

以超音波輔助酵素合成咖啡酸苯乙酯之最優化研究

陳曉菁、謝淳仁、?耀國

E-mail: 319700@mail.dyu.edu.tw

摘要

酚酸化合物已被證實對人體具有抗發炎(anti-inflammatory)、抗氧化(anti-oxidant)等功效。自然界大量存在之咖啡酸(caffeic acid)為水溶性多酚類，其酯類衍生物的親油性高，可強化其抗氧化能力，並提高其應用價值。利用脂解酵素(lipase)合成酚酸酯類，藉此改變酚酸的親油特性，使抗氧化及自由基清除力提高的功能更穩定。脂解酵素可催化咖啡酸及苯乙醇(phenyl ethanol)合成咖啡酸苯乙酯(caffeic acid phenethyl ester, CAPE)，酵素合成比起化學合成方式不僅更天然，且反應條件溫和，製程對環境較友善，所合成出來的產物亦偏向天然，在食品或化妝品的應用上，對於消費者之接受度較高。為了縮短催化酚酸酯化物(phenolic acid ester)的合成時間，本研究藉由生物催化(biocatalysis)技術並配合超音波(ultrasound)系統及連續式酵素填充床生物反應器(continuous packed-bed bioreactor)進行催化合成反應，以符合未來工業量產之應用。將基質咖啡酸與苯乙醇(phenyl ethanol)於異辛烷(isooctane)溶劑中，利用固定化脂解酵素(NovozymR 435)，經由中心混層實驗設計(central composite rotatable design, CCRD)或Box-Behnken實驗設計(Box-Behnken design)，以直接酯化(direct esterification)方式合成咖啡酸苯乙酯，再以反應曲面法(response surface methodology, RSM)進行分析，分別探討合成反應參數對莫耳轉換率的影響，最後，利用脊型分析(analysis of ridge max)詳細探討酵素合成咖啡酸苯乙酯之最優化反應條件。本研究分成三階段進行，第一階段為利用固定化酵素NovozymR 435合成咖啡酸苯乙酯，使提高咖啡酸在親油性環境下之穩定性，增加其有效利用率。將咖啡酸及苯乙醇以直接酯化方式，並利用五階層四變數之中心混層實驗設計，探討合成咖啡酸苯乙酯之最優化條件。實驗結果顯示，反應時間59小時、反應溫度69 °C、咖啡酸與苯乙醇莫耳比1:72及酵素用量351 PLU (propyl laurate unit)，咖啡酸苯乙酯莫耳轉換率之理論值與實驗值分別為91.36%及91.65 ± 0.66%。第二階段藉由超音波輔助系統合成咖啡酸苯乙酯。超音波於液相溶液中形成空穴現象(cavitation)，產生許多微小氣泡及超臨界區域，提供了化學介質力量，使增加了酵素與基質間作用的接觸機會，進而提高合成酯化作用，縮短咖啡酸苯乙酯合成時間。結果顯示，超音波輔助系統利用NovozymR 435於反應溫度70 °C，合成咖啡酸苯乙酯之最優化條件為：反應時間9.6小時、咖啡酸與苯乙醇莫耳比1:71、酵素用量2938 PLU及超音波功率為2 W/cm²，咖啡酸苯乙酯莫耳轉換率之理論值與實驗值分別為96.03%及93.08 ± 0.42%。第三階段為超音波輔助系統結合連續式酵素填充床生物反應器合成咖啡酸苯乙酯，利用超音波加速酵素作用並在連續式生物反應器下，模擬工業連續化生產流程，提供業界大量製備之依據。結果顯示，當咖啡酸與苯乙醇莫耳比1:100，填充15,000 PLU固定化酵素NovozymR 435於連續式酵素填充床生物反應器，合成咖啡酸苯乙酯之最優化條件為：反應溫度72.66 °C、體積流量0.046 mL/min及超音波功率為1.64 W/cm²，莫耳轉換率之理論值與實驗值分別為97.84%及92.11 ± 0.75%。綜合以上實驗結果，利用固定化酵素NovozymR 435合成咖啡酸苯乙酯，並在超音波輔助系統下，克服了酚酸酯類合成耗時的障礙。藉由連續式生物反應器，達到連續生產咖啡酸苯乙酯之目的，探討合成咖啡酸苯乙酯之最優化條件，找出最佳莫耳轉換率，節省操作時間及能源，進一步提供予業界量化的參考。

關鍵詞：生物反應器、生物催化、咖啡酸苯乙酯、脂解酵素、酚酸、超音波、最優化

目錄

封面內頁 簽名頁 授權書iii 中文摘要iv 英文摘要vii 誌謝ix 目錄x 圖目錄xiv 表目錄xvii 1.緒論1 2.文獻回顧5 2.1酚酸酯類之酵素合成5 2.1.1酚類化合物的定義及抗氧化作用5 2.1.2蜂膠與咖啡酸苯乙酯8 2.1.3脂解酵素10 2.1.3.1酵素之優點10 2.1.3.2酵素之固定化優點及應用11 2.1.3.3NovozymR 435介紹12 2.1.4酚酸酯化物之合成13 2.1.4.1酚酸酯類化學合成14 2.1.4.2酚酸酯類酵素合成15 2.1.5反應曲面法之應用17 2.2超音波於酵素合成酯類之研究18 2.2.1超音波的定義及特點18 2.2.2超音波應用型態簡介19 2.2.3超音波水解及合成酯類22 2.3連續式酵素填充床生物反應器於酵素合成酯類之研究24 2.3.1生物反應器介紹24 2.3.2連續式酵素填充床生物反應器之酯類合成26 3.