

Effects of Application of Temperature / Time Control System and Glucono - Delta - Lactone on Sour Meat Quality

楊喬凱、陳明造

E-mail: 364880@mail.dyu.edu.tw

ABSTRACT

The purpose of this research is to study effects of application of temperature/time control system and glucono-delta-lactone (GDL) on sour meat quality. In the experiment, two recipes (A and B) were used as the control, then separately added with GDL as the treatments (A + GDL and B + GDL) and fermented at different temperatures and times. The changes in pH value, titratable acidity, microbial counts, volatile basic nitrogen content (VBN), SDS-PAGE pattern of muscle proteins, organic acid and peptide content of the sour meat samples were analyzed. The results showed as follows: The results showed that the two sour meat samples added GDL could decline the pH value to 5.3 – a safe level which was useful to increase food safety. However, the naturally fermented sour meat samples (the control groups, without GDL) needed to take a longer time to lower the pH value to 5.3. Depending on temperature / time control system, after calculation, it was found that four group samples all met constant of the controls increased with the storage time extended, but the GDL added samples still maintained a low level of VBN. As to total microbial, lactic acid bacterial and aerobic bacterial counts for the control samples were up to 9.24 log CFU/g after fermentation, and the GDL added samples just grew to 6.71 log CFU/g. These results were noted that GDL addition could lower pH value of the sour meat in a short time, and it also could inhibited the microbial growth to extend the storage life of the sour meat. Additionally, the result of organic acid analysis, it was found acetic acid was the highest in the sour meat. However, there was slight amount of lactic acid detected in the sour meat which no lactic acid bacteria (LAB) added. This result indicated that there were LAB presented in the sour meat even it was a natural fermentation. The SDS-PAGE electrophoretogram showed the fragments of muscle proteins for the sour meat added GDL were lower than those of muscle proteins in the natural fermentation sour meat samples. It was found the peptide content was the highest in the samples fermented at 35°C for 24 hours. From these results, it can be concluded that the application of temperature / time control system and GDL to prepare the sour meat is helpful to food safety and extend its storagetime.

Keywords : sour meat、GDL、peptide、T/T control system

Table of Contents

目錄 封面內頁 簽名頁 中文摘要iii 英文摘要iv 誌謝vi 目錄viii 圖目錄xi 表目錄xiii 1. 前言 1.2. 文獻回顧 3.2.1 發酵食品 3.2.2 發酵肉原料簡介 4.2.3 發酵肉品的歷史 5.2.4 發酵肉製品的分類與特性 6.2.5 菌?(starter culture)在發酵肉品上之應用 7.2.6 蛋白質水解作用與胜?忖尾野?0 2.6.1 蛋白質之水解作用 10 2.6.2 胜?忖尾野?1 2.7 溫度 / 時間控制系統 12 2.8 葡萄糖酸內酯 13 3. 材料與方法 15 3.1 實驗藥品與儀器 15 3.1.2 材料 15 3.1.2 藥品 15 3.1.3 儀器設備 16 3.2 實驗方法 17 3.2.1 酸肉配方 17 3.2.2 酸肉製作流程 18 3.3 分析項目 19 3.3.1 酸鹼值測定 19 3.3.1.1 溫度與時間控制系統 19 3.3.2 可滴定酸測定 22 3.3.3 總生菌數測定 22 3.3.4 乳酸菌數測定 23 3.3.5 好氧菌數測定 23 3.3.6 揮發性鹽基氮(VBN)含量分析 23 3.3.7 有機酸測定 25 3.3.7.1 分析條件 25 3.3.7.2 樣品萃取 25 3.3.7.3 標準品配置 26 3.3.7.4 樣品分析 26 3.3.8 SDS-PAGE電泳分析 28 3.3.8.1 水溶性蛋白之萃取 28 3.3.8.2 鹽溶性蛋白之萃取 28 3.3.8.3 蛋白質變性流程 28 3.3.8.4 溶液配置 29 3.3.8.5 鑄膠 30 3.3.8.6 電泳條件 30 3.3.8.7 膠片染色及脫色 30 3.3.9 胜?戌t量之測定 32 3.3.9.1 胜?朴惄?G之製備 32 3.3.9.2 胜?忖妥?w32 3.4 統計分析與繪圖 34 4. 結果與討論 35 4.1 酸肉pH值的變化 35 4.2 可滴定酸 40 4.3 總生菌數、乳酸菌數以及好氧菌數之比較 44 4.4 揚發性鹽基氮(VBN)含量之比較 55 4.5 酸肉之有機酸含量 59 4.6 酸肉中蛋白質之SDS-PAGE電泳分析圖譜 65 4.7 不同酸肉配方之勝?戌t量之變化 74 5. 結論 78 參考文獻 79 圖目錄 圖3.1雙?汝邦 掃u33 圖4.1不同酸肉配方於25°C發酵期間pH之變化 36 圖4.2不同酸肉配方於30°C發酵期間pH之變化 37 圖4.3不同酸肉配方於35°C發酵期間pH之變化 38 圖4.4不同酸肉配方於25°C發酵期間滴定酸之變化 41 圖4.5不同酸肉配方於30°C發酵期間滴定酸之變化 42 圖4.6不同酸肉配方於35°C發酵期間滴定酸之變化 43 圖4.7不同酸肉配方於25°C發酵期間總生菌數之變化 46 圖4.8不同酸肉配方於30°C發酵期間總生菌數之變化 47 圖4.9不同酸肉配方於35°C發酵期間總生菌數之變化 48 圖4.10不同酸肉配方於25°C發酵期間乳酸菌數之變化 49 圖4.11不同酸肉配方於30°C發酵期間乳酸菌數之變化 50 圖4.12不同酸肉配方於35°C發酵期間乳酸菌數之變化 51 圖4.13不同酸肉配方於25°C發酵期間好氧菌數之變化 52 圖4.14不同酸肉配方於30°C發酵期間好氧菌數之變化 53 圖4.15不同酸肉配方於35°C發酵期間好氧菌數之變化 54 圖4.16不同酸肉配方於25°C發酵期間VBN值之變化 56 圖4.17不同酸肉配方於30°C發酵期間VBN值之變化 57 圖4.18不同酸肉配方於35°C發酵期間VBN值之變化 58 圖4.19不同發酵溫度製備A組配方酸肉之水溶性蛋白(WSP)SDS-PAGE電泳圖譜 66 圖4.20不同發酵溫度製備B組配方酸肉之水溶性蛋白(WSP)SDS-PAGE電泳圖譜 67

