Simultaneous Removal of Organic Matter and Production of Bacterial Cellulose from Wastewater Using Rotating Disk Reacto

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ABSTRACT

Bacterial cellulose membrane, an exopolysaccharide production by some bacterial, has unique physical and chemical properties, including well-developed surface area, high mechanical strength and ultrafine 3D network structure. In this study, the factors for bacterial cellulose production by Gluconacetobacter sp. Wu1-1 in Rotating Disks Reactor (RDR) were investigated. In the preliminary study, discs made from polycarbonate fabricated with 400C of sandpaper gave the highest result compared to others. The optimal number of disk and rotation speed for bacterial cellulose production were found to be 8 and 8 rpm. The bacterial cellulose production and COD removal rate were 0.99 g/L and 67.7%, respectively, in aeration treatment of high COD wastewater at hydraulic retention times (HRT) of 16 h. On the other hand, application of bacterial cellulose in wastewater treatment, the main focus of copper sorption from aqueous solution by bacterial cellulose was conducted in batch condition. Kinetic data and equilibrium sorption isotherms were measured. Results indicated that the sorption process follow a pseudo second-order kinetics and the Langmuir model gave a better fit to the experimental data. Additionally, the thermodynamic parameters (G, H?, S?) for the adsorption process were calculated and results suggest that the nature of adsorption is endothermic and the process is spontaneous and favorable. On the other hand, the membrane pressure was enhanced as the thickness increased, and the thin membrane has better permeability. However, the adsorption of copper was reduced when the membranes were pressed by Manual Forming Machine. In additionally, the modified unmodified of bacterial cellulose were characterized by Fourier Transfer Infrared spectroscopy (FTIR) and Scanning electron microscope (SEM).

Keywords: Bacterial cellulose membrane、Gluconacetobacter sp. Wu1-1、rotating disks reactor、chemical modification、membrane pressure

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REFERENCES

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polymer/bentonite composite as an adsorbent for the removal of copper(II) ions from aqueous solutions and electroplating industry wastewater. Industrial and Engineering Chemistry Research 16: 130-139. 26. Apilanez, I., Gutierrez, A. and Diaz, M. 1997. Effect of surface materials on initial biofilm development. Bioresource Technology 98(66): 225-230. 27. Argaman, Y., Sedlak, R. I., Eckenfelder, W. W., Daigger, G. T. and Poison, S. R. 1991. Phosphorus and Nitrogen Removal from Municipal Wastewater-Principles and Practice, 2nd Ed. Lewis Publishers. 28. Argun, M. E., Dursun, S., Ozdemir, C. and Karatas, M. 2007. Heavy metal adsorption by modified oak sawdust: thermodynamics and kinetics. Journal of Hazardous Materials B 141: 77-85. 29.Ayub, S., Ali, S. I. and Khan, N. A. 2001. Study on the removal of Cr(VI) by sugarcane bagasse from wastewater. Pollution Research Journal 2(2): 233-237. 30. Ayub, S., Ali, S. I. and Khan, N. A. 2002. Adsorption studies on the low cost adsorbent for the removal of Cr(VI) from electroplating wastewater. Environmental Pollution Control Journal 5(6): 10-20. 31.Ayub, S., Ali, S. I., Khan, N. A. and Rao, R. A. K. 1998. Treatment of wastewater by agricultural waste. Environmental Protection Control Journal 2(1): 5-8. 32.Bae, S. and Shoda, M. 2004. Bacterial cellulose production by fed-batch fermentation in molasses medium. Biotechnology Progress 20(5): 1366-1371. 33.Baral, S. S., Das, S. N. and Rath, P. 2006. Hexavalent chromium removal from aqueous solution by adsorption on treated sawdust. Biochemical Engineering Journal 31: 216-222. 34.Benziman, M. and Mazover, A. 1973. Nicotinamide adenine dinucleotide- and nicotinamide adenine dinucleotide-phosphate specific glucose-6-phosphate dehydrogenases of Acetobacter xylinum and their role in the regulation of the pentose cycle Journal of Biological Chemistry 248: 1603-1608. 