以反應曲面法探討酵素合成咖啡酸苯乙酯之最優化反應條件28 3.1摘要28 3.2前言29 3.3實驗材料33 3.3.1藥品33 3.3.2儀器設備33 3.4實驗設計與方法34 3.4.1實驗設計34 3.4.2咖啡酸苯乙酯之合成方法34 3.4.3高效能液相層析之分析方法34 3.4.4咖啡酸苯乙酯之莫耳轉換率35 3.5結果與討論37 3.5.1反應時間對NovozymR 435催化合成咖啡酸苯乙酯莫耳轉換率之影響37 3.5.2NovozymR 435合成咖啡酸苯乙酯之變數分析39 3.5.3NovozymR 435合成咖啡酸苯乙酯之最優化探討48 4.以反應曲面法探討超音波輔助合成咖啡酸苯乙酯之最優化反應條件52 4.1摘要52 4.2前言53 4.3實驗材料56 4.3.1藥品56 4.3.2儀器設備56 4.4實驗設計與方法57 4.4.1實驗設計57 4.4.2咖啡酸苯乙酯之合成方法57 4.4.3高效能液相層析之方法分析57 4.5結果與討論59 4.5.1超音波輔助對酵素催化合成咖啡酸苯乙酯莫耳轉換率之影響59 4.5.2反應時間及酵素用量對酵素催化合成咖啡酸苯乙酯莫耳轉換率之影響59 4.5.3超音波功率對酵素合成咖啡酸苯乙酯莫耳轉換率之影響60 4.5.4超音波輔助合成咖啡酸苯乙酯之變數分析64 4.5.5超音波輔助合成咖啡酸苯乙酯之

最優化探討73 5.以反應曲面法探討連續式酵素填充床生物反應器合成咖啡酸苯乙酯之最優化反應條件77 5.1摘要77 5.2前言78 5.3實驗材料82 5.3.1藥品82 5.3.2儀器設備82 5.4實驗設計與方法83 5.4.1實驗設計83 5.4.2咖啡酸苯乙酯合成方法83 5.4.3高效能液相層析之分析方法84 5.5結果與討論86 5.5.1流速及超音波功率對合成咖啡酸苯乙酯莫耳轉換率之影響86 5.5.2連續式酵素填充床生物反應器合成咖啡酸苯乙酯之變數分析89 5.5.3連續式酵素填充床生物反應器合成咖啡酸苯乙酯之最優化探討97 5.5.4酵素再利用率98 6.結論105 參考文獻108 附錄121 圖1.1整體研究架構圖4 圖2.1酚類化合物之分類6 圖2.2抗氧化之酚酸結構7 圖2.3咖啡酸苯乙酯之化學結構式9 圖2.4超音波生物反應器之型態21 圖3.1NovozymR 435催化咖啡酸與苯乙醇生成咖啡酸苯乙酯之酯化反應31 圖3.2NovozymR 435催化咖啡酸及苯乙醇合成咖啡酸苯乙酯之實驗架構圖32 圖3.3反應時間對酵素合成咖啡酸苯乙酯莫耳轉換率之影響38 圖3.4NovozymR 435合成咖啡酸苯乙酯之莫耳轉換率實驗值與預測值之線性關係43 圖3.5反應時間及基質莫耳比對酵素合成咖啡酸苯乙酯莫耳轉換率之反應曲面圖45 圖3.6反應時間及酵素用量對酵素合成咖啡酸苯乙酯莫耳轉換率之反應曲面圖46 圖3.7基質莫耳比及酵素用量對酵素合成咖啡酸苯乙酯莫耳轉換率之反應曲面圖47 圖3.8NovozymR 435合成咖啡酸苯乙酯莫耳轉換率之等高線曲面圖50 圖4.1超音波簡要裝置54 圖4.2超音波輔助咖啡酸及苯乙醇合成咖啡酸苯乙酯之實驗架構圖55 圖4.3比較超音波輔助及機械式振盪對於酵素催化合成咖啡酸苯乙酯之影響61 圖4.4反應時間及酵素用量對超音波輔助合成咖啡酸苯乙酯莫耳轉換率之影響62 圖4.5超音波功率對合成咖啡酸苯乙酯莫耳轉換率之影響63 圖4.6超音波輔助合成咖啡酸苯乙酯莫耳轉換實驗值與預測值之線性關係68 圖4.7反應時間及基質莫耳比對超音波輔助合成咖啡酸苯乙酯莫耳轉換率之反應曲面圖70 圖4.8基質莫耳比及酵素用量對超音波輔助合成咖啡酸苯乙酯莫耳轉換率之反應曲面圖71 圖4.9基質莫耳比及超音波功率對超音波輔助合成咖啡酸苯乙酯莫耳轉換率之反應曲面圖72 圖4.10超音波輔助合成咖啡酸苯乙酯莫耳轉換率之等高線曲線圖75 圖5.1連續式酵素填充床生物反應器裝置80 圖5.2連續式酵素填充床生物反應器催化咖啡酸及苯乙醇合成咖啡酸苯乙酯之實驗架構圖81 圖5.3流速及超音波對連續式酵素填充床生物反應器合成咖啡酸苯乙酯莫耳轉換率之影響87 圖5.4超音波功率對連續式酵素填充床生物反應器合成咖啡酸苯乙酯莫耳轉換率之影響88 圖5.5連續式酵素填充床生物反應器對合成咖啡酸苯乙酯莫耳轉換率之實驗值與預測值之線性關係93 圖5.6反應溫度及流速對連續式酵素填充床生物反應器合成咖啡酸苯乙酯莫耳轉換率之反應曲面圖95 圖5.7反應溫度及超音波功率對連續式酵素填充床生物反應器合成咖啡酸苯乙酯莫耳轉換率之反應曲面圖96 圖5.8不同反應溫度對連續式酵素填充床生物反應器合成咖啡酸苯乙酯莫耳轉換率之等高線曲線圖100 圖5.9不同流速對連續式酵素填充床生物反應器合成咖啡酸苯乙酯莫耳轉換率之等高線曲線圖101 圖5.10不同功率對連續式酵素填充床生物反應器合成咖啡酸苯乙酯莫耳轉換率之等高線曲線圖102 圖5.11酵素在超音波系統及連續式酵素填充床生物反應器合成咖啡酸苯乙酯之再用104 表3.1酵素合成咖啡酸苯乙酯實驗設計反應參數實驗值之範圍36 表3.2NovozymR 435合成咖啡酸苯乙酯之五階層四變數中心混層實驗設計及其莫耳轉換率41 表3.3NovozymR 435合成咖啡酸苯乙酯莫耳轉換率之變異分析42 