

# 木瓜輪點病毒鞘蛋白之N端對植物系統感染之影響

吳宗哲、江主惠

E-mail: 345466@mail.dyu.edu.tw

## 摘要

木瓜輪點病毒(Papaya ringspot virus; PRSV)為RNA病毒，可對應一條複合大蛋白，其中P1和helper component-proteinase (HC-Pro)蛋白可裂解自身的C端，其他蛋白則由nuclear inclusion protein a (NIa)所剪切。由NIa蛋白的切位規則可發現NIb與鞘蛋白(coat protein; CP)間存在有兩個NIa可認知的切位，因此可對應出兩個同一讀框(open reading frame)的異質鞘蛋白，兩者之差別僅在N端的20個胺基酸，本實驗目的即探討此異質鞘蛋白的產生對病毒感染植物的影響。利用PCR突變方式共構築12個CP突變病毒株，其中8個突變株分別在第一個及第二個切位上(CP1及CP2)進行胺基酸的取代，獲得CP1QS、CP1MS、CP1GS、CP2ES、CP2MS、CP2GS、CP12QS/ES及CP12MS/GS突變病毒，另外，4個突變株分別在鞘蛋白N端進行5、10、15和20個胺基酸的刪除，得到CP 5、CP 10、CP 15及CP 20突變病毒。將野生型病毒及此12種突變病毒接種到木瓜上，其中野生型病毒35S-HA及35S-HAGFP分別在接種後第10天及第14天的木瓜上位葉出現病徵，而突變病毒CP1QS、CP1GS與CP 10則於接種後第21天才在木瓜上位葉出現病徵。摘取木瓜植株上位葉進行酵素連結免疫吸附法、RT-PCR、北方墨點法及西方墨點法分析，結果發現接種CP1QS、CP1GS、CP 5與CP 10突變株的木瓜植物皆可偵測到病毒，其他突變病毒株則未偵測到有系統感染的情形。以上結果顯示，CP1及CP2切位的改變皆會影響木瓜輪點病毒的感染能力，具較長N端的鞘蛋白對病毒系統性的感染是必需的，而且其N端第11-15個胺基酸(SNNTH)有可能是影響木瓜輪點病毒系統性感染最關鍵的區域。而CP2切位所產生的較短N端的鞘蛋白，則是病毒感染植物所必需。