35. Benziman, M., Russo, A., Hochman, S. and Weinhouse, H. 1978. Purification and regulatory properties of the oxaloacetate decarboxylase of Acetobacter xylinum. Journal of Bacteriology 134: 1-9. 36.Bhatnagar and Jain. 2005. Bhatnagar, A.K. Jain. A comparative adsorption study with different industrial wastes as adsorbents for the removal of cationic dyes from water. Journal of Colloid and Interface Science 281: 49-55 37.Bhattacharya, S., Bhat, K. K. and Raghuver, K. H. 1992. Rheology of bengal gram cicer arietinum flour suspensions. Journal of Food Engineering 17: 83-96. 38. Bickerstaff, G. F. 1997. Immobilization of Enzymes and Cells. Humana Press. 39.Bielecki, S., Krystynowicz, A., Turkiewicz, M. and Kalinowska, H. 2005.Bacterial Cellulose, Wiley-VCH Verlag, Weinheim, Germany, 2005. 40.Bjorndal, H., Erbing, C., Lindberg, B., Fahraeus, G. and G, L. 1971. Studies on the extracellular polysaccharide from Rhizobium meliloti. Acta Chemica Scandinavica 25: 1281-1286. 41.Borzani, W. and Desouza, S. J. 1995. Mechanism of the film thickness increasing during the bacterial production of cellulose on non-agitated liquid-media. Biotechnology Letters 17(11): 1271-1272. 42. Brown, A. J. 1886. On an acetic ferment which forms cellulose. Journal of the Chemical Society, Transactions 49: 432-439. 43. Budhiono, A., Rosidi, B., Taher, H. and Iguchi, M. 1999. Kinetic aspect of bacterial cellulose formation in nata-de-coco culture system. Carbohydrate Polymers 40: 137-143. 44.Bulut, Y. and Tez, Z. 2003. Removal of heavy metal ions by modified sawdust of walnut. Fresenius Environmental Bulletin 12: 1499-1504. 45. Canale-Parola, E. 1970. Biology of sugar-fermenting sarcinae. Bacteriological Reviews 34: 82-97. 46. Cannon, R. E. and Anderson, S. M. 1991. Biogenesis of BC. Critical Reviews in Microbiology 17(6): 435-447. 47. Chand, S., Aggarwal, V. K. and Kumar, P. 1994. Removal of Hexavalent Chromium from the Wastewater by Adsorption. Indian Journal of Environmental Health 36(3): 151-158. 48. Chao, Y., Mitarai, M., Sugann, Y. and Shoda, M. 2001. Effect of assition of water- soluble polysaccharides on bacterial cellulose production in a 50-L airlift reactor. Biotechnology Progress 17(4): 781-785. 49. Chauhan, D. and Sankararamakrishnan, N. 2008. Highly enhanced adsorption for decontamination of lead ions from battery wastewaters using chitosan functionalized with xanthate. Bioresource Technology 99: 9021-9024. 50. Chen, H., Dai, G. L., Zhao, J., Zhong, A.G., Wu, J. Y. and Yan, H.. 2010. Removal of copper(II) ions by a biosorbent-Cinnamomum camphora leaves powder Journal of Hazardous Materials 177: 228 – 236. 51. Chen, P. P., Wu, S. and Chong, K.-H. 2003. Surface modification of a granular activated carbon by citric acid for enhancement of copper adsorption. Carbon 41: 1979-1986. 52. Chen, S., Zou, Y., Yan, Z., Shen, W., Shi, S., Zhang, X. and Wang, H. 2009. Carboxymethylated-bacterial cellulose for copper and lead ion removal. Journal of Hazardous Materials 161: 1355-1359. 53. Chubar, N., Calvalho, J. R. and Correia, M. J. N. 2004. Heavy metals biosorption on cork biomass: effect of the pre-treatment. Colloids and Surfaces A 238: 51-58. 54. Cooper, C. D. and Alley, F. C. 1992. Air pollution contro: A design approach. P. 343-372. Centralbook, Taipei, Taiwan. 55. Daifullah, A. A. M., Girgis, B. S. and Gad, H. M. H. 2003. Utilization of agro-residues (rice husk) in small waste water treatment plans. Materials Letters 57: 1723-1731. 56. Deepak, G. and Kilambi, P. 2003. The fabrication and evaluation of the formulation variable of a controlled-porosity osmotic drug delivery system. Pharmaceutical technology 27(9): 58-68. 57. Delmer, D. P. 1987. Cellulose biosynthesis. Annual Review of Plant Physiology 32: 259-290. 