gondii* tachyzoites in murine macrophages: role of radical oxygen and inorganic nitrogen oxide in *Toxoplasma* growth-inhibitory activity. *Vet Parasitol* 68:27 – 33. 42. Tanaka, T., Omata, Y., Saito, A., Shimazaki, K., Yamauchi, K., and Takase, M. (1995) *Toxoplasma gondii*: parasitocidal effects of bovine lactoferricin against parasites. *Exp. Parasitol.* 81: 614 – 617. 43. Tanida, T., Rao, F., Hamada, T., Ueta, E. and Osaki, T. (2001) Lactoferrin peptide increases the survival of *Candida albicans*- inoculated mice by upregulating neutrophil and macrophage functions, especially in combination with amphotericin B and granulocyte-macrophage colony-stimulating factor. *Infect. Immun.* 69: 3883 – 3890. 44. Tang, L., Cui, T., Wu, J.J., Liu-Mares, W., Huang, N., and Li, J. (2010a) A rice-derived recombinant human lactoferrin stimulates fibroblast proliferation, migration, and sustains cell survival. *Wound Repair Regen.* 18(1): 123 – 131. 45. Tang, L., Wu, J.J., Ma, Q., Cui, T., Andreopoulos, F.M., and Gil, J., (2010b) Human lactoferrin stimulates skin keratinocyte function and wound re-epithelialization. *Br. J. Dermatol.* 163(1): 38 – 47. 46. Turchany, J. M., Aley, S. B. and Gillin, F. D. (1995) Giardicidal activity of lactoferrin and N-terminal peptides. *Infect. Immun.* 63: 4550 – 4552. 47. Ueta, E., Tanida, T., and Osaki, T. (2001) A novel bovine lactoferrin peptide, FKCRRWQWRM, suppresses *Candida* cell growth and activates neutrophils. *J. Pept. Res.* 57: 240 – 249. 48. Wakabayashi, H., Bellamy, W., Takase, M., and Tomita, M. (1992) Inactivation of *Listeria monocytogenes* by lactoferricin, a potent antimicrobial peptide derived from cow's milk. *J. Food Prot.* 55: 238 – 240. 49. Wakabayashi, H., Abe, S., Teraguchi, S., Hayasawa, H., and Yamaguchi, H. (1998) Inhibition of hyphal growth of azole-resistant strains of *Candida albicans* by triazole antifungal agents in the presence of lactoferrin-related compounds. *Antimicrob. Agents Chemother.* 42: 1587 – 1591. 50. Yamniuk, A.P., Burling, H., and Vogel, H.J. (2009) Thermodynamic characterization of the interactions between the immunoregulatory proteins osteopontin and lactoferrin. *Mol. Immunol.* 46(11 – 12): 2395 – 2402. 51. Yamauchi, K., Tomita, M., Giehl, T. J., and Ellison, R. T. (1993) Antibacterial activity of lactoferrin and a pepsin-derived lactoferrin peptide fragment. *Infect. Immun.* 61: 719 – 728. 52. Yang, N., Strom, M. B., Mekonnen, S. M., Svendsen, J. S. and Redal, O. (2004) The effects of shortening lactoferrin derived peptides against tumour cells, bacteria and normal human cells. *J. Pept. Sci.* 10: 37 – 46. 53. Yoo, Y., Watanabe, S., Watanabe, R., Hata, K., Shimazaki, K., and Azuma, I. (1997) Bovine lactoferrin and lactoferricin, a peptide derived from bovine lactoferrin, inhibit tumor metastasis in mice. *Jpn. J. Cancer Res.* 88: 184 – 190. 54. Yoo, Y., Watanabe, R., Koike, Y., Mitobe, M., Shimazaki, K., and Watanabe, S. (1997) Apoptosis in human leukemic cells induced by lactoferricin, a bovine milk protein-derived peptide: involvement of reactive oxygen species. *Biochem. Biophys. Res. Commun.* 237: 624 – 628.