關鍵詞：木瓜輪點病毒、NIa蛋白裂解?、鞘蛋白、突變、酵素連結免疫吸附法、反轉錄聚合?連鎖反應

## 目錄

封面內頁 簽名頁 中文摘要 iii 英文摘要 v 誌謝 vi 目錄 vii 圖目錄 x 表目錄 xi 1.前言 1 1.1 植物病毒感染植物的策略 1 1.2 馬鈴薯Y群病毒介紹 3 1.3 Potyvirus蛋白之特性 4 1.4 NIa蛋白裂解?之介紹 6 1.5 Potyvirus NIb蛋白?之介紹 8 1.6 potyvirus鞘蛋白的獨特性 8 1.7 木瓜輪點病毒可產生兩條大小不同之鞘蛋白 10 1.8 本實驗目的 11 2. 材料與方法 13 2.1 鞘蛋白N端刪除之突變病毒的構築 13 2.1.1 以PCR進行鞘蛋白N端刪除之突變 13 2.1.2 PCR產生的突變DNA片段構築至TA載體 15 2.1.3 勝任細胞之製備及質體的轉型作用 16 2.1.4 質體DNA之快速篩選(Quick screening) 17 2.1.5 質體DNA之純化 17 2.1.6 使用限制酵素確認質體DNA之正確性 18 2.1.7 鞘蛋白突變株之核?酸解序 18 2.1.8 突變的鞘蛋白DNA片段送入PRSV全長度質體 19 2.2 PRSV鞘蛋白突變病毒在系統性寄主植物感染力測試....20 2.3 以酵素連結免疫分析(Enzyme-Linked Immunosorbent Assay , ELISA)偵測植物中之病毒 20 2.4 以反轉錄-聚合?連鎖反應RT-PCR (Reverse Transcription- Polymerase Chain Reaction)確認植物中之病毒序列 21 2.4.1 植物總核醣核酸萃取 22 2.4.2 PRSV HA鞘蛋白突變位置之專一性引子設計 22 2.4.3 反轉錄-聚合?連鎖反應 23 2.5 北方墨點法(Northern blot)對植物中病毒之偵測 23 2.5.1 變性瓊脂糖凝膠之製備 23 2.5.2 核醣核酸之定量 24 2.5.3 變性瓊脂糖凝膠電泳之進行及轉印 24 2.5.4 RNA固定及甲基藍的定量方法 25 2.5.5 北方墨點法(Northern blot)反應 26 2.6 西方墨點法(Western blotting)偵測植物中之病毒 27 2.6.1 植物蛋白質粗萃取 (Protein extraction) 27 2.6.2 十二烷基礦酸鈉-聚丙烯醯胺膠體之配置 27 2.6.3 蛋白質電泳之進行及電轉印(Electrophoresis and electroblotting) 28 2.6.4 血清雜合與呈色(Serum hybridization and color development) 29 3.結果 30 3.1 PRSV HA CP突變株之構築 30 3.1.1 PRSV HA CP雙切位突變株之重新剪切與黏接 30 3.1.2 PRSV HA CP N端刪除突變株之構築 31 3.2 接種木瓜輪點病毒鞘蛋白突變株在木瓜植物上之病徵觀察與ELISA分析 31 3.3 RT-PCR及核?酸定序確認突變病毒株之感染 32 3.4 北方墨點法分析鞘蛋白突變株在木瓜上之感染情形 33 3.5 西方墨點法分析鞘蛋白突變株在木瓜上之感染情形 33 3.6 PRSV HA CP N端第11至15個胺基酸之點突變構築.....34 4.討論 35 5.結論 40 參考文獻 56 附錄 62 圖目錄 圖1. PRSV CP突變病毒之構築 41 圖2. 利用NCBI軟體比對11種Potyvirus屬病毒鞘蛋白胺基酸 序列 42 圖3. 利用聚合?連鎖反應進行核?酸刪除及點突變 44 圖4. 35S PRSV HA CP突變病毒株的核?酸序列確認 47 圖5. PRSV HA CP突變株接種到木瓜65天後，摘取上位葉進行反轉錄聚合?連鎖反應偵測病毒之感染 48 圖6. 北方墨點法偵測木瓜輪點病毒鞘蛋白突變株在木瓜上之 感染情形 49 圖7. 西方墨點法偵測木瓜輪點病毒鞘蛋白突變株在木瓜上之 感染情形 50 圖8. 針對35S PRSV HA CP N端10-15個胺基酸之突變病毒株的核?酸序列確認 51 表目錄 表1. PCR突變所使用之引子。 52 表2. 木瓜輪點病毒鞘蛋白上之核?酸及其對應的胺基酸突變 53 表3. 木瓜輪點病毒鞘蛋白突變株在寄主植物上之感染力分析 54 表4. 木瓜輪點病毒鞘蛋白N端之10-15個胺基酸突變 55

## 參考文獻

林碧雲. (2010). 木瓜輪點病毒鞘蛋白雙重切位突變病毒株系之感染力分析. 大葉大學分子生物科技學系碩士班。 Adams, M.J., Antoniw, J.F., and Beaudoin, F. (2005). Overview and analysis of the polyprotein cleavage sites in the family Potyviridae. *Mol Plant Pathol* 6, 471-487.

Ala-Poikela, M., Goytia, E., Haikonen, T., Rajamaki, M.L., and Valkonen, J.P. (2011). Helper Component Proteinase of the Genus Potyvirus Is an Interaction Partner of Translation Initiation Factors eIF(iso)4E and eIF4E and Contains a 4E Binding Motif. *J Virol* 85, 6784-6794.