表3.4NovozymR 435合成咖啡酸苯乙酯變數之聯合檢測分析44 表3.5脊型分析評估NovozymR 435合成咖啡酸苯乙酯之莫耳轉換最大值51 表4.1超音波輔助合成咖啡酸苯乙酯之實驗設計反應參數實驗值範圍58 表4.2超音波輔助合成咖啡酸苯乙酯之五階層四變數中心混層實驗設計及其莫耳轉換率66 表4.3超音波輔助合成咖啡酸苯乙酯莫耳轉換率變數之變異分析67 表4.4超音波輔助合成咖啡酸苯乙酯變數之聯合檢測分析69 表4.5脊型分析評估超音波輔助合成咖啡酸苯乙酯之莫耳轉換率最大值76 表5.1連續式酵素填充床生物反應器合成咖啡酸苯乙酯三階層三變數之實驗設計反應參數實驗值範圍85 表5.2連續式酵素填充床生物反應器合成咖啡酸苯乙酯之三階層三變數Box-Behnken實驗設計及其莫耳轉換率91 表5.3連續式酵素填充床生物反應器合成咖啡酸苯乙酯莫耳轉換率對合成變數之變異分析92 表5.4連續式酵素填充床生物反應器合成咖啡酸苯乙酯變數之聯合檢測分析94 表5.5脊型分析評估使用連續式酵素填充床生物反應器合成咖啡酸苯乙酯之莫耳轉換率最大值103

參考文獻

1. 陳國誠。2000。生物固定化技術與產業應用。第121-155頁。茂昌圖書有限公司。台北，台灣。
2. 高靄君。1992。反應曲面法在食品開發上的應用。食品工業月刊24(3): 32-41。
3. Adamczak, M. and Krishna, S. H. 2004. Strategies for improving enzymes for efficient biocatalysis. Food Technol. Biotechnol. 42: 251-264.
4. Aga, H., Shibuya, T., Sugimoto, T., Kurimoto, M. and Nakajima, S. 1994. Isolation and identification of antimicrobial compounds in Brazilian propolis. Biosci. Biotechnol. Biochem. 58: 945-946.
5. Babicz, I., Leite, S. G. F., Souza, R. O. M. A. D. and Antunes, O. A. C. 2010. Lipase-catalyzed diacylglycerol under sonochemical irradiation. Ultrason. Sonochem. 17: 4-6.
6. Bai, S., Guo, Z., Liu, W. and Sun, Y. 2006. Resolution of (\pm)-menthol by immobilized *Candida rugosa* lipase on superparamagnetic nanoparticles. Food Chem. 96: 1-7.
7. Balachandran, S., Kentish, S. E., Mawson, R. and Ashokkumar, M. 2006. Ultrasonic enhancement of the supercritical extraction from ginger. Ultrason. Sonochem. 13: 471-479.
8. Balcao, V. M., Paiva, A. L. and Malcata, F. X. 1996. Bioreactors with immobilized lipases: state of the art. Enzyme Microb. Technol. 18: 392-416.
9. Balcao, V. M. and Malcata, F. X. 1998. Lipase catalyzed modification of milk fat. Biotechnol. Adv. 16: 309-341.
10. Bankova, V., Popov, S. and Marekov, N. 1983. A study of flavonoids of propolis. J. Nat. Prod. 46: 471-474.
11. Banskota, A. H., Nagaoka, T., Sumioka, Y. L., Tezuka, Y., Awale, S., Midorikawa, K., Matsushige, K. and Kadota, S. 2002. Antiproliferative activity of the Netherlands propolis and its active principles in cancer cell lines. J. Ethnopharmacol. 80: 67-73.
12. Banskota, A. H., Tezuka, Y. and Kadota, S. 2001. Recent progress in pharmacological research of propolis. Phytother. Res. 15: 561-571.