備A組配方酸肉之鹽溶性蛋白(SSP)SDS-PAGE電泳圖譜67 圖4.21不同發酵溫度製備AG組配方酸肉之水溶性蛋白(WSP)SDS-PAGE電泳圖譜68 圖4.22不同發酵溫度製備AG組配方酸肉之鹽溶性蛋白(SSP)SDS-PAGE電泳圖譜69 圖4.23不同發酵溫度製備B組配方酸肉之水溶性蛋白(WSP)SDS-PAGE電泳圖譜70 圖4.24不同發酵溫度製備B組配方酸肉之鹽溶性蛋白(SSP)SDS-PAGE電泳圖譜71 圖4.25不同發酵溫度製備BG組配方酸肉之水溶性蛋白(WSP)SDS-PAGE電泳圖譜72 圖4.26不同發酵溫度製備BG組配方酸肉之鹽溶性蛋白(SSP)SDS-PAGE電泳圖譜73 圖4.27不同酸肉配方於25 發酵期間勝?戊t量之變化75 圖4.28不同酸肉配方於30 發酵期間勝?戊t量之變化76 圖4.29不同酸肉配方於35 發酵期間勝?戊t量之變化77 表目錄表2.1乾燥香腸自然熟成之相關微生物9 表3.1恆溫過程標準21 表3.2有機酸標準品之滯留時間27 表3.3分離膠組成31 表3.4排列膠組成31 表4.1不同酸肉配方於pH5.3之恆溫標準比較39 表4.2不同配方之酸肉在不同發酵溫度與時間下草酸含量之變化60 表4.3不同配方之酸肉在不同發酵溫度與時間下甲酸含量之變化61 表4.4不同配方之酸肉在不同發酵溫度與時間下醋酸含量之變化62 表4.5不同配方之酸肉在不同發酵溫度與時間下乳酸含量之變化63 表4.6不同配方之酸肉在不同發酵溫度與時間下檸檬酸含量之變化64