Benitez-Alfonso, Y., Faulkner, C., Ritzenthaler, C., and Maule, A.J. (2010). Plasmodesmata: Gateways to Local and Systemic Virus Infection. *Mol Plant Microbe Interact* 23, 1403-1412. Blanc, S., Ammar, E.D., Garcia-Lampasona, S., Dolja, V.V., Llave, C., Baker, J., and Pirone, T.P. (1998). Mutations in the potyvirus helper component protein: effects on interactions with virions and aphid stylets. *J Gen Virol* 79 ( Pt 12), 3119-3122.

Carrington, J.C., Haldeman, R., Dolja, V.V., and Restrepo-Hartwig, M.A. (1993). Internal cleavage and trans-proteolytic activities of the VPg-proteinase (N1a) of tobacco etch potyvirus in vivo. *J Virol* 67, 6995-7000. Chiang, C.H., and Yeh, S.D. (1997). Infectivity assays of in vitro and in vivo transcripts of papaya ringspot potyvirus. *Bot. Bull. Acad. Sin.* 38, 153-163. Christie, R.G., and Edwardson, J.R. (1977). Light and electron microscopy of plant virus inclusions. *F1. Agric. Exp. St. Monogr*, 150. Chung, B.Y., Miller, W.A., Atkins, J.F., and Firth, A.E. (2008). An overlapping essential gene in the Potyviridae. *Proc Natl Acad Sci U S A* 105, 5897-5902. Dolja, V.V., Haldeman, R., Robertson, N.L., Dougherty, W.G., and Carrington, J.C. (1994). Distinct functions of capsid protein in assembly and movement of tobacco etch potyvirus in plants. *Embo J* 13, 1482-1491. Dougherty, W.G., Cary, S.M., and Parks, T.D. (1989). Molecular genetic analysis of a plant virus polyprotein cleavage site: a model. *Virology* 171, 356-364. Edwardson, J.R. (1974). Some properties of the potato virus Y group. *F1. Agric. Exp. St. Monogr* 4, 225. Ghabrial, S.A., Smith, H.A., Parks, T.D., and Dougherty, W.G. (1990). Molecular genetic analyses of the soybean mosaic virus N1a proteinase. *J Gen Virol* 71 ( Pt 9), 1921-1927. Hajimorad, M.R., Ding, X.S., Flasinski, S., Mahajan, S., Graff, E., Haldman-Cahill, R., Carrington, J.C., and Cassidy, B.G. (1996). N1a and N1b of peanut stripe potyvirus are present in the nucleus of infected cells, but do not form inclusions. *Virology* 224, 368-379. Hamilton, R.I., Edwardson, J.R., Francki, R.I.B., Hsu, H.T., Hull, R., Koenig, R., and Milne, R.G. (1981). Guidelines for the Identification and Characterization of Plant Viruses. *J Gen Virol* 54, 223-241. Harrison, B.D., and Robinson, D.J. (1988). Molecular variation in vector-borne plant viruses: epidemiological significance. *Philos Trans R Soc Lond B Biol Sci* 321, 447-462. Hong, Y., and Hunt, A.G. (1996). RNA Polymerase Activity Catalyzed by a Potyvirus-Encoded RNA-Dependent RNA Polymerase. *Virology* 226, 146-151. Ju, H.-J., Samuels, T.D., Wang, Y.-S., Blancaflor, E., Mark Payton, R.M., Krishnamurthy, K., Nelson, R.S., and Verchot-Lubicz, J. (2005). The Potato Virus X TGBp2 Movement Protein Associates with Endoplasmic Reticulum-Derived Vesicles during Virus Infection. *Plant Physiology* 138, 1877-1895. Karpova, O.V., Zayakina, O.V., Arkhipenko, M.V., Sheval, E.V., Kiselyova, O.I., Poljakov, V.Y., Yaminsky, I.V., Rodionova, N.P., and Atabekov, J.G. (2006). Potato virus X RNA-mediated assembly of single-tailed ternary 'coat protein-RNA-movement protein' complexes. *J Gen Virol* 87, 2731-2740.