58. Denizli, A., Say, R. and Arica., Y. 2000. Removal of heavy metal ions from aquatic solutions by membrane chromatography. Separation and Purification Technology 21: 181-190. 59. Dubey, S. P. and Gopal, K. 2006. Adsorption of chromium(VI) on low cost adsorbents derived from agricultural waste material: a comparative study. Journal of Hazardous Materials.: 465-470. 60. Edele, D. G. 1973. Livestock wastw management in a quality environment. University of Illinois at Urbana-Champaign. 61. Evans, B. R., ONeill, H. M., Malyvanh, V. P., Lee, I. and Woodward, J. 2003. Palladium-bacterial cellulose membranes for fuel cells. Biosensors and Bioelectronics 18: 917-923. 62. Fiedler, S., Fussel, M. and Sattler, K. 1989. Production and application of bacterial cellulose: I. A survey on state of research and nvestigations concerning fermentation kinetics. Zentralblatt fur Mikrobiologie 144: 473-484. 63. Fontana, J. D., de Sousa, A. M., Fontana, C. K., Torriani, I. L., Moreschi, J. C., Gallotti, B. J., de Sousa, S. J., Narcisco, G. P., Bichara, J. A. and Farah, L. F. 1990. Acetobacter cellulose pellicle as a temporary skin substitute. Applied Biochemistry Biotechnology. 25: 253-264. 64. Fontana, J. D., Franco, V. C., de Sousa, S. J., Lyra, I. N. and de Sousa, A. M. 1991. Nature of plant stimulators in the production of Acetobacter xylinum ('tea fungus') biofilm used in skin therapy. Applied Biochemistry Biotechnology 28(29): 341-351. 65. French, A. D. 1985. Physical and theoretical methods for determining the supramolecular structure of cellulose, in: Cellulose chemistry and its applications., R. P. Nevell and S. H. Zeronian (Eds.). Ellis Horwood Ltd., Chichester, England. 66.Ganji, M., Khosravi, M. and Rakhsaee, R. 2005. Biosorption of Pb, Cd, Cu and Zn from wastewater by treated Azolla filiculoides with H2O2/ MgCl2. International Journal of Environmental Science and

Technology 1: 265-271. 67. Geyer, U., Klemm, D. and Schmauder, H. P. 1994. Kinetics of the utilization of different C sources and the cellulose formation by Acetobacter xylinum. Acta Biotechnologica 14: 261-266. 68. Ghimire, K. N., Inoue, K., Yamaguchi, H., Makino, K. and Miyajima, T. 2003. Adsorptive separation of arsenate and arsenite anions from aqueous medium by using orange waste. Water Research 37(20): 4945-4953. 69. Gromet, Z., Schramm, M. and Hestrin, S. 1957. Synthesis of cellulose by Acetobacter xylinum. IV. Enzyme systems present in a crude extract of glucose-grown cells. Biochemical Journal 67: 679-689. 70.Gupta, V. K. and Ali, I. 2004. Removal of lead and chromium from wastewater using bagasse fly ash – a sugar industry waste. Journal of Colloid and Interface Science 271: 321-328. 71. Gupta, V. K., Gupta, M. and Sharma, S. 2001. Process development for the removal of lead and chromium from aqueous solution using red mud - an aluminum industry waste. Water Research 35(5): 1125-1134. 72. Haigler, C. H. 1985. The functions and biogenesis of native cellulose, Ellis Horwood Ltd, Chichester, England. 73. Haigler, C. H., Brown Jr., R. M. and Benziman, M. 1980. Calcoflour white ST alters the in vivo assembly of cellulose microfibrils. Science 21: 903-906. 74. Halim, S. H. A., Shehata, A. M. A. and El-Shahat, M. F. 2003. Removal of lead ions from industrial waste water by different types of natural materials. Water Research 37: 1678-1683. 75. Hanafiah, M. A. K., Ibrahim, S. C. and Yahya, M. Z. A. 2006. Equilibrium adsorption study of lead ions onto sodium hydroxide modified Lalang (Imperata cylindrica) leaf powder. Journal of Applied Polymer Science Research 2: 1169-1174. 76. Hassan, M. L. and El-Wakil, N. A. 2003. Heavy metal ion removal by amidoximated bagasse. Journal of Applied Polymer Science 87: 666-670. 77. Hestrin, S. and Satoshi, M. 1954. Synthesis of cellulose biosynthesis by Acetobacter xylinum. I. Micromethod for the determination of cellulose. Biochemical Journal 56: 2259-2262. 78. Hestrin, S. and Schramm, M. 1954. Synthesis of cellulose by Acetobacter xylinum: II. Preparation of freeze-dried cells capable of polymerizing glucose to cellulose. Biochemical Journal. 