13. Borrelli, F., Izzo, A. A., Di Carlo, G., Maffia, P., Russo, A., Maiello, F. M., Capasso, F. and Mascolo, N. 2002. Effect of a propolis extract and caffeic acid phenethyl ester on formation of aberrant crypt foci and tumors in the rat colon. Fitoterapia. 73: 38-43.
14. Borrelli, F., Maffia P., Pinto, L., Ianaro, A., Russo, A., Capasso, F. and Ialenti, A.

2002. Phytochemical compounds involved in the anti-inflammatory effect of propolis extract. *Fitoterapia*. 73: 53-63. 15. Buisman, G. J. H., Helteren, C. V., Kramer, G. F. H., Veldsink, J. W., Derksen, J. T. P. and Cuperus, F. P. 1998. Enzymatic esterifications of functionalized phenols for the synthesis of lipophilic antioxidants. *Biotechnol. Lett.* 20: 131-136. 16. Burdock, G. A. 1998. Review of the biological properties and toxicity of bee propolis (propolis). *Food Chem. Toxicol.* 36: 347-363. 17. Castaldo, S. and Capasso, F. 2002. Propolis, an old remedy used in modern medicine. *Fitoterapia*. 73: 1-6. 18. Chang, A. C. and Chen, F. C. 2002. The application of 20 kHz ultrasonic waves to accelerate the aging of different wines. *Food Chem.* 79: 501-506. 19. Chang, C., Chen, J. H., Chang, C. M. J., Wu, T. T. and Shieh, C. J. 2009. Optimization of lipase-catalyzed biodiesel by isopropanolysis in a continuous packed-bed reactor using response surface methodology. *New Biotech.* 26: 187-192. 20. Chang, S. W., Shaw, J. F., Yang, C. K. and Shieh, C. J. 2007. Optimal continuous biosynthesis of hexyl laurate by packed bed bioreactor. *Process Biochem.* 42: 1362-1366. 21. Chen, J. H., Shao, Y., Huang, M. T., Chin, C. K. and Ho, C. T. 1996. Inhibitory effect of caffeic acid phenethyl ester on human leukemic HL-60 cells. *Cancer Lett.* 108: 211-214. 22. Chen, J. W. and Wu, W. T. 2003. Regeneration of immobilized *Candida antarctica* lipase for transesterification. *J. Biosci. Bioeng.* 95: 466-469. 23. Chen, Q. H., Fu, M. L., Jin, L., Zhang, H. F., He, G. Q. and Ruan, H. 2009. Optimization of ultrasonic-assisted extraction (UAE) of betulin from white birch bark using response surface methodology. *Ultrason. Sonochem.* 16: 599-604. 24. Chen, Q. Y., Shi, H. and Ho, C. T. 1992. Effects of rosemary extracts and major constituents on lipid oxidation and soybean lipoxygenase activity. *J. Am. Oil Chem.* 69: 999-1002. 25. Chen, W. K., Tsai, C. F., Liao, P. H., Kuo, S. C. and Lee, Y. J. 1999. Synthesis of caffeic acid esters as antioxidant by esterification via acyl chlorides. *Chin. Pharm. J.* 51: 271-278. 26. Chisti, Y. 2003. Sonobioreactors: using ultrasound for enhanced microbial productivity. *Trends Biotechnol.* 21: 89-93. 27. Cirasino, L., Pisati, A. and Fasani, F. 1987. Contact dermatitis from propolis. *Contact Dermatitis.* 16:110-111. 28. Compton, D. L., Laszlo, J. A. and Berhow, M. A. 2000. Lipase-catalyzed synthesis of ferulate esters. *J. Am. Oil Chem. Soc.* 77: 513-519. 29. Cuvelier, M. E., Richard, H. and Berset, C. 1992. Comparison of the antioxidant activity of some acid-phenols: structure-activity relationship. *Biosci. Biotechnol. Biochem.* 56: 324-327. 30. De Castro, S. L. 2001. Propolis: biological and pharmacological activities. Therapeutic uses of this bee-product. *Annu. Rev. Boimed. Sci.* 3: 49-83. 31. Demirel, D. and Mutlu, M. 2005. Performance of immobilized pectinex ultra SP-L on magnetic duolite-polystyrene composite particles. Part II: A magnetic fluidized bed reactor. *J. Food Eng.* 70: 1-6. 32. Figueroa-Espinoza, M. C. and Villeneuve, P. 2005. Phenolic acid enzymatic lipophilization. *J. Agric. Food Chem.* 53: 2779-2787. 33. Freitas, S., Hielscher, G., Merkle, H. P. and Gander, B. 2006. Continuous contact and contamination free ultrasonic emulsification- A useful tool for pharmaceutical development and production. *Ultrason. Sonochem.* 13: 76-85. 34. Gandhi, N. N. 1997. Applications of lipase. *J. Am. Oil Chem. Soc.* 74: 621-634. 35. Garcia, R., Renedo, A., Martinez, M. and Aracil, J. 2002. Enzymatic synthesis of n-octyl (\pm)-2-methylbutanoate ester from racemic (\pm)-2-methylbutanoic acid by immobilized lipase: optimization by statistical analysis. *Enzyme Microb. Technol.* 30: 110-115. 36. Gomez-Romero, M., Arraez-Roman, D., Moreno-Torres, R., Garcia-Salas, P., Segura-Carretero, A. and Fernandez-Gutierrez, A. 2007. Antioxidant compounds of propolis determined by capillary electrophoresis – mass spectrometry. *J. Sep. Sci.