Klein, P.G., Klein, R.R., Rodriguez-Cerezo, E., Hunt, A.G., and Shaw, J.G. (1994). Mutational analysis of the tobacco vein mottling virus genome. *Virology* 204, 759-769. Krishnamurthy, K., Heppeler, M., Mitra, R., Blancaflor, E., Payton, M., Nelson, R.S., and Verchot-Lubicz, J. (2003). The Potato virus X TGBp3 protein associates with the ER network for virus cell-to-cell movement. *Virology* 309, 135-151. Laliberte, J.-F., Nicolas, O., Chatel, H., Lazure, C., and Morosoli, R. (1992). Release of a 22-kDa protein derived from the amino-terminal domain of the 49-kDa N1a of turnip mosaic potyvirus in Escherichia coli. *Virology* 190, 510-514. Lan, P., Yeh, W.B., Tsai, C.W., and Lin, N.S. (2010). A unique glycine-rich motif at the N-terminal region of Bamboo mosaic virus coat protein is required for symptom expression. *Mol Plant Microbe Interact* 23, 903-914.

Li, X.H., Valdez, P., Olvera, R.E., and Carrington, J.C. (1997). Functions of the tobacco etch virus RNA polymerase (N1b): subcellular transport and protein-protein interaction with VPg/proteinase (N1a). *J Virol* 71, 1598-1607. Lin, L., Luo, Z., Yan, F., Lu, Y., Zheng, H., and Chen, J. (2011). Interaction between potyvirus P3 and ribulose-1,5-bisphosphate carboxylase/oxygenase (RubisCO) of host plants. *Virus Genes* 43, 90-92. Lin, S.S., Hou, R.F., and Yeh, S.D. (2001). Complete genome sequence and genetic organization of a Taiwan isolate of Zucchini yellow mosaic virus. *Bot. Bull. Acad. Sin.* 42, 243-250. Maia, I.G., Haenni, A., and Bernardi, F. (1996). Potyviral HC-Pro: a multifunctional protein. *J Gen Virol* 77 ( Pt 7), 1335-1341. Ozeki, J., Hashimoto, M., Komatsu, K., Maejima, K., Himeno, M., Senshu, H., Kawanishi, T., Kagiwada, S., Yamaji, Y., and Namba, S. (2009). The N-terminal region of the *Plantago asiatica* mosaic virus coat protein is required for cell-to-cell movement but is dispensable for virion assembly. *Mol Plant Microbe Interact* 22, 677-685. Peng, Y.H., Kadoury, D., Gal-On, A., Huet, H., Wang, Y., and Raccah, B. (1998). Mutations in the HC-Pro gene of zucchini yellow mosaic potyvirus: effects on aphid transmission and binding to purified virions. *J Gen Virol* 79 ( Pt 4), 897-904. Puustinen, P., and Mäkinen, K. (2004). Uridylation of the potyvirus VPg by viral replicase N1b correlates with the nucleotide binding capacity of VPg. *J Biol Chem* 279, 38103-38110. Puustinen, P., Rajamäki, M.L., Ivanov, K.I., Valkonen, J.P., and Mäkinen, K. (2002). Detection of the potyviral genome-linked protein VPg in virions and its phosphorylation by host kinases. *J Virol* 76, 12703-12711. Quemada, H., L'Hostis, B., Gonsalves, D., Reardon, I.M., Heinrikson, R., Hiebert, E.L., Sieu, L.C., and Slightom, J.L. (1990). The nucleotide sequences of the 3'-terminal regions of papaya ringspot virus strains W and P. *J Gen Virol* 71, 203-210. Revers, F., Le Gall, O., Candresse, T., and Maule, A.J. (1999). New Advances in Understanding the Molecular Biology of Plant/Potyvirus Interactions. *Mol Plant Microbe Interact* 12, 367-376.