58: 345-352. 79.Ho, T. C., Chen, C., Hopper, J. R. and Oberacker, D. A. 1992. Metal capture during fluidized bed incineration of wastes contaminated with lead chloride. Combustion Science and Technology 85: 101 – 116. 80.Ho, T. C., Chen, J. M., Shukla, S. and Hopper, J. R. 1990. Metal capture during fluidized bed incineration of solid wastes. AIChE Symposium Series 276(86): 51-60. 81.Ho, T. C., Chu, H. W. and Hopper, J. R. 1993. Metal volatilization and separation during incineration. Waste Management 13: 455-466. 82. Ho, T. C., Tan, T., Chen, C. and Hopper, J. R. 1991. Characteristics of metal capture during fluidized bed incineration. AIChE Symposium Series 281(87): 118-126. 83.Ho, Y. S. and McKay, G. 1999. Pseudo-second order model for sortpion processes. Process Biochemistry 34: 451-465. 84. Ho, Y. S. and Ofamaja, A. E. 2006. Kinetic studies of copper ion adsorption on palm kernel fibre. Journal of Hazardous Materials 137: 1796-1802. 85.Hon, D. N. S. 1996. Cellulose and its Derivatives: Structures, Reactions, and Medical Uses, in: Polysaccharides in Medicinal Applications, S. Dumitru (Ed.). Marcel Dekker, New York, USA, pp. 87-105. 86. Hornung, M., Ludwigl, M., Gerrard, A. M. and Schmauderl, H.-P. 2006a. Optimizing the production of bacterial cellulose in surface culture: evaluatio of substrate mass transfer influences on the bioreaction (Part 1). Engineering in Life Sciences 6(6): 537-545. 87. Horsfall Jr., M., Abia, A. A. and Spiff, A. I. 2006. Kinetic studies on the adsorption of Cd2+, Cu2+ and Zn2+ ions from agueous solutions by cassava (Manihot sculenta Cranz) tuber bark waste. Bioresource Technology 97: 283-291. 88. Hranja, P. L., Jeso, B. D., Bareille, R., Rouais, F., Baquey, C. and Barbosa, M. A. 2006. Cellulose phosphates as biomaterials in vitro biocompatibility studies. Reactive and Functional Polymers 66: 728-739. 89. Huang, J. C. and Bates, V. T. 1980. Comparative performance of rotating biological contactors using air and pure oxygen. Journal of the Water Pollution Control Federation 52(11): 2686-2703. 90. Iguchi, M. and Yamanaka, S. 1997. Industrial use of bacterial cellulose - A review. Proceedings of International Workshop Green Polymer. Bandung-Bogor: 47-54. 91. Iguchi, M., Yamanaka, S. and Budhiono, A. 2000. Review: Bacterial cellulose - a masterpiece of nature's arts, Journal of Material Science 35(2): 261-270, 92, Ishida, T., Sugano, Y., Nakai, T. and Shoda, M. 2002, Effects of acetan on production of bacterial cellulose by Acetobacter xylinum. Bioscience, Biotechnology, and Biochemistry 66: 1677-1681. 93. Ishikawa, A., Masuoka, M., Tsuchida, T. and Yoshinaga, F. 1995. Increase in cellulose production by sulfa guanidine-resistant mutants derived from Acetobacter xylinum subsp. sucrofermentans. Bioscience, Biotechnology, and Biochemistry 59: 2259-2262. 94. Isihara, M. and Yamanaka, S. 2002. US Patent Application No. 20020065409. 95. Jonas, R. and Farab, L.F. 1998. Production and application of microbial cellulose. Polymer Degradation and Stability 59: 101-106. 96. Joris, K., Billiet, F., Drieghe, S., Brachx, D. and E, V. 1990. Microbial production of -1,4 glucan Meded. Fac. Landbouwwet Rijksuniv Gent. 55: 1563-1566. 97. Junior, O. K., Gurgel, L. V. A., de Melo, J. C. P., Botaro, V. R., Melo, T. M. S., de Freitas Gil, R. P. and Gil, L. F. 2006. Adsorption of heavy metal ion from aqueous single metal solution by chemically modified sugarcane bagasse. Bioresource Technology 98: 1291-1297. 98. Kadirvelu, K., Kavipriya, M., Karthika, C., Radhika, M., Vennilamani, N. and Pattabhi, S. 2003. Utilization of various agricultural wastes for activated carbon preparation and application for the removal of dyes and metal ions from aqueous Solution. Bioresource Technology 87: 129-132. 99. Kadirvelu, K., Thamaraiselvi, K. and Namasivayam, C. 2001a. Adsorption of nickel(II) from aqueous solution onto activated carbon prepared from coirpith. Separation and Purification Technology 24: 497-505. 