* 30: 595-603. 37. Graf, E. 1992. Antioxidant potential of ferulic acid. *Free Radic. Biol. Med.* 13: 435-448. 38. Grunberger, D., Banerjee, R., Eisinger, K., Oltz, E. M., Efros, L., Caldwell, M., Estevez, V. and Nakanishi, V. 1988. Preferential cytotoxicity on tumor cells by caffeic acid phenethyl ester isolated from propolis. *Cell. Mol. Life Sci.* 44: 230-232. 39. Gupta, M. N. and Roy, I. 2004. Enzyme in organic media: forms, function and applications. *Eur. J. Biochem.* 271: 2575-2583. 40. Guyou, B., Bosquette, B., Pina, M. and Graille, J. 1997. Esterification of phenolic acids from green coffee with an immobilized lipase from *Candida antarctica* in solvent-free medium. *Biotechnol. Lett.* 19: 529-532. 41. Guyou, B., Gueule, D., Pina, M., Graille, J., Farines, V. and Farines, M. 2000. Enzymatic synthesis of fatty ester in 5-caffeoyl quinic acid. *Eur. J. Lipid Sci. Technol.* 102: 93-96. 42. Halim, S. F. A., Kamaruddin, A. H. and Fernando, W. J. N. 2009. Continuous biosynthesis of biodiesel from waste cooking palm oil in a packed bed reactor: Optimization using response surface methodology (RSM) and mass transfer studies. *Bioresour. Technol.* 100: 710-716. 43. Halldorsson, A., Magnusson, C. D. and Haraldsson, G. G. 2003. Chemoenzymatic synthesis of structured triacylglycerols by highly regioselective acylation. *Tetrahedron* 59: 9101-9109. 44. Hasan, F., Shah, A. A. and Hameed, A. 2006. Industrial application of microbial lipase. *Enzyme Microb. Technol.* 39: 235-251. 45. Heo, M. Y., Sohn, S. J. and Au, W. W. 2001. Anti-genotoxicity of galangin as a cancer chemopreventive agent candidate. *Mutat. Res.* 488: 135-150. 46. Hernandez, C. E., Chen, H. H., Chang, C. I. and Huang, T. C. 2009. Direct lipase-catalyzed lipophilization of chlorogenic acid from coffee pulp in supercritical carbon dioxide. *Ind. Crop. Prod.* 30: 359-365. 47. Hishikawa, K., Nakaki, T. and Fujita, T. 2005. Oral flavonoid supplementation attenuates atherosclerosis development in apolipoprotein E-deficient mice. *Arterioscler. Thromb. Vasc. Biol.* 25: 442-446. 48. H-Kittikun, A., Kaewthong, W. and Cheirsilp, B. 2008. Continuous production of monoacylglycerols from palm olein in packed-bed reactor with immobilized lipase PS. *Biochem. Eng. J.* 40: 116-120. 49. Hollman, P. C. H. 2001. Evidence for health benefits of plant phenols: local or systemic effects? *J. Sci. Food Agr.* 81: 842-852. 50. Horchani, H., Salem, N. B., Zarai, Z., Sayari, A., Gargouri, Y. and Chaabouni, M. 2010. Enzymatic synthesis of eugenol benzoate by immobilized *Staphylococcus aureus* lipase: optimization using response surface methodology and determination of antioxidant activity. *Bioresour. Technol.* 101: 2809-2817. 51. Hsu, A. F., Jones, K. C., Foglia, T. A. and Marmer, W. N. 2004. Continuous production of ethyl esters of grease using an immobilized lipase. *J. Am. Oil Chem. Soc.* 81: 749-752. 52. Ilhan, A., Iraz, M., Gurel, A., Armutcu, F. and Kyol, O. 2004. Caffeic acid phenethyl ester exerts a neuroprotective effect on CNS against pentylentetrazol-induced seizures in mice. *Neurochem. Res.* 29: 2287-2292. 53. Jayaprakasam, B., Vanisree, M., Zhang, Y., Dewitt, D. L. and Nair, M. G. 2006. Impact of alkyl esters of caffeic and ferulic acids on tumor cell proliferation, cyclooxygenase enzyme, and lipid peroxidation. *J. Agric. Food Chem.* 54: 5376-5381. 54. Jennings, B. H. and Akoh, C. C. 2001. Lipase catalyzed modification of fish oil to incorporate capric acid. *Food Chem.* 72: 273-278. 55. Ji, J., Wang, J., Li, Y., Yu, Y. and Xu, Z. 2006. Preparation of biodiesel with the help of ultrasonic and

