

Purification and characterization of chitinase produced by *Aeromonas* sp.

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ABSTRACT

In this study, 11 strains were screened from soil all over Taiwan to produce chitinases. Among them, strains DN15 and DN23 produced higher level of reducing sugars. According to the DNA sequences of these two strains identified by the NCBI (National Center for Biotechnology Information), Strains DN15 and DN23 were then named as *Aeromonas hydrophila* DYU-Too15 and *Aeromonas punctata* DYU-Too16. The β -chitin content and nitrogen species in the CB (chitin broth) media were altered to search for a better culture condition to produce N-acetylglucosamine. In a CB medium, when the content of β -chitin was altered, there was no effect on the variety of N-acetylglucosamine produced by *A. hydrophila* DYU-Too15 or *A. punctata* DYU-Too16, but the content of chitooligosaccharides increased with the increase of the β -chitin concentration. If the medium contained 4%

-chitin, the highest amount of N-acetylglucosamine (about 3.65 g/L) was produced by *A. hydrophila* DYU-Too15 at 96 h. When the medium contained 5% β -chitin, N-acetylchitotriose was the major product and reached about 1.22 g/L at 96 h. There was no effect on the variety of N-acetylglucosamine produced by *A. hydrophila* DYU-Too15 in a CB medium with various nitrogen sources (yeast extract, peptone, tryptone, yeast extract + peptone, NH4Cl). The major product was N-acetylglucosamine. For *A. punctata* DYU-Too16 in a CB medium with yeast extract and peptone as the nitrogen sources, the major products included N-acetylglucosamine and N-acetylchitotriose. However, the only major product was N-acetylglucosamine if each of the other four nitrogen sources was used as the nitrogen source in the CB medium. In order to obtain N-acetylchitooligosaccharides, the newly screened two strains and the early obtained strain, *Aeromonas hydrophila* DYU-Too14, in our laboratory were examined. *A. hydrophila* DYU-Too14 can produce N-acetylchitopentaose and N-acetylchitohexaose in a CB medium by using NH4Cl as a nitrogen source. Since N-acetylchitopentaose and N-acetylchitohexaose can enhance the immune system, inhibit tumor cell growth and possess other physiological activities, and their values are much higher than N-acetylglucosamine and N-acetylchitotriose produced by the other two strains. Thereafter, *A. hydrophila* DYU-Too14 was used as the target strain for chitinase purification. To separate the chitinase produced by *A. hydrophila* DYU-Too14, this strain was cultivated in a CB medium containing 4% β -chitin as the carbon source and 0.7 g/L NH4Cl as the nitrogen source. The supernatant of the culture containing crude enzyme was first precipitated by ammonium sulfate, and then the precipitate was further purified through dialysis, anion gel (DEAE-Sepharose) chromatography. From DEAE-Sepharose gel chromatographic diagram, two peaks of Fractions 90-93 and 94-98 possessed chitinase activity. Hence, the above chitinase was used to hydrolyze colloidal chitin solution, the hydrolysates were separated through centrifuge and lyophilization, and its composition was analyzed by HPLC. The hydrolysates contained N-acetylchitopentaose and N-acetylchitohexaose. Through electrophoresis, the molecular weight of the chitinase was identified to be 25 kDa.

Keywords : chitinase、*Aeromonas hydrophila* DYU-Too14、N-acetylglucosamine、N-acetylchitotriose、N-acetylchitopentaose、N-acetylchitohexaose

Table of Contents

封面內頁 簽名頁 授權書 iii 中文摘要 iv 英文摘要 vi 致謝 vii 目錄 viii 圖目錄 xii 表目錄 xv 1. 緒論 1 2. 文獻回顧 2 2.1 幾丁質 2 2.2 N-乙醯幾丁寡醣相關衍生物與應用 2 2.2.1 抗菌活性 2 2.2.2 免疫活性 4 2.2.3 基因輸送載體 6 2.2.4 藥物輸送載體 7 2.3 N-乙醯幾丁寡醣的製備 9 2.3.1 化學法 9 2.3.2 酶素法 10 2.4 N-乙醯幾丁寡醣的分離與純化 11 2.4.1 膠體過濾層析法 11 2.4.2 離子交換層析法 12 3.材料方法 13 3.1 實驗藥品 13 3.2 實驗器材 14 3.3 實驗試劑 15 3.3.1 培養基組成 15 3.3.2 膠態幾丁質之製備 17 3.3.3 McIlvaine buffer之配製 17 3.3.4 呈色劑之配置 17 3.4 實驗方法 17 3.4.1 菌株篩選、保存及活化 19 3.4.2 幾丁質分解？ “吨懶R 19 3.4.3 還原醣含量之測定 21 3.4.4 蛋白質濃度測定 21 3.4.5 幾丁質水解產物之HPLC分析 21 3.4.6 分離純化幾丁質？ 22 3.4.7 聚丙烯醯胺膠體電泳分析 24 4.結果與討論 27 4.1 菌株於膠態幾丁質培養基生長情形 27 4.2 分解幾丁質菌株之篩選 27 4.2.1 菌株 *Aeromonas hydrophila* DYU-Too15 32 4.2.2 菌株 *Aeromonas punctata* DYU-Too16 32 4.2.3 菌株 *Aeromonas* sp. DYU-Too14之特性 32 4.3 菌株培養於不同含量 β -幾丁質之CB培養基 35 4.3.1 菌株 *A. hydrophila* DYU-Too15 35 4.3.1.1 幾丁質？ “吨坐懶R 35 4.3.1.2 還原醣量與pH值變化 35 4.3.1.3 幾丁質水解產物分析 37 4.3.2 菌株 *A. punctata* DYU-Too16 37 4.3.2.1 幾丁質？ “吨坐懶R 42 4.3.2.2 還原醣量與pH值變化 42 4.3.2.3 幾丁質水解產物分析 42 4.4 不同氮源培養菌株 46 4.4.1 菌株 *A. hydrophila* DYU-Too15 46 4.4.1.1 幾丁質？ “吨坐懶R 46 4.4.1.2 還原醣量與pH值變化 51 4.4.1.3 幾丁質水解產物分析 51 4.4.2 菌株 *A. punctata* DYU-Too16 55 4.4.2.1 幾丁質？ “吨坐懶R 55 4.4.2.2 還原醣量與pH值變化 55 4.4.2.3 幾丁質水

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