100. Kadirvelu, K., Thamaraiselvi, K. and Namasivayam, C. 2001b. Removal of heavy metal from industrial wastewaters by adsorption onto activated carbon prepared from an agricultural solid waste. Bioresource Technology 76: 63-65. 101. Kamide, K., Matsuda, Y., IiJima, H. and Okajima, K. 1990. Effect of culture conditions of acetic acid bacteria on cellulose biosynthesis. British Polymer Journal 22: 167-171. 102. Keshk, S. and Sameshima, K. 2006. Influence of lignosulfonate on crystal structure and productivity of bacterial cellulose in a static culture. Enzyme and Microbial Technology 40: 4-8. 103. Khan, N. A., Ali, S. I. and Ayub, S. 2001. Effect of pH on the removal of chromium (Cr) (VI) by sugar cane baggase. Science and Technology 6: 13-19. 104. Khan, N. A., Shaaban, M. G. and Hassan, M. H. A. Removal of heavy metal using an inexpensive adsorbent. Proc. UM Research Seminar 2003 organized by Institute of Research Management and Consultancy (IPPP), University of Malaya, Kuala Lumpur. 105. Klemm, D., Schumann, D., Udhardt, U. and Marsch, S. 2001. Bacterial synthesized cellulose-artificial blood vessels for microsurgery. Progress in Polymer Science 26(9): 1561-1603. 106. Klimmek, K. S., Stan, H. J., Wilke, A., Bunke, G. and Buchholz, R. 2001. Comparative analysis of the

```
biosorption of cadmium, lead, nickel, and zinc by algae. Environmental Science and Technology 35(21): 4283-4288. 107. Kobya, M., Demirbas, E.,
Senturk, E. and Ince, M. 2005. Adsorption of heavy metal ions from aqueous solutions by activated carbon prepared from apricot stone.
Bioresource Technology 96: 1518-1521. 108. Kouda, T., Naritomi, T., Yano, H. and Yoshinaga, F. 1997. Effects of oxygen and carbon dioxide
pressures on bacterial cellulose production by Acetobacter in aerated and agitated culture. Journal of Fermentation and Bioengineering 84(2):
124-127. 109. Krystynowicz, A., Czaja, W., Wiktorowska-Jezierska, A., Goncalves-Mi?kiewicz, M., Turkiewicz, M. and Bielecki, S. 2002. Factors
affecting the yield and properties of bacterial cellulose. Journal of Industrial Microbiology and Biotechnology 29: 89-195. 110.Kuga, S., Takagi, S.
and Brown Jr., R. M. 1993. Native folded chain cellulose II. Polymer Polymer: 3293 – 3297. 111. Kula, I., U?urlu, M., Karao?lu, H. and Celik, A.
2007. Adsorption of Cd(II) ions from aqueous solutions using activated prepared from olive stone by ZnCl2 activation. Bioresource Technology
99(3): 492-501. 112. Kumar, U. and Bandyopadhyay, M. 2006. Sorption of cadm ium from aqueous solution using pretreated rice husk.
Bioresource Technology 97: 104-109. 113. Lagergren, S. 1989. About the theory of so called adsorption of soluble substances. Ksver
Veterskapsakad Hand 24: 1-6. 114. Lagergren, S. 2004. Adsorption of Congo red from aqueous solution onto calcium-rich fly ash. Journal of
Colloid and Interface Science 274: 371-379. 115.Laus, R., Costa, T.G., Szpoganicz, B. and Favere, V.T. 2010. Adsorption and desorption of
Cu(II), Cd(II) and Pb(II) ions using chitosan crosslinked with epichlorohydrin-triphosphate as the adsorbent. Journal of Hazardous Materials 183:
233-241, 116.Lee, S. H. D. and Johnson, I. J. 1980. Removal of gaseous alkali metal compounds from hot flue gas by particulate sorbents. Journal
of Engineering for Power 102: 397-402. 117. Leyva-Ramos, R., Bernal-Jacome, L. A. and Acosta-Rodriguez, I. 2005. Adsorption of cadmium(II)
from aqueous solution on natural and oxidized corncob. Separation and Purification Technology 45: 41-49. 118.Li, J. X., Hu, J., Sheng, G. D.,
Zhao, G. X. and Huang, Q. 2009. Effect of pH, ionic strength, foreign ions and temperature on the adsorption of Cu(II) from aqueous solution
toGMZ bentonite. Colloids and Surfaces A: 195-201 119.Li, N., Zhang, L., Chen, Y., Tian, Y. and Wang, H. 2011. Adsorption behavior of Cu(II)
onto titanate nanofibers prepared by alkali Treatment. Journal of Hazardous Materials 189: 265-272. 120.Li, Q., Zhai, J., Zhang, W., Wang, M.