REFERENCES

- 1.中國國家標準。1982。冷凍魚類檢驗法。CNS 1451 N6029。經濟部中央標準局。台北。
- 2.王宇、孔保華、李明清、夏秀芳、劉騫。2010。葡萄糖酸內酯對豬肉肌原纖維蛋白功能性的影響。食品科學。31(9):67-70。
- 3.王德法。2001。D-葡萄糖醛酸- β -X酯的開發和應用。安徽化工。2001(4):21-22。
- 4.丘志威、吳定峰。2005。食品微生物學精要 / John Garbutt原著。藝軒出版社。台北。
- 5.李敏雄、黃建雄、鍾玉明、賴喜美、蘇南雄、邱淑媛、徐岩。2006。發酵食品微生物學 / Brian J. B. Wood原著。藝軒圖書出版社。台北。
- 6.周光宏。2008。肉品加工學。中國農業出版社。北京。
- 7.岳振峰、吳暉。1998。D-葡萄糖酸- β -X酯在食品工業中的應用。食品科技。1998(6): 32-33。
- 8.林松筠。1986 菌?在發酵香腸扮演的角色。食品工業 , 18(4): 37-41。
- 9.林耕年。1990。食品營養。復文圖書有限公司。
- 10.林淑妮。2000。調節氣體包裝與添加物對減脂中式香腸品質影響之探討。靜宜大學食品營養學系碩士論文。台中。
- 11.林陽山。1993。應用乳酸菌製造肉及其產品特性之研究。國立中興大學畜產學研究所碩士論文。台中。
- 12.施明智。1996。食物學原理。藝軒圖書出版社。台北。
- 13.郭智宏。2001。腸道吸收-胺基酸與胜?式C食品工業。33(5):15-23。
- 14.陳玉真。2004。乳酸菌發酵吳郭魚保健食品產製技術及生理活性之探討。國立臺灣海洋大學食品科學系碩士學位論文。基隆。
- 15.陳怡宏。1999。生物活性勝?付峇鈴X成。食品工業 , 31(1):1-8。
- 16.陳明造、賴滋漢、許哲彰。2000。添加葡萄糖酸內酯與不同米飯類對肉品質特性和生物胺含量之影響。中國畜牧學會會誌。29(3):255-264。
- 17.陳俞伶。2005。發酵豆乳中活性勝?付坏茭H國立臺灣海洋大學食品科學系碩士學位論文。基隆。
- 18.陳勁初。1991。以乳酸菌保存食品之機制。科學與技術 , 23(9): 17-21。
- 19.黃龍男。2006。以五種不同方式處理原料豬肉對其所製肉絨品質之影響。國立屏東科技大學熱帶農業暨國際合作研究所碩士論文。屏東。
- 20.劉毓蕙。2004。水解蛋白的特性及應用食品工業 。36(2):19-24。
- 21.Chan, K. M., Decker, E. A. and Means, W. J. 1993. Extraction and activity of carnosine, a naturally occurring antioxidant in beef muscle. *J. Food Sci.*, 58: 1-4.
- 22.Church, F. C., Swaisgood, H. E., Porter, D. H. and Catignani, G. L. 1983. Spectrophotometric Assay Using o-Phthaldialdehyde for Determination of Proteolysis in Milk and Isolated Milk Proteins. *J. Dairy Sci.* 66: 1219-1227.
- 23.Eskeland B, Nordal J. 1980. Nutritional evaluation of protein in dry sausages during the fermentation process with special emphasis on amino acid digestibility. *J. Food Sci.* 45: 1153-1160.
- 24.Fadda, S., G. Oliver, and G. Vignolo. 2002. Protein degradation by *Lactobacillus plantarum* and *Lactobacillus casei* in a sausage model system. *Journal of Food Science* 67: 1179-1183.
- 25.Frister, H., Meisel, H. and Schlimme, E. 1988. OPA method modified by use of N, N-dimeyhl-2-mercaptopethylammonium chlorides as thiol component. *Fresen. J. Anal. Chem.* 330: 631-633.
- 26.Frokjaer, S. 1994. Use of hydrolysates for protein supplementation. *Food technol.* 48(10): 86-88.
- 27.Goll, D. E., Y. Otsuka, P. A. Nagainis, J. D. Shannon, S. K. Sathe and M. Muguruma. 1983. Role of muscle proteinases in maintenance of muscle integrity and mass. *J. Food Biochem.* 7: 137-177.
- 28.Grimble, G. K. and Silk, D. 1989. Peptides in human nutrition. *Nutr. Research Rev.* 2: 87-108.
- 29.Hong G.P, Ko S.H, Choi M.J, Min S.G. 2008. Effect of glucono- β -lactone and α -carrageenan combined with high pressure treatment on the physicochemical properties of restructured pork. *Meat Science*. 79(2):236-243.
- 30.Hoogenkamp, H. W. 1992. Sausage:past, present and future in vegetable protein (Technology Value in Meat Poultry and Vegetarian Foods). *Protein Technologies International Inc.* 35-40.
- 31.Incze, K. 1992. Raw fermented and dried meat products. *Fleischwirtsch.* 72:58-62.
- 32.Jay, J. M. 1996. Fermentation and fermented dairy products. In *Modern Food Microbiology*. 131-137. 4th ed. International Thomson Publishing, New York.
- 33.Kooohmaraie, M. 1992. The role of Ca++-dependent proteinases(calpains) in post-mortem proteolysis and meat tenderness. *Biochimie.* 74: 239-245.
- 34.Laemmli, U. K. 1970. Cleavage of structural proteins during the assembly of the head bacteriophage T4. *Nature* 277: 680-685.
- 35.Lahl, W. J. and Braum, S. D. 1994. Enzymatic production of protein hydrolysates for food use. *Food Technol.* 48(10): 68-71.
- 36.Lin, M. Y., and C. L. Yen. 1999. Antioxidation ability of lactic acid bacteria. *Journal of Agricultural and Food Chemistry.* 47: 1460-1466.
- 37.Lopezbote, C., T. Antequera, J. J. Cordba, C. Garocia, M. A. Asensio, and J. Ventanas. 1990. Proteolytic and lypolytic breakdown during the ripening of Iberian hams. *Processdings of 36th ICOSMT.* 3:883-887. Havana, Cuba.
- 38.Loukas, S., Varoucha, D., Zioudrou, C., Streaty, R. A. and Klee, W. A. 1983. Opioid activities and structures of α -casein-derived exorphins. *Biochemistry* 22: 4567-4573.
- 39.Lucke, F. K. 1986. Microbiological processes in the manufac- ture of dry sausage and raw ham. *Fleischwirtsch.* 66:1505- 1509.
- 40.Maijala, R. L., S. H. Herola, M. A. Aho and J. A. Hirn. 1993. The effect of GDL-induced pH decrease on the formation of biogenic amines in meat. *J. Food Prot.* 56 : 125-129.
- 41.Mandigo, R. 1991. History of meat fermentation. *Meat & poultry.* 9:12.
- 42.Nakamura, Y., N. Yamamoto. K. Sakai. A. Okubo, S. Yamazaki, and T. Takano. 1995. Antihypertensive effect of sour milk and peptides isolated from it that are inhibitors to angiotensin I-converting enzyme. *J. Dairy*