hydrodynamic cavitation. *Ultrasonics* 44: 411-414. 56. Ju, H. Y., Yang, C. K., Yen, Y. H. and Shieh, C. J. 2009. Continuous lipase-catalyzed synthesis of hexyl laurate in a packed-bed reactor: optimization of the reaction conditions in a solvent-free system. *J. Chem. Biotechnol.* 84: 29-33. 57. Jung, W. K., Lee, D. Y., Kim, J. H., Choi, I., Park, S. G., Seo, S. K., Lee, S. W., Lee, C. M., Park, Y. M., Jeon, Y. J., Lee, C. H., Jeon, B. T., Qian, Z. J., Kim, S. K. and Choi, I. W. 2008. Anti-inflammatory activity of caffeic acid phenethyl ester (CAPE) extracted from *Rhodiola sacra* against lipopolysaccharide-induced inflammatory responses in mice. *Process Biochem.* 43: 783-787. 58. Kajiyama, T. and Ohkatsu, Y. 2001. Effect of para-substituents of phenolic antioxidants. *Polym. Degrad. Stabil.* 71: 445-452. 59. Karam, R., Karboune, S., St-Louis, R. and Kermasha, S. 2009. Lipase-catalyzed acidolysis of fish liver oil with dihydroxyphenylacetic acid in organic solvent media. *Process Biochem.* 44: 1193-1199. 60. Karboune, S., Safari, M. Lue, B. M., Yeboah, F. K. and Kermasha, S. 2005. Lipase-catalyzed biosynthesis of cinnamoylated lipids in a selected organic solvent medium. *J. Biotechnol.* 119: 281-290. 61. Karboune, S., St-Louie, R. and Kermasha, S. 2008. Enzymatic synthesis of structured phenolic lipids by acidolysis of flaxseed oil with selected phenolic acids. *J. Mol. Catal. B-Enzym.* 52-53: 96-105. 62. Kimbaris, A. C., Siatis, N. G., Daferera, D. J., Tarantilis, P. A., Pappas, C. S. and Polissiou, M. G. 2006. Comparison of distillation and ultrasound-assisted extraction methods for the isolation of sensitive aroma compounds from garlic (*Allium sativum*). *Ultrason. Sonochem.* 13: 54-60. 63. Kirk, O., Borchert, T. V. and Fuglsang, C. C. 2002. Industrial enzyme applications. *Curr. Opin. Biotechnol.* 13: 345-351. 64. Kristensen, J. B., Xu, X. and Mu, H. 2005. Diacylglycerol synthesis by enzymatic glycerolysis: screening of commercially available lipases. *J. Am. Oil Chem. Soc.* 82: 329-334. 65. Laszlo, J. A., Compton, D. L., Eller, F. J., Taylor, S. L. and Isbell, T. A. 2003. Packed-bed bioreactor synthesis of feruloylated monoacyl- and diacyl-glycerols: clean production of a "green" sunscreen. *Green Chem.* 5: 382-386. 66. Lee, G. S., Widjaja, A. and Ju, Y. H. 2006. Enzymatic synthesis of cinnamic acid derivatives. *Biotechnol. Lett.* 28: 581-585. 67. Lee, S. H., Nguyen, H. M., Koo, Y. M. and Ha, S. H. 2008. Ultrasound-enhanced lipase activity in the synthesis of sugar ester using ionic liquids. *Process Biochem.* 43: 1009-1012. 68. Lee, K. T., Akoh, C. C. and Dawe, D. L. 1999. Effects of structured lipid containing omega-3 and medium chain fatty acids on serum lipids and immunological variables in mice. *J. Food Biochem.* 23: 197-208. 69. Lee, Y. J., Liao, P. H., Chen, W. K. and Yang, C. Y. 2000. Preferential cytotoxicity of caffeic acid phenethyl ester analogues on oral cancer cells. *Cancer Lett.* 153: 51-56. 70. Liu, L., Robert Hudgins, W., Shack, S., Yin, M. Q. and Samid, D. 1995. Cinnamic acid: A natural product with potential use in cancer intervention. *Int. J. Cancer.* 62: 345-350. 71. Liu, X. Y., Guo, F. L., Wu, L. M. and Liu, Z. L. 1996. Remarkable enhancement of antioxidant activity of vitamin C in an artificial bilayer by marking it lipo-soluble. *Chem. Phys. Lipids* 62: 345-350. 72. Liu, Y., Jin, Q., Shan, L., Liu, Y., Shen, W. and Wang, X. 2008. The effect of ultrasound on lipase-catalyzed hydrolysis of soy oil in solvent-free system. *Ultrason. Sonochem.* 15: 402-407. 73. Long, W. S., Kamaruddin, A. and Bhatia, S. 2005. Chiral resolution of racemic ibuprofen ester in an enzymatic membrane reactor. *J. Membr. Sci.* 247: 185-200. 74. Lopez Giraldo, L. J., Laguerre, M., Lecomte, J., Figueroa-Espinoza, M. C., Barouh, N., Barea, B. and Villeneuve, P. 2007. Lipase-catalyzed synthesis of chlorogenate fatty esters in solvent-free medium. *Enzyme Microb. Technol.* 41: 721-726. 75. Malcata, F. X., Reyes, H. R., Garcia, H. S., Hill Jr., C. G. and Amundson, C. H. 1990. Immobilized lipase reactors for modification of fats and oils. A review. *J. Am. Oil Chem. Soc.* 67: 890-910. 76. Margulis, M. A. 1992. Fundamental aspects of sonochemistry. *Ultrasonics* 30: 152-154. 77. Marinova, E. M. and Yanishlieva, N. V. 1994. Effect of lipid unsaturation in the antioxidative activity of some phenolic acid. *J. Am. Oil Chem. Soc.* 71: 427-434. 78. Markham, K. R., Mitchell, K. A., Wilkins, A. L., Daldy, J. A. and Lu, Y. 1996. HPLC and GC-MS identification of the major organic constituents in NEW Zealand propolis. *Phytochemistry* 42: 205-211. 79. Masudo, T. and Okada, T. 2001. Ultrasonic Irradiation-Novel principle for microparticle separation. *Anal. Sci.* 17: 341-344. 