and Zhou, J. 2006. Kinetic studies of adsorption of Pb(II), Cr(III) and Cu(II) from aqueous solution by sawdust and modified peanut husk. Journal
of Hazardous Materials B 141: 163-167. 121.Liang, S., Guo, X., Feng, N. and Tian, Q. 2010. Isotherms, kinetics and thermodynamic studies of
adsorption of Cu2+ from aqueous solutions by Mg2+/K+ type orange peel adsorbents. Journal of Hazardous Materials 174: 756-762. 122.Lin, J.,
Zhan, Y. and Zhu Z. 2011. Adsorption characteristics of copper (II) ions from aqueous solution onto humic acid-immobilized surfactant-modified
zeolite. Colloids and Surfaces A: Physicochemical and Engineering Aspects 384: 9-16. 123.Liu, M., Deng, Y., Zhan, H. and Zhang, X. 2002.
Adsorption and Desorption of Copper(II) from Solutions on New Spherical Cellulose Adsorbent. Journal of Applied Polymer Science 84: 478
- 485. 124.Loncar, E.S., Petrovic, S.E., Malbasa, R.V. and Verac, R.M. 2000. Biosynthesis of glucuronic acid by means of tea fungus.
Nahrung-Food 44: 138-139. 125.Low, K. S., Lee, C. K. and Mak, S. M. 2004. Sorption of copper and lead by citric acid modified wood. Wood
Science and Technology 38: 629-640. 126. Makhija, S. N. and Vavia, P. R. 2003. Controlled porosity osmotic pump-based controlled release
systems of pseudoephedrine: I. Cellulose acetate as a semipermeable membrane. Journal of Controlled Release 89(1): 5-18. 127.Mall, I. D.,
Srivastava, N. C. and Agarwal, N. K. 2006. Removal of Orange-G and Methyl Violet dyes by adsorption onto bagasse fly ashkinetic strdy and
equilibrium isotherm analyses. Dyes and Pigments 69: 210-223. 128. Marchetti, M., Clement, A., Loubinoux, B. and Gerardin, P. 2000.
Decontamination of synthetic solutions containing heavy metals using chemically modified sawdusts bearing polyacrylic acid chains. Journal of
Wood Science 46: 331-333, 129, Marshall, W. E., Wartelle, L. H., Boler, D. E., Johns, M. M. and Toles, C. A. 1999, Enhanced metal adsorption by
soybean hulls modified with citric acid. Bioresource Technology 69: 263-268. 130. Martin-Dupont, F., Gloaquen, V., Granet, R., Guilloton, M.,
Morvan, H. and Krausz, P. 2002. Heavy Metal Adsorption by Crude Coniferous Barks: A Modelling Study. Journal of Environmental Science and
Health 37(6): 1063-1073. 131. Masaoka, S., Ohe, T. and Sakota, N. 1993. Production of cellulose from glucose by Acetobacter xylinum. Journal of
Fermentation and Bioengineering 75(1): 18-22. 132. Matsuoka, M., Tsuchida, T., Matsushita, K., Adachi, O. and Yoshinaga, F. 1996. A synthetic
medium for bacterial cellulose production by Acetobacter xylinum subsp. sucrofermentans. Bioscience, Biotechnology, and Biochemistry 60(4):
575-579. 133. Matthysse, A. G. 1986. Initial interactions of Agrobacterium tumefaciens with plant host cells. Critical Reviews in Microbiology 13:
281-307. 134.McGarry, M. G. 1970. Alga flocculation with aluminium sulfate and polyelectrolytes. Journal of the Water Pollution Control
Federation 42: 191-201. 135. Memon, S. Q., Memon, N., Shah, S. W., Khuhawar, M. Y. and Bhanger, M. I. 2007. Sawdust – a green and
economical sorbent for the removal of cadmium(II) ions. Journal of Hazardous Materials B 139: 116-121. 136.Metcalf and Eddy, I. 1991.
Wastewater Engineering: Treatment and Reuse. 3rd ed. New York: McGraw-Hill. 137.Min, S. H., Han, J. S., Shin, E. W. and Park, J. K. 2004.
Improvement of cadmium ion removal by base treatment of juniper fiber. Water Research 38: 1289-1295. 138. Mohan, D. and Singh, K. P. 2002.