Sci. 78:1253-1257. 43.Niinivaar, F. P. 1991. Starter culture in the processing of meat by fermentation and dehydration. Proceedings 44th Annual Reciprocal meat conference. 59-63. 44.O' Halloran, G. R., D. J. Troy, D. J. Buckley, and W. J. Reville. 1997. The role of endogenous proteases in the tenderisation of fast glycolysing muscle. Meat Sci. 47: 187-210. 45.Ockerman, H. W. 1985. "Quality Control of Post-mortem Muscle Tissue". The Ohio State University and The Ohio Agricultural Research and Development Center . 1:51-53. 46.Radovanovic, R., D. Cavoski, D. Velikovic, and G. Carapic. 1990. Study of a traditional dry beef meat product: " Uzice Beef Prshuta " quantitative and qualitative Characteristics. Proceedings 36th ICoMST. 3:905-912. Havana, Cuba. 47.Sharma, U.,and Mukhopadhyay. 1992 . Processing of fermented sausage using starter culture. Processings of 38th ICoMST. 4:827-830. Clermont-Ferrand, France. 48.Smith, J. L. and S. A. Palumbo. 1981. Microorganisms as food additives. J. Food Prot. 44:936-955. 49.Smith, J. L. and S. A. Palumbo. 1983. Use of starter cultures in meats. J. Food Prot. 46:997-1006. 50.Vanderzant, C. and D. F. Splittstoesser. 1992. "Compendium of Methods for the Microbiological Examination of Foods" Third edition, The American Public Health Assoication. U.S.A. 51.Wilson George D. 1982. Fermented dry and semi-dry sausage. American Meat Institute. 1-10. 52.Zioudrou, C., Streaty, R. A. and Klee, W. A. 1979. Opioid peptides derived from food proteins. J. Bio. Chem. 254: 2446-2449.