80. Michaluart, P., Masferrer, J. L., Carothers, A. M., Subbaramaiah, K., Zweifel, B. S., Koboldt, C., Mestre, J. R., Grunberger, D., Sacks, P. G., Tanabe, T. and Dannenberg, A. J. 1999. Inhibitory effects of caffeic acid phenethyl ester on the activity and expression of cyclooxygenase-2 in human oral epithelial cells and in rat model of inflammatory. *Cancer Res.* 59: 2347-2352. 81. Miethchen, R. 1992. Selected applications of sonochemistry in organic chemistry. *Ultrasonics* 30: 173-179. 82. Mirzoeva, O. K. and Calder, P. C. 1996. The effect of propolis and its components on eicosanoid production during the inflammatory response. *Prostaglandins Leukot. Essent. Fatty Acids* 55: 441-449. 83. Nagaoka, T., Banskota, A. H., Tezuka, Y., Saiki, I. and Kadota, S. 2002. Selective antiproliferative activity of caffeic acid phenethyl ester analogues on highly liver-metastatic murine colon 26-L5 carcinoma cell line. *Bioor. Med. Chem.* 10: 3351-3359. 84. Natella, F., Nardini, M., Felice, M. D. and Scaccini, C. 1999. Benzoic and cinnamic acid derivatives as antioxidant: structure-activity relation. *J. Agric. Food Chem.* 47: 1453-1459. 85. Nielsen, N. S., Yang, T., Xu, X. and Jacobsen, C. 2006. Production and oxidative stability of a human milk fat substitute produced from lard by enzyme technology in a pilot packed-bed reactor. *Food Chem.* 94: 53-60. 86. Noelker, C., Bacher, M., Gocke, P., Weib, X., Klockgether, T., Du, Y. and Dodel, R. 2005. The α -avanoid caffeic acid phenethyl ester blocks 6-hydroxydopamine-induced neurotoxicity. *Neurosci. Lett.* 383: 39-43. 87. Papay, V., Toth, L., Soltész, M., Nagy, E. and Litkei, G. 1986. Isolated compounds from Hungarian propolis and populi gemma. *Stud. Org. Chem.* 23: 233-240. 88. Patist, A. and Bates, D. 2008. Ultrasonic innovations in the food industry: From the laboratory to commercial production. *Innov. Food Sci. Emerg. Technol.* 9: 147-154. 89. Pitt, W. G. and Rodd, A. 2003. Ultrasound increases the rate of bacterial growth. *Biotechnol. Prog.* 19: 1030-1044. 90. Posorske, L. H. 1984. Industrial-scale application of enzymes to the fats and oil industry. *J. Am. Oil Chem. Soc.* 61: 1758-1760. 91. Puri, S., Kaur, B., Parmar, A. and Kumar, H. 2009. Ultrasound-promoted greener synthesis of 2H-chromen-2-ones catalyzed by copper perchlorate in solventless media. *Ultrason. Sonochem.* 16: 705-707. 92. Piao, J., Kobayashi, T., Adachi, S., Nakanshi, K. and Matsuno, R. 2004. Continuous synthesis of lauroyl and oleoyl erythritol by a packed bed reactor with an immobilized lipase. *Process Biochem.* 39: 113-119. 93. Ribeiro, C. M. R., Passaroto, E. N. and Brenelli, E. C. S. 2001. Ultrasound in enzymatic resolution of ethyl 3-hydroxy-3-phenylpropanoate. *Tetrahedron Lett.* 42: 6477-6479. 94. Ramachandran, K. B., Al-Zuhair, S., Fong, C. S. and Gak, C. W. 2006. Kinetic study on hydrolysis of oils by lipase with ultrasonic

emulsification. *Biochem. Eng. J.* 32: 19-24. 95. Ratoarinoro, Contamine, F., Wilhelm, A. M., Berlan, J. and Delmas, H. 1995. Activation of a solid-liquid chemical reaction by ultrasound. *Chem. Eng. Sci.* 50: 554-558. 96. Royon, D., Daz, M., Ellenrieder, G. and Locatelli, S. 2007. Enzymatic production of biodiesel from cotton seed oil using t-butanol as a solvent. *Bioresour. Technol.* 98: 648-653. 97. Sabally, K., Karboune, S., St-Louis, R. and Kermasha, S. 2006. Lipase-catalyzed transesterification of trilinolein or trilinolenin with selected phenolic acids. *J. Am. Oil Chem. Soc.* 83: 101-107. 98. Shahidi, F. and Ho, C. T. 2005. Phenolic compounds in foods and natural health products. p. 2-5. American Chemical Society, Washington, DC, USA. 99. Stamatis, H., Sereti, V. and Kolisis, F. N. 2001. Enzymatic synthesis of hydrophilic and hydrophobic derivatives of natural phenolic acids in organic media. *J. Mol. Catal. B-Enzym.* 11: 323-328. 100. Stavarache, C., Vinatoru, M., Nishimura, R. and Maeda, Y. 2005. Fatty acids methyl esters from vegetable oil by means of ultrasonic energy. *Ultrason. Sonochem.* 12: 367-372. 101. Stepanovi?, S., Anti?, N., Daki?, I. and Vlahovi?, M. 2003. In vitro antimicrobial activity of propolis and synergism between propolis and antimicrobial drugs. *Microbiol. Res.* 158: 353-357. 102. Stevenson, D. E., Parkar, S. G., Zhang, J., Stanley, R. A., Jensen, D. J. and Cooney, J. M. 2007. Combinatorial enzymic synthesis for functional testing of phenolic acid esters catalysed by *Candida antarctica* lipase B (NovozymR 435). *Enzyme Microb. Technol.