Singleand multi-component adsorption of cadmium and zinc using activated carbon derived from bagasse-an agricultural waste. Water Research
36: 2304-2318. 139. Mohanty, K., Jha, M., Meikap, B. C. and Biswas, M. N. 2005. Removal of chromium(VI) from dilute aqueous solutions by
activated carbon developed from Terminalia arjuna nuts activated with zinc chloride. Chemical Engineering Science 60: 3049-3059. 140. Moon, S.
H., Park, J. M., Chun, H. Y. and Kim, S. J. 2006. Comparison of physical properties of bacterial celluloses produced in different culture conditions
using Saccharified food wastes. Biotechnology and Bioprocess Engineering 11: 26-31. 141. Munaf, E. and Zein, R. 1997. The use of rice husk for
removal of toxic metals from wastewater. Environmental Technology 18: 359-362. 142. Nakastuka, S., Nakate, I. and Miyano, T. 1996. Drinking
water treatment by using ultrafiltration hallow membranes. Desalination 106: 55-61. 143. Namasivayam, C. and Kanchana, N. 1992. Waste
Banapith as adsorbent for colour removal from wastewater. Chemsphere 25: 1691-1705. 144. Namasivayam, C. and Sangeetha, D. 2006. Recycling
of agricultural solid waste, coirpith: removal of anions, heavy metals, organics and dyes from water by adsorption onto ZnCl2 activated coirpith
```

carbon. Journal of Hazardous Materials B 135: 449-452. 145. Namasivayam, C., Jayakumar, R. and Yaumuna, R. T. 1994. Dye Removal from wastewater by adsorption on waste Fe (III) hydroxide, Waste Management, 14 (7): 643-648. 146.Nasernejad, B., Zadeh, T. E., Pour, B. B., Bygi, M. E. and Zamani, A. 2005. Comparison for biosorption modeling of heavy metals (Cr(III), Cu(II), Zn(II)) adsorption from wastewater by carrot residues. Process Biochemistry 40: 1319-1322. 147. Nassar, M. M., Hamoda, M. F. and Radwan, G. H. 1995. Adsorption equilibria of basic dyestuff onto palm fruit particles. Water Science and Technology 32: 31-41. 148. Nishi, Y., Uryu, M., Yamanaka, S., Watanabe, K., Kitajima, K. and Iguchi, M. 1990. The structure and mechanical properties of sheets prepared from bacterial cellulose. Part 2: Improvement of the mechanical properties of sheets and their applicability to diaphragms of electro-acoustic transducers. Journal of Material Science 149. Noeline, B. F., Manohar, D. M. and Anirudhan, T. S. 2005. Kinetic and equilibrium modeling of lead(II) sorption from water and wastewater by polymerized banana stem in a batch reactor. Separation and Purification Technology 45: 131-140. 150. Ofomaja, A. E. and Naidoo, E. B. 2010. Kinetic modeling of the interaction between copper(II) and calcium hydroxide treated pine cone powder. Journal of the Taiwan Institute of Chemical Engineers 42: 480-485. 151.?ikawa, T., Morino, T. and Ameyama, M. 1995a. Production of cellulose from D-Arabitol by Acetobacter xylinum Ku-1. Bioscience, Biotechnology, and Biochemistry 59(8): 1564-1565. 152. ikawa, T., Ohtori, T. and Ameyama, M. 1995b. Production of cellulose from D-Mannitol by Acetobacter xylinum Ku-1. Bioscience, Biotechnology, and Biochemistry 59(2): 331-332. 153. Okieimen, F. E., Sogbaike, C. E. and Ebhoaye, J. E. 2005. Removal of cadmium and copper ions from aqueous solution with cellulose graft copolymers. Separation and Purification Technology 44: 85-89. 154.Oshima, T., Kondo, K., Ohto, K., Inoue, K. and Baba, Y. 2008. Preparation of phosphorylated bacterial cellulose as an adsorbent for metal ions. Reactive and Functional Polymers 68: 376-383. 155. Oshima, T., Taguchi, S., Ohe, K. and Baba, Y. 2011. Phosphorylated bacterial cellulose for adsorption of proteins. Carbohydrate Polymers 83: 953-958. 156.Ozera, A. and Pirincci, H. B. 2006. The adsorption of Cd(II) ions on sulfuric acid-treated wheat bran. Journal of Hazardous Materials B 137: 849-855. 