Rodriguez-Cerezo, E., Findlay, K., Shaw, J.G., Lomonosoff, G.P., Qiu, S.G., Linstead, P., Shanks, M., and Risco, C. (1997). The coat and cylindrical inclusion proteins of a potyvirus are associated with connections between plant cells. *Virology* 236, 296-306. Rojas, M.R., Zerbini, F.M., Allison, R.F., Gilbertson, R.L., and Lucas, W.J. (1997). Capsid protein and helper component-proteinase function as potyvirus cell-to-cell movement proteins. *Virology* 237, 283-295. Samuels, T.D., Ju, H.J., Ye, C.M., Motes, C.M., Blancaflor, E.B., and Verchot-Lubicz, J. (2007). Subcellular targeting and interactions among the Potato virus X TGB proteins. *Virology* 367, 375-389. Shukla, D.D., and Ward, C.W. (1989). Identification and classification of potyviruses on the basis of coat protein sequence data and serology. Brief review. *Arch Virol* 106, 171-200.

Taliansky, M., Torrance, L., and Kalinina, N.O. (2008). Role of plant virus movement proteins. *Methods Mol. Biol.* 451, 33-54. Tatineni, S., Van Winkle, D.H., and French, R. (2010). The N-terminal region of wheat streak mosaic virus coat protein is a host- and strain-specific long-distance transport factor. *J Virol* 85, 1718-1731. Torrance, L., Andreev, I.A., Gabrenaite-Verhovskaya, R., Cowan, G., Makinen, K., and Taliansky, M.E. (2006). An unusual structure at one end of potato potyvirus particles. *J Mol Biol* 357, 1-8. Urcuqui-Inchima, S., Haenni, A.L., and Bernardi, F. (2001). Potyvirus proteins: a wealth of functions. *Virus Res* 74, 157-175. van Regenmortel, M.H.V., Fauquet, C.M., Bishop, D.H.L., Carstens, E.B., Estes, M.K., Lemon, S.M., Maniloff, J., Mayo, M.A., McGeoch, D.J., Pringle, C.R., and Wickner, R.B. (2000). Virus taxonomy: classification and nomenclature of viruses. Seventh Report of the International Committee on Taxonomy of Viruses. (San Diego: Academic Press). Varrelmann, M., and Maiss, E. (2000). Mutations in the coat protein gene of plum pox virus suppress particle assembly, heterologous encapsidation and complementation in transgenic plants of *Nicotiana benthamiana*. *J Gen Virol* 81, 567-576. Verchot-Lubicz, J. (2005). A New Cell-to-Cell Transport Model for Potexviruses. *Mol Plant Microbe Interact* 18, 283-290. Verchot-Lubicz, J., Ye, C.M., and Bamunusinghe, D. (2007). Molecular biology of potexviruses: recent advances. *J Gen Virol* 88, 1643-1655. Wei, T., Zhang, C., Hong, J., Xiong, R., Kasschau, K.D., Zhou, X., Carrington, J.C., and Wang, A. (2010). Formation of complexes at plasmodesmata for potyvirus intercellular movement is mediated by the viral protein P3N-PIPO. *PLoS Pathog* 6, e1000962. Wen, R.H., and Hajimorad, M.R. (2010). Mutational analysis of the putative pipo of soybean mosaic virus suggests disruption of PIPO protein impedes movement. *Virology* 400, 1-7. Yeh, S.D., Gonsalves, D., and Provvidenti, R. (1984). Comparative Studies on Host Range and Serology of Papaya Ringspot Virus and Watermelon Mosaic Virus 1. *Phytopathology* 74(9), 1081-1085. Yeh, S.D., Jan, F.J., Chiang, C.H., Doong, T.J., Chen, M.C., Chung, P.H., and Bau, H.J. (1992). Complete nucleotide sequence and genetic organization of papaya ringspot virus RNA. *J Gen Virol* 73 ( Pt 10), 2531-2541.