* 40: 1078-1086. 103. Sud'ina, G. F., Mirzoeva, O. K., Pushkareva, M. A., Korshunova, G. A., Sumbatyan, N. V. and Varfolomeev, S. D. 1993. Caffeic acid phenethyl ester as a lipoxygenase inhibitor with antioxidant properties. *FEBS Lett.* 329: 21-24. 104. Talukder, M. M. R., Zaman, M. M., Hayashi, Y., Wui, J. C. and Kawanishi, T. 2006. Ultrasonication enhanced hydrolytic activity of lipase in water/isooctane two-phase systems. *Biocatal. Biotransfor.* 24: 189-194. 105. Twu, Y. K., Shih, I. L., Yen, Y. H., Ling, Y. F. and Shieh, C. J. 2005. Optimization of lipase-catalyzed synthesis of octyl hydroxyphenylpropionate by response surface methodology. *J. Agric. Food Chem.* 53: 1012-1016. 106. Vilku, K., Mawson, R., Simons, L. and Bates, D. 2008. Applications and opportunities for ultrasound assisted extraction in the food industry-A review. *Innov. Food Sci. Emerg. Technol.* 9: 161-169. 107. Vosmann, K., Wiege, B., Weitkamp, P. and Weber, N. 2008. Preparation of lipophilic alkyl (hydroxy) benzoates by solvent-free lipase-catalyzed esterification and transesterification. *Appl. Microbiol. Biotechnol.* 80: 29-36. 108. Watanabe, Y., Shimada, Y., Sugihara, A. and Tominaga, Y. 2001. Enzymatic conversion of waste edible oil to biodiesel fuel in a fixed-bed bioreactor. *J. Am. Oil Chem. Soc.* 78: 703-707. 109. Wei, X., Zhao, L., Ma, Z., Holtzman, D. M., Yan, C., Dodel, R. C., Hampel, H., Oertel, W., Farlow, M. R. and Du, Y. 2004. Caffeic acid phenethyl ester prevents neonatal hypoxicischaemic brain injury. *Brain* 127: 2629-2635. 110. Weitkamp, P., Vosmann, K. and Weber, N. 2006. Highly efficient preparation of lipophilic hydroxycinnamates by solvent-free lipase-catalyzed transesterification. *J. Agric. Food Chem.* 54: 7062-7068. 111. Wu, W. M., Lu, L., Long, Y., Wang, T., Liu, L., Chen, Q. and Wang, R. 2007. Free radical scavenging and antioxidative activities of caffeic acid phenethyl ester (CAPE) and its related compounds in solution and membranes: A structure – activity insight. *Food Chem.* 105: 107-115. 112. Xiao, Y. M., Wu, Q., Cai, Y. and Lin, X. F. 2005. Ultrasound-accelerated enzymatic synthesis of sugar esters in nonaqueous solvents. *Carbohydr. Res.* 340: 2097-2103. 113. Xin, J. Y., Zhang, L., Chen, L. I., Zhang, Y., Wu, X. M. and Xia, C. G. 2009. Lipase-catalyzed synthesis of ferulyl oleins in solvent-free medium. *Food Chem.* 112: 640-645. 114. Xu, X. 2000. Production of specific-structured triacylglycerols by lipase-catalyzed reactions: a review. *Eur. J. Lipid Sci. Tech.* 102: 287-303. 115. Yadav, G. D. and Dhoot, S. B. 2009. Immobilized lipase-catalyzed synthesis of cinnamyl laurate in non-aqueous media. *J. Mol. Catal. B-Enzym.* 57: 34-39. 116. Yang, T., Rebsdore, M., Engelrud, U. and Xu, X. 2005. Enzymatic production of monoacylglycerols containing polyunsaturated fatty acid through an efficient glycerolysis system. *J. Agric. Food Chem.* 53: 1475-1481. 117. Yu, D., Tian, L., Wu, H., Wang, S., Wang, Y., Ma, D. and Fang, X. 2010. Ultrasonic irradiation with vibration for biodiesel production from soybean oil by NovozymR 435. *Process Biochem.* 45: 519-525. 118. Yu, X., Li, Y. and Wu, D. 2004. Enzymatic synthesis of gallic acid esters using microencapsulated tannase: effect of organic solvents and enzyme specificity. *J. Mol. Catal. B-Enzym.* 30: 69-73. 119. Yu, Z. R., Chang, S. W., Wang, H. Y. and Shieh, C. J. 2003. Study on synthesis parameters of lipase-catalyzed hexyl acetate in supercritical CO₂ by response surface methodology. *J. Am. Oil Chem. Soc.* 80: 139-144. 120. Zhang, J., Chen, M., Ju, W., Liu, S., Xu, M., Chu, J. and Wu, T. 2010. Liquid chromatograph/tandem mass spectrometry assay for the simultaneous determination of chlorogenic acid and cinnamic acid in plasma and its application to a pharmacokinetic study. *J. Pharm. Biomed. Anal.* 51: 685-690. 121. Zhang, Y., Stanculescu, M. and Ikura, M. 2009. Rapid transesterification of soybean oil with phase transfer catalysts. *Appl. Catal. A-Gen.* 366: 176-183. 122. Zheng, L. and Sun, D. W. 2006. Innovative applications of power ultrasound during food freezing processes-A review. *Trends Food Sci. Technol.* 17: 16-23.