157. Ozera, A., Ozera, D. and Ozer, A. 2004. The adsorption of copper(II) ions onto dehydrated wheat bran (DWB): determination of equilibrium and thermodynamic parameters. Process Biochemistry 39: 2183-2191. 158.Palma, L.D., Merli, C., Paris, M. and Petrucci, E. 2003. A steady-state model for the evaluation of disk rotational speed influence on RBC kinetic: model presentation. Bioresource Technology 86: 193-200. 159.Pehlivan, E., Cetin, S. and Yan?k, B. H. 2006. Equilibrium studies for the sorption of zinc and copper from aqueous solutions using sugar beet pulp and fly ash. Journal of Hazardous Materials B 135: 193-199. 160. Quesada-Chanto, A., Schroeder, A. G. S., Schmidt-Meyer, A. C., Lopez, J. A., Silveira, M. M., Jonas, R. and Naturforsch, Z. 1997. 52c: 193-196. 161.Rahman, M. S. and Islam, M. R. 2009. Effects of pH on isotherms modeling for Cu(II) ions adsorption using maple wood sawdust. Chemical Engineering Journal 149: 273-280. 162. Rehman, H., Shakirullah, M., Ahmad, I., Shah, S. and Hamedullah. 2006. Sorption studies of nickel ions onto sawdust of Dalbergia sissoo. Journal of the Chinese Chemical Society 53: 1045-1052. 163. Romano, M., Franzosi, G., Seves, A. and S, S. 1989. Study of the production of cellulose gel and cellulose by Acetobacter xylinum. Cellulose Chemistry and Technology 23: 217-223. 164. Sattler, K. and Fiedler, S. 1990. Production and application of bacterial cellulose: I. Cultivation in a rotating drum fermentor. Zentralbl Microbiol 145: 247-252. 165. Schmauder, H. P., Einfeldt, L., Geyer, U. and Klemm, D. 1992. Biosynthesis of cellulose by Acetobacter xylinum using glucose and modified carbonsources. . Meded. Fac. Landbouwwet Rijksuniv Gent 57(4a): 1797-1799. 166. Schramm, M., Gromet, Z. and Hestrin, S. 1957. Synthesis of cellulose by Acetobacter xylinum. IV Enzyme systems present in a crude extract of glucose grown cells. Biochemical Journal 67(7): 679-689. 167. S?iban, M., Klasnja, M. and Skrbi?, B. 2006. Modified softwood sawdust as adsorbent of heavy metal ions from water. Journal of Hazardous Materials B 136: 266-271, 168. Selvi, K., Pattabhi, S. and Kadirvelu, K. 2001, Removal of Cr (VI) from aqueous solution by adsorption onto activated carbon. Bioresource Technology 80: 87-89. 169. Serafica, G.C. 1997. Production of bacterial cellulose using a rotating disk film bioreactor by Acetobacter xylinum. PhD Thesis, Rensselaer Polytechnic Institute. 170. Serafica, G.C., Mormino, R. and Bungay, H.R. 2002. Inclusion of Solid Particlesin Bacterial Cellulose, Journal of Applied Microbiology and Biotechnology 58: 756 - 760. 171. Shen, W., Chen, S., Shi, S., Li, X., Zhang, X. and Hu, W. 2009a. Adsorption of Cu(II) and Pb(II) onto diethylenetriamine-bacterial cellulose. Carbohydrate Polymers 75: 110-114. 172. Shin, E. W. and Rowell, R. M. 2005. Cadmium ion sorption onto lignocellulosic biosorbent modified by sulfonation: the origin of sorption capacity improvement. Chemosphere 60: 1054-1061. 173. Shoda, M. and Sugano, Y. 2005. Recent advances in bacterial cellulose production. Biotechnology and Bioprocess Engineering 10: 1-8. 174. Shukla, S. R. and Pai, R. S. 2005a. Adsorption of Cu(II), Ni(II) and Zn(II) on dye loaded groundnut shells and sawdust. Separation and Purification Technology 43: 1-8. 175. Shukla, S. R. and Pai, R. S. 2005b. Adsorption of Cu(II), Ni(II) and Zn(II) on modified jute fibres. Bioresource Technology 96: 1430-1438. 176. Sokolnicki, A. M., Fisher, R. J., Harrah, T. P. and Kaplan, D. L. 2006. Permeability of bacterial cellulose membranes. Journal of Membrane Science 272: 15-27. 177. Somasundaran, P., Somil C. M., Yu, X. and Krishnakumar, S. 2009. Handbook of surface and colloid chemistry. In: Colloid Systems and Interfaces Stability of Di